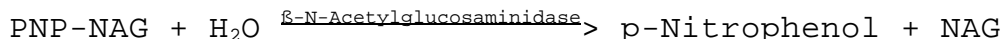


**Enzymatic Assay of  $\beta$ -N-ACETYLGLUCOSAMINIDASE  
(EC 3.2.1.30)  
from Jack Beans**

**PRINCIPLE:**



Abbreviations:

PNP-NAG = p-Nitrophenyl N-Acetyl- $\beta$ -D-Glucosaminide

NAG = N-Acetyl- $\beta$ -D-Glucosamine

**CONDITIONS:** T = 25°C, pH = 5.0, A<sub>400nm</sub>, Light path = 1 cm

**METHOD:** Spectrophotometric Stop Rate Determination

**REAGENTS:**

- A. 100 mM Citrate Buffer with 200 mM Sodium Chloride and 0.02% (w/v) Bovine Serum Albumin, pH 5.0 at 25°C  
(Prepare 100 ml in deionized water using Citric Acid, Free Acid, Monohydrate, Sigma Prod. No. C-7129, Sodium Chloride, Sigma Prod. No. S-9625, and Albumin, Bovine, Sigma Prod. No. A-4503. Adjust to pH 5.0 at 25°C with 1 M NaOH.)
- B. 10 mM p-Nitrophenyl N-Acetyl- $\beta$ -D-Glucosaminide Solution (PNP-NAG)  
(Prepare 5 ml in deionized water using p-Nitrophenyl N-Acetyl- $\beta$ -D-Glucosaminide, Sigma Prod. No. N-9376.)
- C. 200 mM Borate Buffer, pH 9.8 at 25°C  
(Prepare 100 ml in deionized water using Boric Acid, Sigma Prod. No. B-0252. Adjust to pH 9.8 at 25°C with 1 M NaOH.)
- D.  $\beta$ -N-Acetylglucosaminidase Enzyme Solution  
(Immediately before use, prepare a solution containing 0.05 - 0.1 unit/ml of  $\beta$ -N-Acetylglucosaminidase in cold Reagent A.)

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**PROCEDURE:**

Pipette (in milliliters) the following reagents into suitable containers:

	<u>Test</u>	<u>Blank</u>
Reagent A (Buffer)	0.40	0.40
Reagent B (PNP-NAG)	0.50	0.50

Mix by inversion and equilibrate to 25°C. Then add:

Reagent D (Enzyme Solution)	0.10	-----
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Mix by inversion and incubate for exactly 10 minutes at 25°C. Then add:

Reagent C (Borate Buffer)	3.00	3.00
Reagent D (Enzyme Solution)	-----	0.10

Mix by inversion and transfer to suitable cuvettes. Record the  $A_{400\text{nm}}$  for both the Test and Blank using a suitable spectrophotometer.

**CALCULATIONS:**

$$\text{Units/ml enzyme} = \frac{(\Delta A_{400\text{nm}} \text{ Test} - \Delta A_{400\text{nm}} \text{ Blank})(4)(\text{df})}{(10)(18)(0.1)}$$

4 = Total volume (in milliliters) of Assay

10 = Time of assay (in minutes) as per the Unit Definition

18 = Millimolar extinction coefficient of p-Nitrophenol  
at 400 nm<sup>1</sup>

0.1 = Volume (in milliliter) of enzyme used

$$\text{Units/mg solid} = \frac{\text{units/ml enzyme}}{\text{mg solid/ml enzyme}}$$

$$\text{Units/mg protein} = \frac{\text{units/ml enzyme}}{\text{mg protein/ml enzyme}}$$

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**UNIT DEFINITION:**

One unit will hydrolyze 1.0  $\mu$ mole of p-nitrophenyl N-acetyl- $\beta$ -D-glucosaminide to p-nitrophenol and N-acetyl- $\beta$ -D-glucosamine per minute at pH 5.0 at 25°C.

**FINAL ASSAY CONCENTRATION:**

In a 1.00 ml reaction mix, the final concentrations are 50 mM citric acid, 100 mM sodium chloride, 0.01% (w/v) bovine serum albumin, 5 mM p-nitrophenyl N-acetyl- $\beta$ -D-glucosaminide and 0.005 - 0.01 unit  $\beta$ -N-acetylglucosaminidase.

**REFERENCES:**

Bessey, O.A., Lowry, O.H., and Brock, M.J., (1946) *Journal of Biological Chemistry* **164**, 321-329

Li, S.-C. and Li, Y.-T. (1970) *Journal of Biological Chemistry* **245**, 5153-5160

Borooah, J., Leaback, D.H. and Walker, P.G. (1961) *Biochemical Journal* **78**, 106-110

**NOTES:**

1. The millimolar extinction coefficient is described in Bessey, O.A. et al. (1946).
2. This assay is based on the cited references.

**This procedure is for informational purposes. For a current copy of Sigma's quality control procedure contact our Technical Service Department.**