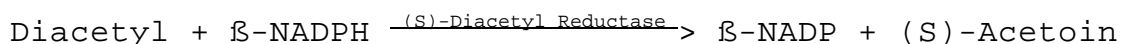


## Enzymatic Assay of (S)-DIACETYL REDUCTASE

### PRINCIPLE:



Abbreviations used:

$\beta$ -NADPH =  $\beta$ -Nicotinamide Adenine Dinucleotide Phosphate,  
Reduced Form

$\beta$ -NADP =  $\beta$ -Nicotinamide Adenine Dinucleotide Phosphate,  
Oxidized Form

**CONDITIONS:** T = 25°C, pH = 6.9,  $A_{340\text{nm}}$ , Light path = 1 cm

**METHOD:** Continuous Spectrophotometric Rate Determination

### REAGENTS:

- A. 100 mM Potassium Phosphate Buffer, pH 6.9 at 25°C  
(Prepare 100 ml in deionized water using Potassium Phosphate, Monobasic, Anhydrous, Sigma Prod. No. P-5379. Adjust to pH 6.9 at 25°C with 1 M KOH.)
- B. 1 mM DL-Dithiothreitol Solution (DTT)  
(Prepare 25 ml in Reagent A using DL-Dithiothreitol, Sigma Prod. No. D-0632.)
- C. 114 mM Diacetyl Solution (Diacetyl)  
(Prepare 10 ml in deionized water using Diacetyl, Sigma Prod. No. D-3634.)
- D. 1.10 mM  $\beta$ -Nicotinamide Adenine Dinucleotide Phosphate, Reduced Form Solution ( $\beta$ -NADPH)  
(Dissolve the contents of one 1 mg vial of  $\beta$ -Nicotinamide Adenine Dinucleotide Phosphate, Tetrasodium Salt, Preweighed Vial, Sigma Stock No. 201-201 in the appropriate volume of deionized water.)
- E. (S)-Diacetyl Reductase Enzyme Solution  
(Immediately before use, prepare a solution containing 0.05 - 0.10 unit/ml of (S)-Diacetyl Reductase in cold Reagent A.)

## Enzymatic Assay of (S)-DIACETYL REDUCTASE

### PROCEDURE:

Pipette (in milliliters) the following reagents into suitable cuvettes:

	<u>Test</u>	<u>Blank</u>
Reagent B (DTT)	0.75	0.75
Reagent D ( $\beta$ -NADPH)	0.10	0.10
Reagent E (Enzyme Solution)	0.10	0.10

Mix by inversion and equilibrate to 25°C. Monitor the  $A_{340\text{nm}}$  until constant using a suitably thermostatted spectrophotometer. Then add:

Reagent C (Diacetyl)	0.10	-----
Deionized Water	-----	0.10

Immediately mix by inversion and record the decrease in  $A_{340\text{nm}}$  for approximately 5 minutes. Obtain the  $r A_{340\text{nm}}/\text{min}$  using the maximum linear rate for both the Test and Blank.

### CALCULATIONS:

$$\text{Units/ml enzyme} = \frac{(r A_{340\text{nm}}/\text{min Test} - r A_{340\text{nm}}/\text{min Blank})(1.05)(df)}{(6.22)(0.1)}$$

1.05 = Total volume (in milliliters) of assay

df = Dilution factor

6.22 Millimolar extinction coefficient of  $\beta$ -NADPH at 340 nm

0.1 = Volume (in milliliter) of enzyme used in assay

$$\text{Units/mg solid} = \frac{\text{units/ml enzyme}}{\text{mg solid/ml enzyme}}$$

$$\text{Units/mg protein} = \frac{\text{units/ml enzyme}}{\text{mg protein/ml enzyme}}$$

### UNIT DEFINITION:

One unit will convert 1  $\mu\text{mole}$  of diacetyl and  $\beta$ -NADPH to (S)-acetoin and  $\beta$ -NADP per minute at pH 6.9 at 25°C.

## Enzymatic Assay of (S)-DIACETYL REDUCTASE

### FINAL ASSAY CONCENTRATIONS:

In a 1.05 ml reaction mix, the final concentrations are 81 mM potassium phosphate, 0.7 mM DL-dithiothreitol, 1.1 mM  $\beta$ -nicotinamide adenine dinucleotide phosphate, reduced form, 11 mM diacetyl, and 0.005 - 0.01 unit (S)-diacetyl reductase.

### REFERENCE:

Heidlas, J. and Tressl, R. (1990) *European Journal of Biochemistry* **188**, 165-174

### NOTES:

1. This assay is based on the cited reference.

**This procedure is for informational purposes. For a current copy of Sigma's quality control procedure contact our Technical Service Department.**