Product Information

**Anti-Nitric Oxide Synthase, Brain (1409-1429)**
produced in rabbit, IgG fraction of antiserum

*Catalog Number N7280*

**Synonym:** Anti-bNOS

**Product Description**

Anti-Nitric Oxide Synthase, Brain (1409-1429) is produced in rabbit using as immunogen the synthetic peptide K-RSESIAFIEESKKDADEVFSS corresponding to the C-terminal fragment of bNOS of rat brain origin (amino acids 1409-1429 with N-terminally added lysine) conjugated to KLH. This sequence differs from the corresponding human sequence by a single amino acid. Whole antiserum is purified to provide an IgG fraction of antiserum.

Anti-Nitric Oxide Synthase, Brain (1409-1429) is specific for nitric oxide synthase (NOS) derived from brain (150-160 kDa), but does not recognize NOS derived from activated macrophages (iNOS) and endothelial cells (eNOS). By immunoblotting of rat brain tissue extract, the staining of bNOS band is specifically inhibited with the immunizing peptide.

Nitric Oxide Synthases are enzymes involved in the synthesis of nitric oxide (NO), a messenger molecule generated under physiological conditions by virtually all mammalian cells.\(^1,2,3\) NO is formed from oxidation of L-arginine by NOS, releasing NO and L-citrulline. NO mediates diverse functions including vasodilation, synaptic neurotransmission, and cytotoxicity. NO participates in signal transduction pathways by activating soluble guanylate cyclase. In addition, NO has been implicated as a pathogenic mediator in a variety of conditions, such as central nervous system (CNS) disease states, including focal cerebral ischemia, and the animal model of multiple sclerosis (MS), experimental allergic encephalomyelitis.\(^4\)

At least three distinct isoforms of NOS have been described which are the products of different mammalian genes. The NOS isoform found in neurons is a 150-160 kDa protein. It is also termed brain NOS, \(nNOS\), neuronal NOS (nNOS), neuronal constitutive NOS or Ca\(^{2+}\) -regulated NOS (cNOS, ncNOS). The NOS isoform characterized in macrophages is a 130 kDa protein, also known as macrophage NOS (mNOS), NOS2 or inducible NOS (iNOS). The NOS isoform found in endothelial cells is a 135 kDa protein, also called endothelial NOS, NOS 3 (eNOS, or ecNOS). nNOS and eNOS are constitutively expressed and are dependent on Ca\(^{2+}\) and calmodulin for NO production. iNOS is Ca\(^{2+}\) independent and is expressed in cytokine-activated macrophages and microglial cells in response to bacterial lipopolysaccharide (LPS) stimulation.

NOS are complex enzymes forming homodimers under native conditions, and require three cosubstrates, L-arginine, NADPH, and O\(_2\), and five cofactors FAD, FMN, tetrahydrobiopterin (BH\(_4\)), heme, and calmodulin. The C-terminal half of NOS possesses a high level of homology with NADPH-cytochrome P-450 reductase, where the predicted sites for binding NADPH and flavins are also located. However, the predicted heme and calmodulin binding sites of NOS are located within its N-terminal half.

NOS isoforms may subserve a multiplicity of disparate biological functions.\(^1,5,6\) For instance, bNOS is present also in skeletal muscle, where it is complexed with dystrophin and is absent in Duchenne’s muscular dystrophy (DMD).\(^6\) iNOS not only occurs in macrophages but in several other cell types including hepatocytes, chondrocytes, endothelial cells, and fibroblasts. eNOS is not restricted to the endothelium of blood vessels but exists in the epithelium of several tissues, including the bronchial tree. It is also localized in neurons in the brain, especially the pyramidal cells of the hippocampus, where it may function in long-term potentiation. NOS seems to be a highly conserved enzyme between the various types. Human bNOS and eNOS share 52% amino acid identity, and rat and human bNOS share 93% amino acid identity.

The production of isoform specific antibodies to NOS allows investigators to identify which isoform(s) is present in a specific cell or tissue. These antibodies are valuable for elucidating the expression of these isoforms in a variety cell types and tissues.
Reagent
Solution in 0.01 M phosphate buffered saline, pH 7.4, containing 15 mM sodium azide as a preservative.

Precautions and Disclaimer
This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability
Store at 2-8 °C for up to one month. For extended storage, solution may be frozen in working aliquots. Repeated freezing and thawing, or storage in "frost-free" freezers, is not recommended. If slight turbidity occurs upon prolonged storage, clarify by centrifugation before use.

Procedure
Immunoblotting Procedure of Whole Cell Extract and Rat Brain Extract

Reagents and Equipment
1. Bovine endothelial lung cells culture (eNOS control).
2. Rats (Sprague-Dawley).
3. Macrophage cell line RAW264.7 (mouse), activated with interferon-γ (IFN-γ) and lipopolysaccharide (LPS), (iNOS control).
4. Lipopolysaccharide (LPS), from E. coli, Catalog No. L2654.
6. Phenylmethylsulfonylfluoride (PMSF) Catalog No. P7626, 0.5M in EtOH.
7. Pepstatin A, Catalog No. P4265, 2 mg/ml in DMSO.
8. Leupeptin, Catalog No. L2884.
10. DTT, Catalog No. D9760.
11. Homogenization Buffer (Rat brain), Ice Cold: 20 mM Tris-HCl buffer, pH 7.5, containing 0.25 M sucrose, 10 mM EGTA, 2 mM EDTA, and protease inhibitors: 2 mM PMSF, 50 µg/ml leupeptin, 25 µg/ml aprotinin, 10 µg/ml pepstatin A and 2 mM dithiothreitol (DTT).

Note: Add all protease inhibitors freshly before tissue homogenization.

12. Mechanical homogenizer.
13. Refrigerated High-Speed Centrifuge.
14. Protein assay kit .
15. Laemmli sample (3X) buffer containing 2-mercaptopethanol.
16. Laemmli sample (1X) buffer containing 2-mercaptopethanol.

Procedure
Preparation of Whole Cell Culture Extract

1. Grow bovine endothelial cells to confluence in 10 cm plate containing 10% FCS in DMEM.
2. Grow RAW 264.7 macrophage cells to confluence in 10 cm plate containing 10% FCS in DMEM.
3. Activate RAW 264.7 cells by incubation with LPS (1 µg/ml) and IFN-γ (10 ng/ml) overnight at 37 °C.
4. Remove medium from culture dishes.
5. Rinse plates with ice PBS pH 7.4 (2 x 10 ml).
6. Add 1 ml/plate of (1x) boiling sample buffer and scrape cells.
7. Boil sample for 5 min. at 95 °C.
8. Aliquot samples of cells extracts and store at −70 °C.

Preparation of Rat Brain Extract

Note: All procedure steps are carried out on ice, unless noted otherwise.

Rapidly dissect out whole brains (5 g) from Sprague-Dawley rats (4-5 rats, 250-300 g) and collect into ice cold homogenization buffer.

1. Homogenize tissue in 5x volumes of ice cold homogenization buffer (w/v) using mechanical homogenizer at maximum speed (3 x 10 sec pulses with 1 min rest in between).
2. Homogenize briefly on ice with fine glass homogenizer.
3. Centrifuge homogenate at 1200 x g for 10 min at 4 °C. Collect supernatant.
4. Centrifuge supernatant at 15,000 x g for 20 min at 2-8 °C. Collect clear supernatant.
5. Remove 0.5 ml aliquot of supernatant for protein determination using the Bradford method with BSA as standard.
6. Add to supernatant 3x sample buffer to final dilution of 1x sample buffer.
7. Boil sample for 5 min. at 95 °C.
8. Aliquot sample of rat brain extract and store at −70 °C.
Immunoblotting Reagents and Equipment

1. Bovine endothelial cell extract (eNOS control).
2. Rat brain extract.
3. Activated-RAW 264.7 macrophage cells extract (iNOS control).
4. 7% polyacrylamide slab minigel with 5% stacking gel (80 x 80 x 1.5 mm).
5. Nitrocellulose membrane (0.45 mm).
7. Blocking Buffer: 10% dry milk (w/v) in 10 mM phosphate buffered saline (PBS), pH 7.4.
8. Dilution Buffer: 1% BSA in PBS pH 7.4 containing 0.05% TWEEN® 20.
9. Washing Buffer: PBS pH 7.4 containing 0.05% Tween-20.
10. NOS brain peptide (amino acids 1409-1429 with N-terminally added lysine). Dissolve in double distilled water at 0.5 mg/ml. Store aliquots at –20 °C.
11. Primary antibody: Anti-bNOS (1409-1429), Catalog No. N7280, at appropriate dilution in dilution buffer.
14. Electrophoresis and transfer apparatus.

Immunoblotting Procedure

Note: In order to obtain best results in different preparations it is recommended to optimize procedure conditions (antibody dilutions, incubation times, blocking conditions etc.), for a specific application.

1. Resolve whole cell extracts (250 µL/slab) and rat brain extract (250 µg/slab), on precast 7% polyacrylamide minigel.
2. Run SDS-PAGE at room temperature.
3. Perform transfer for 1 hour at room temperature to nitrocellulose membrane.
4. Block nitrocellulose membrane in blocking buffer for at least 1 hour at room temperature.
5. Incubate membrane with primary antibody dilutions for 2 hours at room temperature.(a)
6. Wash membrane with washing buffer 4 x 5 min.
7. Incubate membrane with secondary antibody at recommended dilution in dilution buffer for 1 hour at room temperature.
8. Wash membrane with washing buffer 4 x 5 min. Wash 1 x 5 min. in deionized water.
10. Wash membrane thoroughly with deionized water.
11. Air-dry blots on filter paper.

(a)Note: For specific inhibition of NOS brain (bNOS) band (150-160 kDa band) it is recommended to incubate prediluted antibody with bNOS peptide (1409-1429), (concentration 10 µg/ml), for 2 hours at room temperature or overnight at 2-8 °C.

Product Profile
Immunoblotting: a minimum working dilution of 1:10,000 is determined by using a rat brain extract.

Note: In order to obtain best results, it is recommended that each individual user determine their optimum working dilutions by titration assay.

Total Protein is determined by absorbance at 280 nm (E$_{280}^{1%}$ = 14.0).

References