Product Information

**CDC7/DBF4, active, GST-tagged, human PRECISIO® Kinase recombinant, expressed in Sf9 cells**

Catalog Number **SRP5231**
Storage Temperature –70 °C

Synonyms: CDC7, CDC7L1, HsCDC7, Hsk1, huCDC7, MGC117361, MGC126237, MGC126238 DBF4, ASK, CHIF, DBF4A, ZDBF1

**Product Description**

CDC7 is a cell division cycle 7 homolog protein that is critical for the G1/S transition and is also essential for initiation of DNA replication as cell division occurs. CDC7 is expressed in many normal tissues, but the overexpression of CDC7 may be associated with neoplastic transformation for some tumors and transformed cell lines. CDC7/DBF4 kinase promotes S phase by alleviating an inhibitory activity in Mcm4 that evolved to integrate several protein kinases.

Recombinant full-length human CDC7 and DBF4 proteins were co-expressed by baculovirus in Sf9 insect cells using N-terminal GST tags. The CDC7 gene accession number is NM_003503; DBF4 is NM_006716. Recombinant protein stored in 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM glutathione, 0.1 mM EDTA, 0.25 mM DTT, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~94 kDa and 125 kDa

Purity: 70–95% (SDS-PAGE, see Figure 1)

Specific Activity: 7.2–9.8 nmole/min/mg (see Figure 2)

**Precautions and Disclaimer**

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

**Storage/Stability**

The product ships on dry ice and storage at –70 °C is recommended. After opening, aliquot into smaller quantities and store at –70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

**Procedure**

**Preparation Instructions**

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl2, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50ng/μl BSA solution.
Kinase Solution – Dilute the active CDC7/DBF4 (0.1 µg/µl) with Kinase Dilution Buffer to the desired concentration. 

*Note:* The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active CDC7/DBF4 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 µl aliquots at –20 °C.

γ-33P-ATP Assay Cocktail (250 µM) – Combine 5.75 ml of Kinase Assay Buffer, 150 µl of 10 mM ATP Stock Solution, 100 µl of γ-33P-ATP (1 mCi/100 µl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – PDKtide peptide substrate (KTFCGTPYLAPEYLAPEVRREPRILSEEEQEMFRD-FDYIADWC) diluted in distilled water to a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the 33P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

1. Thaw the active CDC7/DBF4, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ-33P-ATP Assay Cocktail may be thawed at room temperature.
2. In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 µl:
   10 µl of Kinase Solution
   5 µl of Substrate Solution
   5 µl of cold water (4 °C)
3. Set up a blank control as outlined in step 2, substituting 5 µl of cold water (4 °C) for the Substrate Solution.
4. Initiate each reaction with the addition of 5 µl of the γ-33P-ATP Assay Cocktail, bringing the final reaction volume to 25 µl. Incubate the mixture in a water bath at 30 °C for 15 minutes.
5. After the 15 minute incubation, stop the reaction by spotting 20 µl of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.
6. Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of –10 minutes each.
7. Set up a radioactive control to measure the total γ-33P-ATP counts introduced into the reaction. Spot 5 µl of the γ-33P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

**Calculations:**

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

\[ SR = \frac{cpm \text{ of } 5 \mu l \text{ of } \gamma-33P-ATP \text{ Assay Cocktail}}{nmole \text{ of ATP}} \]

   cpm – value from control (step 7)
   nmole – 1.25 nmole (5 µl of 250 µM ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

\[ \text{nmole/min/mg} = \frac{\Delta \text{cpm} \times (25/20)}{SR \times E \times T} \]

\[ \Delta \text{cpm} = \text{cpm of the sample – cpm of the blank (step 3)} \]
\[ 25 = \text{total reaction volume} \]
\[ 20 = \text{spot volume} \]
\[ T = \text{reaction time (minutes)} \]
\[ E = \text{amount of enzyme (mg)} \]

**References**


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