

**Determination of the Concentration and Molecular Weight  
of L-a-GLYCEROPHOSPHATE**

**PRINCIPLE:**

L-a-Glycerophosphate +  $\beta$ -NAD + Hydrazine  $\xrightarrow{\text{GDH}}$  DAP Hydrazone + NADH

Abbreviations used:

$\beta$ -NAD =  $\beta$ -Nicotinamide Adenine Dinucleotide, Oxidized Form

GDH =  $\alpha$ -Glycerophosphate Dehydrogenase

DAP = Dihydroxyacetone Phosphate

$\beta$ -NADH =  $\beta$ -Nicotinamide Adenine Dinucleotide, Reduced Form

**CONDITIONS:** T = 25°C, pH = 9.5, A<sub>340nm</sub>, Light path = 1 cm

**METHOD:** Spectrophotometric Determination

**REAGENTS:**

- A. 1 M Glycine Buffer with 609 mM Hydrazine Sulfate and 5.3 mM Ethylenediaminetetraacetic Acid, pH 9.5 at 25°C (Prepare 50 ml by dissolving Glycine, Free Base, Sigma Prod. No. G-7126, Hydrazine Sulfate, Sigma Prod. No. H-3376, and Ethylenediaminetetraacetic Acid, Tetrasodium Salt, Hydrate, Sigma Stock No. ED4SS, in 2 M NaOH until dissolved. Adjust to pH 9.5 with either 1 M NaOH or 1 M HCl. Then dilute to 50 ml with deionized water.<sup>1</sup> **PREPARE FRESH.**)
- B. 20 mM  $\beta$ -Nicotinamide Adenine Dinucleotide, Oxidized Form, Solution ( $\beta$ -NAD) (Prepare by dissolving the contents of one 50 mg vial  $\beta$ -Nicotinamide Adenine Dinucleotide, Sigma Stock No. 260-150, in the appropriate volume of deionized water. **PREPARE FRESH.**)
- C. Glycerophosphate Dehydrogenase Enzyme Solution (GDH) (Immediately before use, prepare a solution containing 1200 units/ml of  $\alpha$ -Glycerophosphate Dehydrogenase, Sigma Prod. No. G-6751, in cold deionized water.)
- D. L-a-Glycerophosphate Solution (L-a-GP) (Immediately before use, prepare a solution containing approximately 0.3  $\mu$ moles/ml of L-a-Glycerophosphate in deionized water.)

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**PROCEDURE:**

Pipette (in milliliters) the following reagents into suitable cuvettes:

	<u>Test</u>	<u>Blank</u>
Deionized Water	0.22	1.22
Reagent A (Buffer)	1.35	1.35
Reagent D (L-a-GP)	1.00	-----
Reagent B (β-NAD)	0.38	0.38

Mix by inversion and equilibrate to 25°C using a suitably thermostatted spectrophotometer. Record the initial  $A_{340\text{nm}}$  for the Test and Blank. Then add:

Reagent C (Enzyme Solution)	0.05	0.05
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Immediately mix by inversion and allow the reaction to proceed to completion (approximately 5 minutes). Record the final  $A_{340\text{nm}}$  for both the Test and Blank.

**CALCULATIONS:**

$$r A = A_f - A_i$$

$A_f$  = Final Absorbance

$A_i$  = Initial Absorbance

$$\text{Micromoles L-a-GP/ml RM} = \frac{(r A_{\text{Test}} - r A_{\text{Blank}})(3)}{(6.22)}$$

L-a-GP = L-a-Glycerophosphate

3 = Total volume (in milliliters) of assay

6.22 = Millimolar extinction coefficient of β-NADH at 340 nm

RM = Reaction Mixture

$$\text{Apparent Molecular Weight} = \frac{\text{mg/ml L-a-GP} \times 1000}{\mu\text{moles of L-a-GP/ml RM}}$$

1000 = Conversion factor from mg to μg

L-a-GP = L-a-Glycerophosphate

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**FINAL ASSAY CONCENTRATION:**

In a 3.00 ml reaction mix, the final concentrations are 450 mM glycine, 274 mM hydrazine sulfate, 2.4 mM ethylenediaminetetraacetic acid, 2.5 mM  $\beta$ -nicotinamide adenine dinucleotide, 60 units a-glycerophosphate dehydrogenase, and varying amounts of L-a-glycerophosphate.

**REFERENCE:**

Bergmeyer, H.U., Gawehn, K., and Grassl, M. (1974) in *Methods of Enzymatic Analysis*, (Bergmeyer, H.U. ed) Volume I, 2nd ed., 468-469, Academic Press, New York, NY

**NOTES:**

1. Once the pH of this solution has been adjusted to 9.5 at 25°C, it must be used within two hours. After that period of time, a baseline drift will occur when the solution is measured spectrophotometrically.
2. Where Sigma Product or Stock numbers are specified, equivalent reagents may be substituted.

**This procedure is for informational purposes. For a current copy of Sigma's quality control procedure contact our Technical Service Department.**