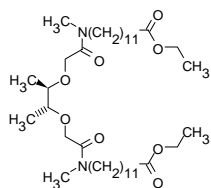


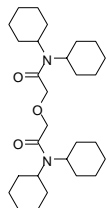
## Calcium



### Calcium ionophore I

(ETH 1001; (-)-(R,R)-N,N'-Bis-[11-(ethoxycarbonyl)undecyl]-N,N',4,5-tetramethyl-3,6-dioxaoctane-diamide; Diethyl N,N'-[(4R,5R)-4,5-dimethyl-1,8-dioxo-3,6-dioxaoctamethylene]bis(12-methylaminododecanoate))  
 $C_{38}H_{72}N_2O_8$   $M_r$  689.99 [58801-34-6]

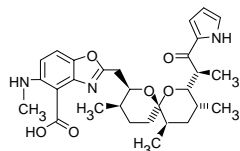
[21192](#) **Selectophore®**, function tested 50 mg, 250 mg



### Calcium ionophore II

(ETH 129; N,N,N',N'-Tetra[cyclohexyl]diglycolic acid diamide; N,N,N',N'-Tetracyclohexyl-3-oxapentanediamide)  
 $C_{28}H_{48}N_2O_3$   $M_r$  460.69 [74267-27-9]

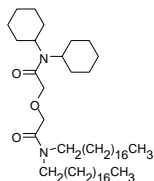
[21193](#) **Selectophore®**, function tested 50 mg, 250 mg



### Calcium ionophore III

(A 23187; Calcimycin)  
 $C_{29}H_{37}N_3O_6$   $M_r$  523.62 [52665-69-7]

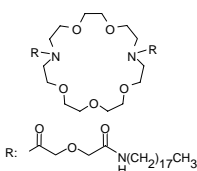
[21186](#) **Selectophore®**, function tested 5 mg, 25 mg



### Calcium ionophore IV

(ETH 5234; N,N-Dicyclohexyl-N',N'-dioctadecyl-diglycolic diamide; N,N-Dicyclohexyl-N',N'-dioctadecyl-3-oxapentanediamide)  
 $C_{52}H_{100}N_2O_3$   $M_r$  801.36 [126572-74-5]

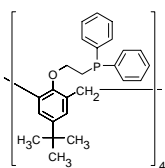
[21198](#) **Selectophore®**, function tested 50 mg, 250 mg



### Calcium ionophore V

(K23E1; 10,19-Bis[(octadecylcarbamoyl)methoxyacetyl]-1,4,7,13,16-pentaoxa-10,19-diazacycloheneicosane)  
 $C_{58}H_{112}N_4O_{11}$   $M_r$  1041.53 [160563-01-9]

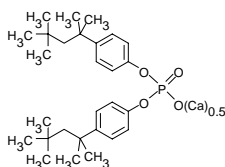
[21203](#) **Selectophore®**, function tested 25 mg, 250 mg



### Calcium ionophore VI

(*tert*-Butyl-calix[4]arene tetrakis[2-(diphenylphosphoryl)ethyl ether])  
 $C_{100}H_{108}O_8P_4$   $M_r$  1561.82 [171979-66-1]

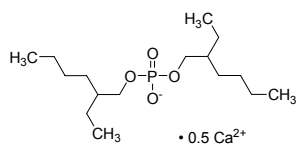
[72385](#) **Selectophore®**, function tested 50 mg, 250 mg



### Bis[4-(1,1,3,3-tetramethylbutyl)phenyl]phosphate Calcium salt

(hemi-Calcium bis[4-(1,1,3,3-tetramethylbutyl)phenyl] phosphate)  
 $C_{28}H_{42}Ca_{0.5}O_4P$   $M_r$  493.66 [40835-97-0]

[15180](#) **Selectophore®**, function tested 1 g

**Bis(2-ethylhexyl) phosphate hemicalcium salt**

(hemi-Calcium bis(2-ethylhexyl) phosphate)

C<sub>32</sub>H<sub>70</sub>Ca<sub>0.5</sub>O<sub>8</sub>P<sub>2</sub> M<sub>r</sub> 684.92 [10442-05-4][08733](#) **Selectophore®** 1 g**Calcium ionophore I - Cocktail A**

Calcium-selective membrane solution for microelectrodes

[21048](#) **Selectophore®** package with 0.1 mL**Calcium ionophore II - Cocktail A**

Calcium-selective membrane solution for microelectrodes

[21196](#) **Selectophore®** package with 0.1 mL**Ion-Selective Electrodes****Microelectrodes****Ion-Selective Field Effect Transistors****Ion-Selective Conductometric Microsensors****Optical Transduction**

## Electrochemical Transduction

### Ion-Selective Electrodes

#### Application 1 and Sensor Type <sup>1,2,3</sup>

Assay of Ca<sup>2+</sup> activity in whole blood, plasma, serum (ionized or total calcium) with solvent polymeric membrane electrodes based on Calcium ionophore I.

#### Recommended Membrane Composition

3.30	wt%	Calcium ionophore I ( <a href="#">21192</a> )
63.70	wt%	Bis(1-butylpentyl)decan-1,10-diyl diglutarate ( <a href="#">30585</a> )*
2.10	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
30.90	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

\* The use of bis(1-butylpentyl)adipate ([02150](#)) or bis(2-ethylhexyl)sebacate ([84818](#)) leads to membrane electrodes of similar performance

#### Recommended Cell Assembly

Reference || sample solution || ion-selective membrane | 0.001 M CaCl<sub>2</sub> | AgCl, Ag

#### Electrode Characteristics and Function

Selectivity coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M solutions of the chlorides)

	required <sup>1)</sup>	found
$\log K_{Ca, H}^{Pot}$	< -2.3	-2.9
$\log K_{Ca, Na}^{Pot}$	< -3.6	-3.7
$\log K_{Ca, K}^{Pot}$	< -0.6	-3.7
$\log K_{Ca, Mg}^{Pot}$	< -1.9	-4.7

Stability: Drift 0.01 mV/h

Standard deviation: 0.03 mV

Reproducibility: 0.13 mV

Lifetime:	$\log P_{TLC}^{2)}$ ionophore	> 8.4	7.5
	plasticizer	> 12.8	10.8

<sup>1)</sup> for measurements in whole blood (1% interference, worst case)<sup>4,5</sup>

<sup>2)</sup> lipophilicity, determined by thin layer chromatography<sup>6</sup>

<sup>1</sup> D. Ammann, P. Anker, E. Metzger, U. Oesch, W. Simon, in: Ion Measurements in Physiology and Medicine, Eds. M. Kessler, D.K. Harrison, J. Höper, Springer-Verlag, Berlin, Heidelberg 102 (1985).

<sup>2</sup> P. Anker, E. Wieland, D. Ammann, R.E. Dohner, R. Asper, W. Simon, Neutral carrier based ion-selective electrode for the determination of total calcium in blood serum. **Anal. Chem.** **53**, 1970 (1981).

<sup>3</sup> P. Anker, D. Ammann, P.C. Meier, W. Simon, Neutral carrier electrode for continuous measurement of blood Ca<sup>2+</sup> in the extracorporeal circulation. **Clin. Chem.** **30**, 454 (1984).

<sup>4</sup> A. Lewenstam, Ion selective electrodes in clinical chemistry. **Anal. Proc.** **28**, 106 (1991).

<sup>5</sup> U. Oesch, P. Anker, D. Ammann, W. Simon, in: Ion-Selective Electrodes, Eds. E. Pungor, I. Buzás, Akadémiai Kiadó, Budapest 81 (1985).

<sup>6</sup> O. Dinten, U.E. Spichiger, N. Chaniotakis, P. Gehrig, B. Rusterholz, W.E. Morf, W. Simon, Lifetime of neutral-carrier-based liquid membranes in aqueous samples and blood and the lipophilicity of membrane components, **Anal. Chem.** **63**, 596 (1991).

### Application 2 and Sensor Type <sup>7,8</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on Calcium ionophore II, the detection limit lying in the sub-nanomolar range.

#### Recommended Membrane Composition

1.00	wt%	Calcium ionophore II ( <a href="#">21193</a> )
0.60	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
65.60	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )
32.80	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

#### Recommended Cell Assembly

Reference || sample solution || ion-selective membrane | 0.01 M CaCl<sub>2</sub> + 5 • 10<sup>-2</sup> EDTA, pH 8.5 | Ag, AgCl

#### Electrode Characteristics and Function

Selectivity coefficients logK<sub>Ca, M</sub><sup>Pot</sup> as obtained by the fixed interference method (0.1 M buffered solutions of the chlorides).

logK<sub>Ca, K</sub><sup>Pot</sup> - 10.1                      logK<sub>Ca, Na</sub><sup>Pot</sup> - 8.3

logK<sub>Ca, Mg</sub><sup>Pot</sup> - 9.3

Slope of linear regression: 33.4±0.1 mV/dec (membranes conditions in 0.01 M NaCl)

Detection limit (Ca<sup>2+</sup>-buffered solutions containing 94 mM Na<sup>+</sup>): log a<sub>Ca</sub> ~ - 9.7

Detection limit (Ca<sup>2+</sup>-buffered solutions containing 125 mM Na<sup>+</sup>): log a<sub>Ca</sub> ~ - 10.1

Lifetime: log P<sub>TLC</sub><sup>1)</sup> ionophore: 7.2

Response time: 90% response time 2.5 s

<sup>1)</sup> lipophilicity, determined by thin layer chromatography<sup>6</sup>

### Application 3 and Sensor Type <sup>9</sup>

Assay of Ca<sup>2+</sup> activity in blood serum with solvent polymeric membrane electrodes with good potential stability and reproducibility, based on Calcium ionophore II.

#### Recommended Membrane Composition

1.0	wt%	Calcium ionophore II ( <a href="#">21193</a> )
1.0	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
65.0	wt%	Bis(2-ethylhexyl)phthalate ( <a href="#">80030</a> )
33.0	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

#### Recommended Cell Assembly

Reference || sample solution || liquid membrane | 0.001 M CaCl<sub>2</sub>, 0.1 M NaCl | AgCl, Ag

#### Electrode Characteristics and Function

Selectivity coefficients logK<sub>Ca, M</sub><sup>Pot</sup> as obtained by the fixed interference method in Ca<sup>2+</sup>-buffered solution

(M: Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Cl<sup>-</sup>)

logK<sub>Ca, Na</sub><sup>Pot</sup> -2.9                      logK<sub>Ca, NH<sub>4</sub></sub><sup>Pot</sup> -2.9

logK<sub>Ca, K</sub><sup>Pot</sup> -2.7                      logK<sub>Ca, Mg</sub><sup>Pot</sup> -3.2

Slope of linear regression: 28 mV/dec (2 • 10<sup>-7</sup> to 10<sup>-1</sup> M Ca<sup>2+</sup>)

Detection limit: 2 • 10<sup>-9</sup> M Ca<sup>2+</sup>

Practical pH measuring range: 3-8

Response time: 90% response time 4 s (10<sup>-5</sup> to 10<sup>-4</sup> M Ca<sup>2+</sup>)

<sup>7</sup> E. Bakker, Determination of Unbiased Selectivity Coefficients of Neutral Carrier-Based Cation-Selective Electrodes. **Anal. Chem.** **69**, 1061 (1997).

<sup>8</sup> Bedlechowicz-Sliwakowska I, Lingenfelter P, Sokalski T, Lewenstam A, Maj-Zurawska M., Ion-selective electrode for measuring low Ca<sup>2+</sup> concentrations in the presence of high K<sup>+</sup>, Na<sup>+</sup> and Mg<sup>2+</sup> background. **Anal. Bioanal. Chem.** **385(8)**, 1477 (2006).

<sup>9</sup> Y. Ma, X. Rao, S. Zhong, S. Ren, T. Yu, Q. Zhen, A study of calcium ion-selective PVC membrane electrode based on neutral carrier N,N,N',N'-tetracyclo-3-oxapentanediamide (correction of oxapentanediamide). **J. Tongji Med. Univ.** **12**, 98 (1992).

#### Application 4 and Sensor Type <sup>10</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on the highly lipophilic Calcium ionophore IV.

##### Recommended Membrane Composition

1.00	wt%	Calcium ionophore IV ( <a href="#">21198</a> )
0.28	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
65.82	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )
32.90	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

##### Recommended Cell Assembly

Reference || sample solution || ion-selective electrode | 0.01 M CaCl<sub>2</sub> | AgCl,Ag

##### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M solutions of the chlorides).<sup>10</sup>

$\log K_{Ca, H}^{Pot}$	-3.1	$\log K_{Ca, K}^{Pot}$	-7.5
$\log K_{Ca, Li}^{Pot}$	-5.8	$\log K_{Ca, Mg}^{Pot}$	-4.4
$\log K_{Ca, Na}^{Pot}$	-5.9		

Slope of linear regression: 29.7±1.7 mV/dec (10<sup>-8</sup> to 10<sup>-1</sup> M CaCl<sub>2</sub>)

Detection limit (CaCl<sub>2</sub> ion background of 125 mM KCl): log a<sub>Ca</sub> ~-9.7

Lifetime: log P<sub>TLC</sub><sup>1)</sup> ionophore: 22.6 ± 3.7

Response time: 90% response time 1.2 s (10<sup>-4</sup> to 10<sup>-3</sup> M CaCl<sub>2</sub>)

<sup>1)</sup> lipophilicity, determined by thin layer chromatography<sup>6</sup>

#### Application 5 and Sensor Type <sup>11</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on Calcium ionophore V.

##### Recommended Membrane Composition

2.0	wt%	Calcium ionophore V ( <a href="#">21203</a> )
0.9	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
66.0	wt%	Nitrophenyl octyl ether ( <a href="#">73732</a> )
31.1	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

##### Recommended Cell Assembly

Reference || sample solution || ion-selective electrode | 0.1 M CaCl<sub>2</sub> | AgCl,Ag

##### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M of the chloride salts).

$\log K_{Ca, H}^{Pot}$	-3.6	$\log K_{Ca, Cs}^{Pot}$	-4.0
$\log K_{Ca, Li}^{Pot}$	-4.1	$\log K_{Ca, NH_4}^{Pot}$	-4.2
$\log K_{Ca, Na}^{Pot}$	-4.1	$\log K_{Ca, Mg}^{Pot}$	-5.0
$\log K_{Ca, K}^{Pot}$	-4.4	$\log K_{Ca, Sr}^{Pot}$	-1.0
$\log K_{Ca, Rb}^{Pot}$	-4.2	$\log K_{Ca, Ba}^{Pot}$	-2.1

Lifetime: log P<sub>TLC</sub><sup>1)</sup> ionophore: 14.6

<sup>1)</sup> lipophilicity, determined by thin layer chromatography<sup>6</sup>

<sup>10</sup> P. Gehrig, B. Rusterholz, W. Simon, Very lipophilic calcium. ion-selective ionophore for chemical sensors of high life-time. *Chimia* **43**, 377 (1989).

<sup>11</sup> K. Suzuki, K. Watanabe, Y. Matsumoto, M. Kobayashi, S. Sato, D. Siswanta, H. Hisamoto, Design and Synthesis of Calcium and Magnesium Ionophores Based on Double-Armed Diazacrown Ether Compounds and Their Application to an Ion Sensing Component for an Ion-Selective Electrode. *Anal. Chem.* **67**, 324 (1995).

### Application 6 and Sensor Type <sup>12</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on Calcium ionophore VI.

#### Recommended Membrane Composition

0.66	wt%	Calcium ionophore VI ( <a href="#">72385</a> )
0.07	wt%	Potassium tetrakis(4-chlorophenyl)borate ( <a href="#">60591</a> )
66.18	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )
33.09	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

#### Recommended Cell Assembly

Reference || sample solution || ion-selective electrode | 0.1 M CaCl<sub>2</sub> | AgCl,Ag

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M of the chloride salts).

$$\log K_{Ca, Na}^{Pot} = -2.2 \qquad \log K_{Ca, Li}^{Pot} = -1.6$$

$$\log K_{Ca, K}^{Pot} = -2.7 \qquad \log K_{Ca, Mg}^{Pot} = -2.6$$

$$\log K_{Ca, NH_4}^{Pot} = -2.0$$

Slope of linear regression: 26.35 mV (10<sup>-4</sup> to 10<sup>-1</sup> CaCl<sub>2</sub>)

### Application 7 and Sensor Type <sup>13</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on the liquid ion-exchanger Bis[4-(1,1,3,3-tetramethylbutyl)phenyl]phosphate Calcium salt.

#### Recommended Membrane Composition

7.00	wt%	Bis[4-(1,1,3,3-tetramethylbutyl)phenyl]phosphate Calcium salt ( <a href="#">15180</a> )
29.86	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
63.14	wt%	Di-n-octylphenylphosphonate ( <a href="#">12584</a> )

#### Recommended Cell Assembly

Reference || sample solution || ion-selective electrode | 0.1 M CaCl<sub>2</sub> | AgCl,Ag

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M of the chloride salts).

$$\log K_{Ca, Na}^{Pot} = -4.4 \qquad \log K_{Ca, Mg}^{Pot} = -4.9$$

$$\log K_{Ca, K}^{Pot} = -4.5$$

Slope of linear regression: 30.5 at 25°C (10<sup>-5</sup> to 10<sup>-0</sup> Ca<sup>2+</sup>)

Detection limit: 3.2 · 10<sup>-6</sup> M Ca<sup>2+</sup>

pH range for 10<sup>-3</sup> M CaCl<sub>2</sub>: 4.8 to 8.8

Response time: <10 s

Operational lifetime: 3 months

Membrane resistance: 3 MΩ

<sup>12</sup> T. McKittrick, D. Diamond, D.J. Marrs, P. O'Hagan, M. Anthony McKervey. Calcium-selective electrode based on a calix[4]arene tetraphosphine oxide, **Talanta** **43**, 1145 (1996).

<sup>13</sup> G.J. Moody, The role of polymeric materials in the fabrication of ion-selective electrodes and biosensors. **Polym. Mat. Sci. Eng.** **64**, 362, (1991) and ref. cited therein.

### Application 8 and Sensor Type <sup>14</sup>

Assay of Ca<sup>2+</sup> activity with solvent polymeric membrane electrodes based on the liquid ion-exchanger Bis[4-(1,1,3,3-tetramethylbutyl)phenyl]phosphate Calcium salt.

#### Recommended Membrane Composition

0.10	wt%	Bis[4-(1,1,3,3-tetramethylbutyl)phenyl]phosphate Calcium salt ( <a href="#">15180</a> )
33.43	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
66.43	wt%	Bis(2-ethylhexyl)sebacate ( <a href="#">84818</a> )

#### Recommended Cell Assembly

Reference || sample solution || ion-selective electrode | 0.1 M CaCl<sub>2</sub> | AgCl,Ag

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the separate solution method (0.1 M of the chloride salts).

$\log K_{Ca, NH_4}^{Pot}$	-3.3	$\log K_{Ca, H}^{Pot}$	-2.4
$\log K_{Ca, Li}^{Pot}$	-4.1	$\log K_{Ca, Mg}^{Pot}$	-6.2
$\log K_{Ca, Na}^{Pot}$	-4.0	$\log K_{Ca, Ba}^{Pot}$	-3.0
$\log K_{Ca, K}^{Pot}$	-3.0		

Slope of linear regression: 31.9 ± 2.3 mV (10<sup>-4</sup> to 10<sup>-1</sup> CaCl<sub>2</sub>)

Membrane resistance: 9.9 ± 1.0 MΩ

<sup>14</sup> U. Schaller, E. Bakker, E. Pretsch, Carrier mechanism of acidic ionophores in solvent polymeric membrane ion-selective electrodes. *Anal. Chem.* **67**, 3123 (1995).

## Microelectrodes

### Application 1 and Sensor Type general 15, 16, 17, application 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31

Assay of Ca<sup>2+</sup> activity in extra- and intracellular (single-cell) liquids with Ca<sup>2+</sup> microelectrodes of tip diameter > 1 μm based on Calcium ionophore I.

Calcium ionophore I - Cocktail A ([21048](#))

#### Cocktail Composition:

10.0	wt%	Calcium ionophore I ( <a href="#">21192</a> )
89.0	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )
1.0	wt%	Sodium tetraphenylborate ( <a href="#">72018</a> )

#### Recommended Cell Assembly

Reference || sample solution || cocktail | 0.001 M CaCl<sub>2</sub> + 0.011 M NTA + 0.047 M Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub> | AgCl,Ag

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the fixed interference method in Ca<sup>2+</sup> buffered solutions (for M: Na<sup>+</sup>, K<sup>+</sup>) or Ca<sup>2+</sup>-unbuffered solutions (for M: Mg<sup>2+</sup>).<sup>15</sup>

$\log K_{Ca, Na}^{Pot}$	-5.5	$\log K_{Ca, Mg}^{Pot}$	<-4.9
$\log K_{Ca, K}^{Pot}$	-5.4		

Slope of linear regression: 28.1 ± 1.8 mV (10<sup>-7</sup> to 10<sup>-2</sup> M CaCl<sub>2</sub>)

Detection limit (Ca<sup>2+</sup>-buffered solutions, constant ion background of 125 mM K<sup>+</sup>): log a<sub>Ca</sub> ~-7.4

Electrical resistance tip diameter 1 to 2 μm: ~2·10<sup>10</sup> Ω

Response time 90% response time: ≤ 5 s<sup>15</sup>

Time constant: τ = 7 ms

<sup>15</sup> F. Lanter, R.A. Steiner, D. Ammann, W. Simon, Critical evaluation of the applicability of neutral carrier-based calcium selective microelectrodes. **Anal. Chim. Acta** **135**, 51 (1982).

<sup>16</sup> R.Y. Tsien, T.J. Rink, Ca<sup>2+</sup>-selective electrodes: a novel PVC-gelled neutral carrier mixture compared with other currently available sensors. **J. Neurosci. Methods** **4**, 73 (1981).

<sup>17</sup> E. Ujec, E.E.O. Keller, N. Kõiz, V. Pavlik, J. Machek, Low-impedance, coaxial, ion-selective, double-barrel microelectrodes and their use in biological measurements. **Bioelectrochem. Bioenerg.** **7**, 363 (1980).

<sup>18</sup> E. Marban, T.J. Rink, R.W. Tsien, R.Y. Tsien, Free calcium in heart muscle at rest and during contraction measured with Ca<sup>2+</sup>-sensitive microelectrodes. **Nature** **286**, 845 (1980).

<sup>19</sup> D.M. Bers, D. Ellis, Changes of intracellular calcium and sodium activities in sheep heart Purkinje fibres measured with ion-selective micro-electrodes. **J. Physiol.** **310**, 73P (1981).

<sup>20</sup> R. Pumain, I. Kurcewicz, J. Louvel, Fast extracellular calcium transients: involvement in epileptic processes. **Science** **222**, 177 (1983).

<sup>21</sup> R. Weingart, P. Hess, Free calcium in sheep cardiac tissue and frog skeletal muscle measured with Ca<sup>2+</sup>-selective microelectrodes. **Pflügers Arch.** **402**, 1 (1984).

<sup>22</sup> C. Nicholson, Modulation of extracellular calcium and its functional implications. **Fed. Proc.** **39**, 1519 (1980).

<sup>23</sup> A.L.F. Gorman, S. Levy, E. Nasi, D. Tillotson, Intracellular calcium measured with calcium-sensitive micro-electrodes and Arsenazo III in voltage-clamped Aplysia neurones. **J. Physiol.** **353**, 127 (1984).

<sup>24</sup> R. DiPolo, H. Rojas, J. Vergara, R. Lopez, C. Caputo, Measurements of intracellular ionized calcium in squid giant axons using calcium-selective electrodes. **Biochim. Biophys. Acta** **728**, 311 (1983).

<sup>25</sup> F.J. Alvarez-Leefmans, T.J. Rink, R.Y. Tsien, Free calcium ions in neurones of Helix aspersa measured with ion-selective micro-electrodes. **J. Physiol.** **315**, 531 (1981).

<sup>26</sup> M.J. Berridge, Preliminary measurements of intracellular calcium in an insect salivary gland using a calcium-sensitive microelectrode. **Cell Calcium** **1**, 217 (1980).

<sup>27</sup> S. Levy, A. Fein, Relationship between light sensitivity and intracellular free Ca concentration in Limulus ventral photoreceptors. A quantitative study using Ca-selective microelectrodes. **J. Gen. Physiol.** **85**, 805 (1985).

<sup>28</sup> A. Picard, M. Dorée, Is calcium the second messenger of 1-methyladenine in meiosis reinitiation of starfish oocytes? **Exp. Cell. Res.** **145**, 325 (1983).

<sup>29</sup> K.P. Dresdner, R.P. Kline, Extracellular calcium ion depletion in frog cardiac ventricular muscle. **Biophys. J.** **48**, 33 (1985).

<sup>30</sup> H. Yamaguchi, Recording of intracellular Ca<sup>2+</sup> from smooth muscle cells by sub-micron tip, double-barrelled Ca<sup>2+</sup>-selective microelectrodes. **Cell Calcium** **7**, 203 (1986).

<sup>31</sup> E. Kelepouris, Z.S. Agus, M.M. Civan, Intracellular calcium activity in split frog skin epithelium: effect of cAMP. **J. Membr. Biol.** **88**, 113 (1985).

### Application 2 and Sensor Type <sup>32,33,34</sup>

Assay of Ca<sup>2+</sup> activity in extra- and intracellular (single cell) liquids with Ca<sup>2+</sup> microelectrodes based on Calcium ionophore II.

Calcium ionophore II - Cocktail A

#### Cocktail Composition

5.0	wt%	Calcium ionophore II ( <a href="#">21193</a> )
94.0	wt%	2-Nitrophenyl octyl ether (o-NPOE) ( <a href="#">73732</a> )
31.	wt%	Sodium tetraphenylborate ( <a href="#">72018</a> )

#### Recommended Cell Assembly

Reference | sample solution | | cocktail | 10<sup>-7</sup> M CaCl<sub>2</sub> | AgCl, Ag

#### Use

Before use, mix cocktail A with 14 wt% poly(vinyl chloride) ([81392](#)) to obtain a stable and reproducible response<sup>32</sup>.

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the fixed interference method in Ca<sup>2+</sup>-buffered solutions (M: Na<sup>+</sup>, K<sup>+</sup>) or Ca<sup>2+</sup>-unbuffered solutions M: Mg<sup>2+</sup>).

$\log K_{Ca, Na}^{Pot}$	-5.6	$\log K_{Ca, Mg}^{Pot}$	-6.7
$\log K_{Ca, K}^{Pot}$	-7.2		

Slope of linear regression: 29.9 mV (10<sup>-8</sup> to 10<sup>-3</sup> M Ca<sup>2+</sup>)

Detection limit (Ca<sup>2+</sup>-buffered solutions, constant ion background of 125 mM K<sup>+</sup>): log a<sub>Ca</sub> ~-9.2

Electrical resistance, tip diameter 1 to 1.5 μm, filling height 78 to 100 μm: ~4 · 10<sup>10</sup> Ω

Response Time 90% response time: ≤ 5 s

### Application 3 and Sensor Type <sup>32,33,34</sup>

Assay of Ca<sup>2+</sup> activity in intra- and extracellular (single cell) liquids with Ca<sup>2+</sup>-microelectrodes of tip diameter <1 μm based on Calcium ionophore II.

#### Cocktail Composition for electrode tip diameter <1 μm

21.50	wt%	Calcium ionophore II ( <a href="#">21193</a> )
3.50	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
75.00	wt%	Tetrahydrofuran ( <a href="#">87396</a> )

#### Recommended Cell Assembly

Reference | sample solution | | cocktail | 0.01 M CaCl<sub>2</sub> | AgCl, Ag

#### Electrode Characteristics and Function

See Calcium ionophore II - Cocktail A ([21196](#))

<sup>32</sup> D. Ammann, T. Bührer, U. Schefer, M. Müller, W. Simon, Intracellular neutral carrier-based Ca<sup>2+</sup> microelectrode with subnanomolar detection limit. *Pflügers Arch.* **409**, 223 (1987).

<sup>33</sup> H.M. Brown, S.K. Marron, Fabrication method to enhance stability of N,N,N',N'-tetracyclohexyl-3-oxapentanediamide calcium microelectrodes. *Anal. Chem.* **62**, 2153 (1990).

<sup>34</sup> D. Ammann, P. Caroni, Preparation and use of micro- and macroelectrodes for measurement of transmembrane potentials and ion activities. *Methods in Enzymol.* **172**, 136 (1989).

## Ion-Selective Field Effect Transistors

### Application 1 and Sensor Type <sup>35</sup>

Assay of Ca<sup>2+</sup> activity with Urushi matrix ion-selective field effect transistors of good durability based on Calcium ionophore I.

#### Recommended Membrane Composition:

5.0	wt%	Calcium ionophore I ( <a href="#">21192</a> )
44.0	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )
50.0	wt%	Urushi latex
1.0	wt%	Sodium tetraphenylborate ( <a href="#">72018</a> )

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$  as obtained by the fixed interference method in Ca<sup>2+</sup> buffered solutions (for M: Na<sup>+</sup>, K<sup>+</sup>) or Ca<sup>2+</sup>-unbuffered solutions (for M: Mg<sup>2+</sup>).<sup>15</sup>

$\log K_{Ca, Na}^{Pot}$	-4.8	$\log K_{Ca, NH_4}^{Pot}$	-4.4
$\log K_{Ca, K}^{Pot}$	-5.8	$\log K_{Ca, Mg}^{Pot}$	-4.6

Slope of linear regression: 25 mV (10<sup>-5.5</sup> to 10<sup>-1.5</sup> M Ca<sup>2+</sup>)

### Application 2 and Sensor Type <sup>36</sup>

Determination of Ca<sup>2+</sup> with an ion-selective field effect transistor based on a photo-curable polysiloxane membrane containing Calcium ionophore IV.

#### Recommended Membrane Composition:

0.99	wt%	Calcium ionophore IV ( <a href="#">21198</a> )
0.55	wt%	Potassium tetrakis[3.5-bis(trifluoromethyl)phenyl]borate ( <a href="#">60588</a> )
0.88	wt%	2,2-Dimethoxy-2-phenylacetophenone ( <a href="#">38781</a> )
0.88	wt%	Dibutyltin dilaurate ( <a href="#">34930</a> )
8.79	wt%	3-(Trimethoxysilyl)propyl methacrylate ( <a href="#">64210</a> )
87.91	wt%	10-12%(3-cyanopropyl)methyl/88-90% (dimethylsiloxane) copolymer

#### Electrode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Pot}$

$\log K_{Ca, Na}^{Pot}$	< -3.5	$\log K_{Ca, Mg}^{Pot}$	< -4.0
$\log K_{Ca, K}^{Pot}$	< -3.5		

Slope of linear regression: Nernstian behaviour (10<sup>-5</sup> to 10<sup>-1</sup> M CaCl<sub>2</sub>)

<sup>35</sup> S.I. Wakida, M. Yamane, K. Higashi, K. Hiro, Y. Ujihara, Urushi matrix sodium, potassium, calcium and chloride-selective field-effect transistors. **Sens. Actuators B1**, 412 (1990).

<sup>36</sup> P.D. Van der Wal, A. Van den Berg, N.A. de Rooij, Universal approach for the fabrication of Ca<sup>2+</sup>-, K<sup>+</sup>- and NO<sub>3</sub><sup>-</sup>-sensitive membrane ISFETs. **Sens. Actuators B 18-19**, 200 (1994).

## **Ion-Selective Conductometric Microsensors**

### **Application and Sensor Type** <sup>37</sup>

Assay of Ca<sup>2+</sup> activity with ion-selective conductometric microsensors (ISCOM). Detection is accomplished by measurement of the bulk conductance of the solvent polymeric membrane based on Calcium ionophore IV.

#### *Recommended Membrane Composition:*

5.0	wt%	Calcium ionophore IV ( <a href="#">21198</a> )
30.0	w%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
65.0	wt%	2-Nitrophenyl octyl ether ( <a href="#">73732</a> )

#### *Electrode Characteristics and Function*

Detection limit:  $\sim 10^{-7}$  M Ca<sup>2+</sup> (measurements in 1 M NaNO<sub>3</sub>)

Response time:  $\sim 2$  s

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<sup>37</sup> A.A. Shul'ga, B. Ahlers, K. Cammann, Ion-selective conductometric microsensors based on the phenomenon of specific salt extraction. **J. Electroanal. Chem.** **395**, 305 (1995).

## Optical Transduction

### Application 1 and Sensor Type <sup>38,39</sup>

Assay of Ca<sup>2+</sup> activity in aqueous pH-buffered solutions with polymeric optode membranes based on Chromoionophore I (ETH 5294) and Calcium ionophore I.

#### Recommended Membrane Composition

2.20	wt%	Chromoionophore I ( <a href="#">27086</a> )
8.01	wt%	Calcium ionophore I ( <a href="#">21192</a> )
4.49	wt%	Bis(2-ethylhexyl)sebacate ( <a href="#">84818</a> )
57.27	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
28.03	wt%	Sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate ( <a href="#">72017</a> )

#### Recommended pH Buffer

0.16 M sodium acetate, adjusted with acetic acid to pH 5.3 for recording the calibration curve.<sup>40</sup>

#### Optode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Opt}$  as obtained by the separate solution method in pH-buffered solutions.

$\log K_{Ca, Na}^{Opt}$	-3.6	$\log K_{Ca, Mg}^{Opt}$	-4.1
$\log K_{Ca, K}^{Opt}$	-3.8		

### Application 2 and Sensor Type <sup>41</sup>

Assay of Ca<sup>2+</sup> activity in aqueous pH-buffered solutions with solvent polymeric optode membranes based on Chromoionophore III (ETH 5350) and Calcium ionophore II.

#### Recommended Membrane Composition

9.20	wt%	Calcium ionophore II ( <a href="#">21193</a> )
2.20	wt%	Chromoionophore III ( <a href="#">27088</a> )
4.50	wt%	Sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate ( <a href="#">72017</a> )
56.40	wt%	Bis(2-ethylhexyl)sebacate ( <a href="#">84818</a> )
27.80	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

#### Recommended pH Buffer

0.1 M sodium acetate adjusted to pH 5.4 with 3 M acetic acid.

#### Optode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, Na}^{Opt}$  -6.1 as obtained by the fixed interference method.<sup>38</sup>

Response time: ~1 min

<sup>38</sup> Fluka [58166](#): K. Seiler, Ion-selective Optode Membranes, monograph, describing theory, preparation and application of ion-selective optode membranes as well as recent developments in this field. With 237 references. published by Fluka Chemie GmbH, Buchs, Switzerland (1993).

Fluka [58165](#): K. Seiler, Ionenselektive Optodenmembranen, dt. Monographie, herausgegeben von Fluka Chemie GmbH, Buchs, Switzerland (1993).

<sup>39</sup> W.E. Morf, K. Seiler, B. Rusterholz, W. Simon, Design of a novel calcium-selective optode membrane based on neutral ionophores. **Anal. Chem.** **62**, 738 (1990).

<sup>40</sup> D.D. Perrin, B. Dempsey, Buffers for pH and Metal Ion Control. Chapman & Hall, London, New York (1983).

<sup>41</sup> T. Rosatzin, P. Holy, K. Seiler, B. Rusterholz, W. Simon, Immobilization of components in polymer membrane-based calcium-selective bulk optodes. **Anal. Chem.** **64**, 2029 (1992).

### Application 3 and Sensor Type<sup>38,42</sup>

Assay of Ca<sup>2+</sup> activity in aqueous pH-buffered solutions and in diluted blood plasma with solvent polymeric optode membranes based on Chromoionophore II (ETH 2439) and Calcium ionophore I.

#### Recommended Membrane Composition

1.47	wt%	Chromoionophore II ( <a href="#">27087</a> )
5.98	wt%	Calcium ionophore I ( <a href="#">21192</a> )
2.56	wt%	Sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate ( <a href="#">72017</a> )
60.90	wt%	Bis(2-ethylhexyl)sebacate ( <a href="#">84818</a> )
29.09	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )

#### Recommended pH Buffer

0.02 M sodium hydroxide adjusted to pH 3.3 with acetic acid.

### Application 4 and Sensor Type<sup>43</sup>

Assay of Ca<sup>2+</sup> activity in diluted human plasma with solvent polymeric optode membranes based on Chromoionophore I (ETH 5294) and Calcium ionophore I.

#### Recommended Membrane Composition

2.20	wt%	Chromoionophore I ( <a href="#">27086</a> )
8.01	wt%	Calcium ionophore I ( <a href="#">21192</a> )
4.49	wt%	Bis(2-ethylhexyl)sebacate ( <a href="#">84818</a> )
57.27	wt%	Poly(vinyl chloride) high molecular weight ( <a href="#">81392</a> )
28.03	wt%	Sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate ( <a href="#">72017</a> )

#### Recommended pH Buffer

Sodium acetate type at pH 3.52

#### Optode Characteristics and Function

Selectivity Coefficients  $\log K_{Ca, M}^{Opt}$  as obtained by the fixed interference method.<sup>38</sup>

$\log K_{Ca, Li}^{Opt}$	-3.1	$\log K_{Ca, K}^{Opt}$	-3.8
$\log K_{Ca, Na}^{Opt}$	-3.6	$\log K_{Ca, Mg}^{Opt}$	-4.1

### Application 5 and Sensor Type<sup>44</sup>

Flow-through type Ca<sup>2+</sup> ion selective optodes for determination of Ca<sup>2+</sup> in biological samples such as human serum based on Calcium ionophore V.

#### Recommended Membrane Composition

1.0	wt%	Calcium ionophore V ( <a href="#">21203</a> )
1.2	wt%	Chromophore (LAD-3)*
31.0	wt%	o-trifluoromethylphenyl dodecyl ether (TFPDE)*
66.8	wt%	ODS beads*

\*not available from Sigma-Aldrich

#### Recommended pH Buffer

0.05 M Tris-HCl, pH 7.0

<sup>42</sup> K. Seiler, R. Eugster, W.E. Morf, K. Wang, M. Czösz, B. Rusterholz, W. Simon, U.E. Spichiger, Application of calcium optode in human plasma. **Fresenius J. Anal. Chem.** **337**, 109 (1990).

<sup>43</sup> U.E. Spichiger, K. Seiler, K. Wang, G. Suter, W.E. Morf, W. Simon, Optical quantification of sodium, potassium, and calcium ions in diluted human plasma based on ion-selective liquid membranes. **Proc. SPIE-Int. Soc. Opt. Eng.** **1510**, 118 (1991).

<sup>44</sup> H. Hisamoto, K. Watanabe, E. Nakagawa, D. Siswanta, Y. Shichi, K. Suzuki, Flow-through type calcium ion selective optodes based on novel neutral ionophores and a lipophilic anionic dye. **Anal. Chim. Acta** **299**, 179 (1994).

*Optode Characteristics and Function*

Selectivity Coefficients  $\log K_{Ca, M}^{Opt}$  as obtained by the separate solution method in pH buffered solutions at 516 nm.

$\log K_{Ca, Li}^{Opt}$	-4.0	$\log K_{Ca, NH_4}^{Opt}$	-4.4
$\log K_{Ca, Na}^{Opt}$	-4.8	$\log K_{Ca, Mg}^{Opt}$	-4.0
$\log K_{Ca, K}^{Opt}$	-4.1	$\log K_{Ca, Ba}^{Opt}$	-1.7

Concentration range:  $10^{-7}$  M to  $10^{-1}$  M.