

## **Storage & Handling PEPscreen® Custom Peptide Libraries**

### **GUIDELINES**



## Guidelines for Handling PEPscreen® Custom Peptide Libraries

The first step in working with your PEPscreen peptide set is to obtain a uniform solution of dissolved peptides, which can be aliquotted, diluted, assayed and stored. Dissolving the peptides is a critical step for successful use of a PEPscreen peptide set. This document provides guidelines for dissolving, handling and storing your peptide set.

When working with a peptide library, it is not convenient to determine a solubilization strategy for each individual peptide. It is most practical to follow a single solubilization strategy for the entire set of peptides.

### Peptide Solubilization

Unfortunately there is no universal solvent suitable for dissolving all peptide types that is compatible with all biological assays. The solubility of a particular peptide is dependent on the amino acid composition and the sequence. Sequences comprised of several charged residues (D, E, K, R, and H) are soluble in aqueous solvents. Sequences comprised of several hydrophobic residues (A, V, M, L, I, F, and W) are more soluble in organic solvents such as dimethylformamide (DMF), dimethyl sulfoxide (DMSO), or N-Methylpyrrolidone (NMP).

All peptides possess a certain level of solubility in the solvent. Depending on your application, it may not be necessary to dissolve the entire peptide sample to achieve valid results in your screening experiment. When choices are available for specific sequence characteristics, it is advisable to try solvents that are more easily removed by lyophilization, in the event the solvent does not dissolve the sample.

### Choosing the Solvent

**The following characteristics should be considered when choosing your solvent:**

1. The ability of the solvent to dissolve a variety of peptide types
2. Solvent characteristics: toxicity, chemical reactivity and stability
3. Solvent purity
4. Solvent availability and cost
5. Special handling requirements

Pure water is often the preferred choice; however there are likely hydrophobic peptides in your peptide library that will not be soluble in water. Thus, other solvents need to be considered. The preferred solvent for researchers is either DMF or DMSO because of their ability to dissolve a wide variety of peptide sequences and they are relatively benign on the peptides and the assays.

In general we recommend a solution of 0.2ml of 80% DMSO/20% water mixture (v/v) as the most useful solvent for peptide sets. However, in applications involving living cells the final concentration of DMSO or DMF must be minimized by multiple-fold dilutions due to the mild toxicity of these solvents. If the tolerance of specific biological assays to DMF or DMSO is not well-established, other suitable solvents might need to be explored.

## Solubilization Guideline

The guideline below is one example of a useful solvent mixture, acetonitrile (ACN) and water (H<sub>2</sub>O).

A good solvent for dissolving peptide sets is a 50% (v/v) ACN/ H<sub>2</sub>O mixture, because many peptides are soluble in this mixture. Another benefit of this solvent mixture is that the solvent can be completely removed from the peptide sample by lyophilization, if the peptide proves to be insoluble in the ACN/ H<sub>2</sub>O mixture. Once the sample is lyophilized, another solvent can be tried. Acetonitrile is a widely used solvent and is less toxic to living cells, provided a high purity grade is used. It is recommended that a high concentration of acetonitrile (50% ACN in H<sub>2</sub>O or greater) be used for dissolving the peptides. Most applications will require dilution in aqueous buffers to remove any solvent toxicity. A final acetonitrile level of 0.3% (v/v) or less should be safe for common cell culture systems.

## Dissolving the PEPscreen Peptide Set

The guideline below is a useful procedure for dissolving peptide sets.

1. Remove the caps from the desired tubes, in which you wish to dissolve. It is advised that you uncap the tubes individually or in 8-column rows. A “decapper” tool is included in each PEPscreen shipment to aid in this process. Add 400  $\mu$ L (or 2 X 200  $\mu$ L) of 50%(v/v) ACN/ H<sub>2</sub>O solution to each tube. For sets containing mostly hydrophobic peptide sequences, add 200  $\mu$ L of ACN, followed by 200  $\mu$ L of H<sub>2</sub>O.
2. Recap the tubes; ensuring the cap is tightly sealed. Incomplete capping can result in solvent leakage from the tubes. It is critical that the caps and tubes not be interchanged to prevent cross-contamination of the samples. If there is any doubt as to which caps belong to which tubes, we recommend you wash and dry all the caps before recapping or use new caps. Cross-contamination of peptides can lead to invalid results.
3. Return the capped tubes to the rack. Apply the rack lid and secure by locking the lid. When the lid is “locked”, invert the rack several times allowing dissolution to take place.
4. The capped tubes can be removed from the rack for inspection. Gentle tilting of the tube will expose whether or not the peptides have dissolved.
5. Continue the inspection for each tube, making note of the peptides not completely dissolved. Sonication may help in dissolving the peptides. If sonication does not completely dissolve the peptide, you may want to explore the strategies outlined in steps 6 and 7.
6. If there are several peptides that have not dissolved, add additional aliquots of the ACN/ H<sub>2</sub>O mixture and repeat the mixing and sonication steps. A volume of up to 0.7mL is practical for use in the tubes supplied. If there are still peptides that are not dissolved, follow the procedure outlined in step 7.
7. Label a 10mL polypropylene tube corresponding to each undissolved peptide. Remove the cap from the corresponding tube of undissolved peptide and transfer the content of the tube into the labeled 10mL polypropylene tube. Lyophilize the 10mL tubes to remove the solvent. DMSO is likely the most suitable solvent for hydrophobic peptides. After lyophilization, add a minimum volume of solvent (typically 100  $\mu$ L of DMSO) and dilute the solution with a 50/50 (v/v) ACN/H<sub>2</sub>O mixture to a total volume of 400  $\mu$ L. This will maintain an equivalent peptide concentration for all of the peptides in the set. Vortexing the sample may assist in dissolving the peptide. Carefully transfer the peptide solution or suspension back into its original tube, cap the tube, and position the tube in its original location in the rack.

It is important to take a sensible approach when working with peptide sets. Although some peptides may not be completely dissolved, there will likely be enough dissolved peptide to provide meaningful results in many applications, such as binding assays where an excess of peptide is typically present. Peptide solutions containing DMSO must be frozen when not in use to prevent oxidation of cysteines and methionines in the peptide sequence.

## Other Solvent Choices

Other solvent mixtures for dissolving peptides before final lyophilization are different combinations of acetic acid/acetonitrile/ water mixtures. The acetic acid and the acetonitrile concentrations should range from 5 to 20%, depending on the relative hydrophobicities of the peptides. When high concentration of acetic acid is used (>10%), the peptides should be lyophilized or frozen quickly to avoid potential hydrolysis reactions.

If using DMSO, it will be beneficial to add water to the DMSO (e.g. 20%) to assist hydration of highly hydrophilic peptides. Since DMSO is a weak oxidizing agent, peptides containing cysteine (C), methionine (M), or tryptophan (W) can be oxidized. For this reason, DMF or N-methylpyrrolidone (NMP) may be better choices for dissolving your peptide set.

## Handling Guidelines for PEPscreen Peptide Sets

The dissolved peptides constitute a set of stock peptide solutions. This peptide set should be handled with much care, avoiding the possibility of cross-contamination to the peptide set.

Making replicate sets from the stock peptide set is useful for many applications in order to minimize repetitive freeze-thaw cycles. Aliquotting can be done into microtitre plates or into a set of capped polypropylene tubes (as originally supplied). It is recommended that you uncap individual tubes or, at the most, one row of tubes at a time to reduce the possibility of cross-contamination to your stock set of peptides. The majority of applications will require dilution from the stock peptide set.

The stock and replicate sets should be stored at -20°C or colder. It is not recommended to keep peptides in a liquid state for long-term storage. Peptides in solution may degrade by chemical processes or grow microbiological contaminants resulting in an unusable sample. For long-term storage, the peptides are much more stable when stored in a lyophilized state.

If liquid state storage is unavoidable, we recommend adding 0.1%(w/v) sodium azide, which serves as a preservative and eliminates microbial growth in the solution.

## Conclusion

This document serves only as a general guideline for working with PEPscreen peptide sets. Because each peptide set is unique, it may be necessary to tailor special methods for dissolving your peptide set. If you have specific questions regarding your peptide set, please contact us at **1-800-234-5362** or email us at **peptides@sial.com**.

**We appreciate your business and wish you great success in your research!**

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
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