
Facile HPLC Separation of Peptides: Resolution of Chiral/Achiral Single Amino Acid Analogues using Teicoplanin-based Chiral Stationary Phases

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Abstract

- Recently, a new wave of peptide therapeutics are coming through pharmaceutical and biotech companies. Compared to small molecules, peptides are more potent with higher specificity, and most importantly, peptides clear from the body faster and have fewer toxicology problems.
- The separation of closely-related peptides that are different from one another by a particular amino acid, or by the chirality of an amino acid, often involves HPLC methods that are either complex or not ESI-MS compatible. This study describes the HPLC method development for resolution of small chiral peptides and single amino acid chiral/achiral analogues in peptide sequences utilizing teicoplanin-based chiral stationary phases. The mobile phases used are generally acetonitrile-buffer systems that are LC-MS compatible, enabling higher sensitivity than UV detection. More importantly, these phases provide different selectivity and higher resolution when compared to the conventional C₁₈, ion pairing and ion exchange methods.
- This presentation is aimed to give analytical chemists a tool on LC-MS platforms to test the quality and quantity of peptide products. Amino acid variation for peptides with up to 13 amino acid residues have been separated so far, but it is not known what the limit in peptide length will be. Contrary to amino acids, the elution order for peptides with chiral analogue has shown to be D before L. Examples of separation on different peptide families and several racemic di- and tri-peptides will be given. Detailed information on the method development and optimization procedures will be outlined. Finally, the complementary effect on the CHIROBIOTIC T/T2/TAG columns will also be presented.

Separation of Peptide Analogues

These are closely related peptides that differ from each other because of a particular amino acid or chirality of an amino acid either within the peptide or at its terminus

- Current LC Methods:
 - Reversed phase (C_{18}) – often needs ion pairing reagents that modify the column and are not MS compatible
 - Ion exchange – uses buffers that are generally not MS compatible
- CHIROBIOTIC Phases
 - Use ACN/ buffers that are LC-MS compatible
 - Provide different selectivity from C_{18} and ion exchange methods

REFERENCE:

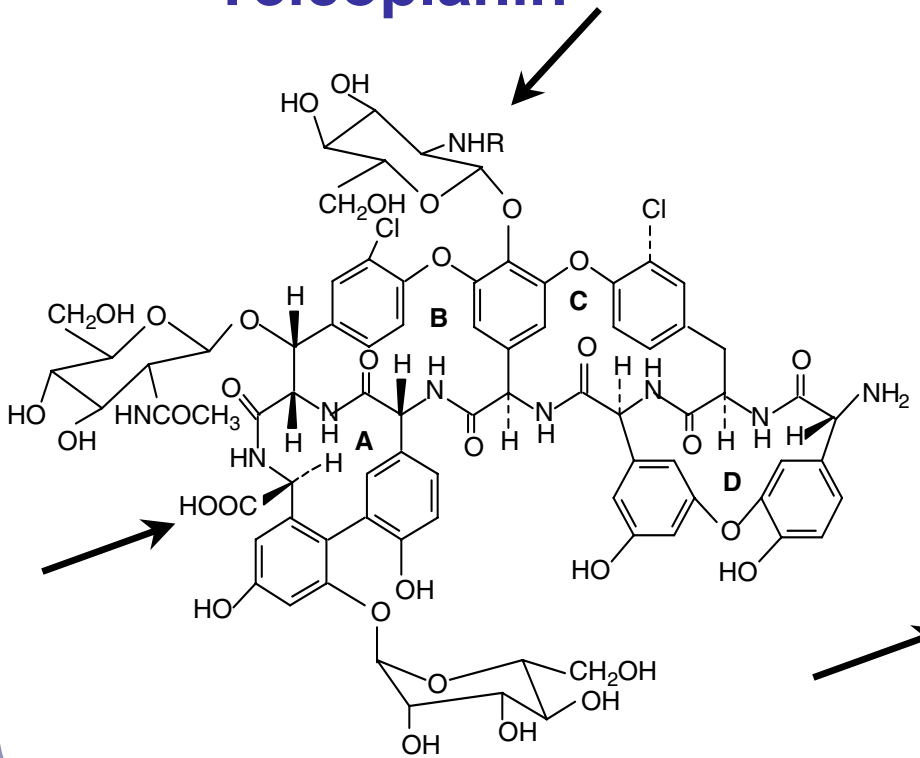
B Zhang, R Soukup & D W Armstrong; J Chrom A, 1053 (2004) 89-99

Separation of Peptide Analogues

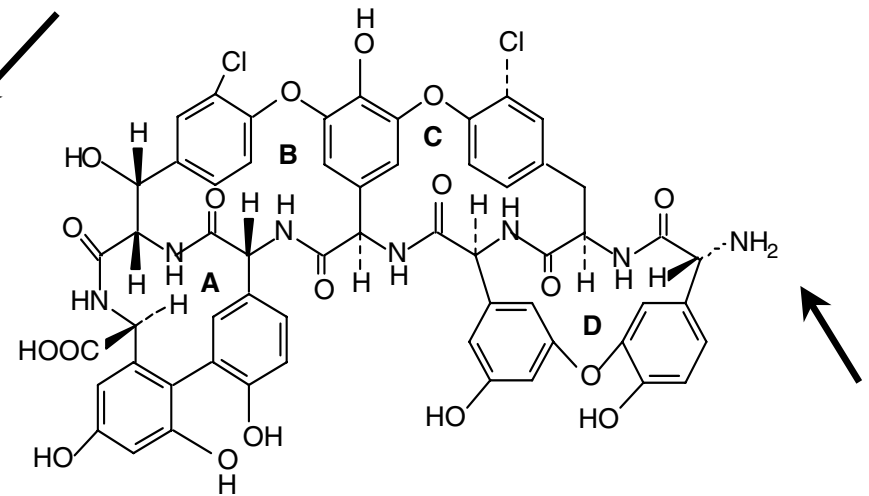
- Astec CHIROBIOTIC phases have demonstrated large selectivity factors for:
 - Single amino acid differences within peptides of up to 13 amino acid residues
 - Single chiral amino acid differences where one amino acid in the peptide is present as either the D or L form
- Maximum peptide chain length that can be separated is as yet unknown.
- D elutes before L when it is a chiral analogue.

Proposed Structures of CSPs Macrocyclic Glycopeptides

Teicoplanin



Teicoplanin Aglycone



→ Key sites

Methods Development

Peptide type	Di/Tri-peptides	Bi-functional	Neutral
Starting mobile phase: 50/50: ACN/Buffer	5mM NH ₄ OAc, pH 4.1	0.1% HCOOH	5mM NH ₄ formate
Optimization: Organic	% ACN or combination of ACN and MeOH	% ACN (U-shape effect)	% ACN (U-shape effect)
pH	2.6 – 6.8		2.6 – 6.2
Buffer	2-50mM	0.01 – 0.5%	2-50mM
Temperature	5-45°C	5-45°C	5-45°C

All peptide types showed complementary effects between CHIROBIOTIC T, T2 AND TAG CSPs

Optimization: [ACN] Effects Tri-peptide

Sample: Gly-DL-Leu-DL-Ala

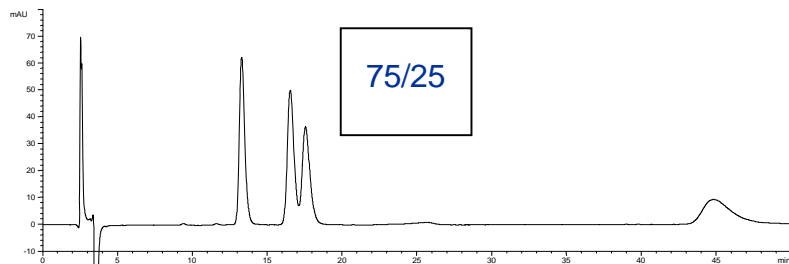
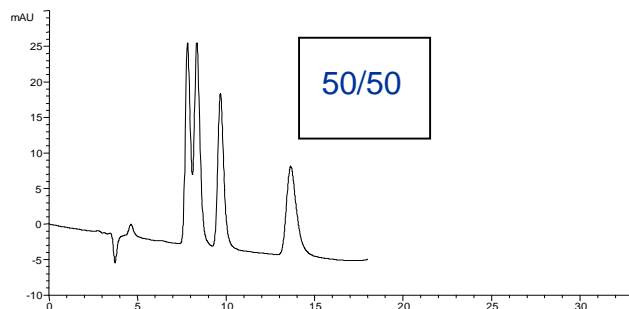
Column: CHIROBIOTIC TAG, 250x4.6mm

Mobile phase: ACN/5mM NH₄OAc, pH 4.1

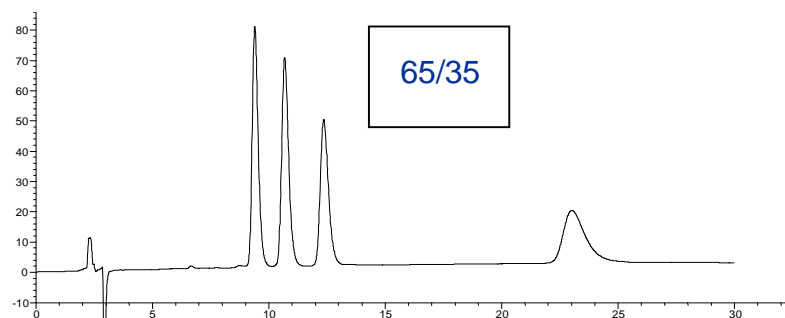
Temperature: 25°C

Flow rate: 1 mL/min

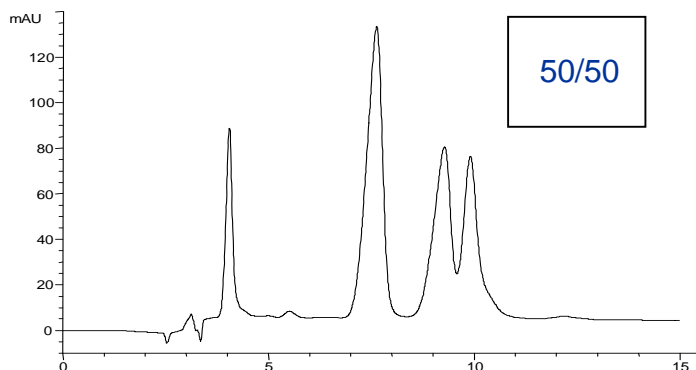
UV: 220 nm



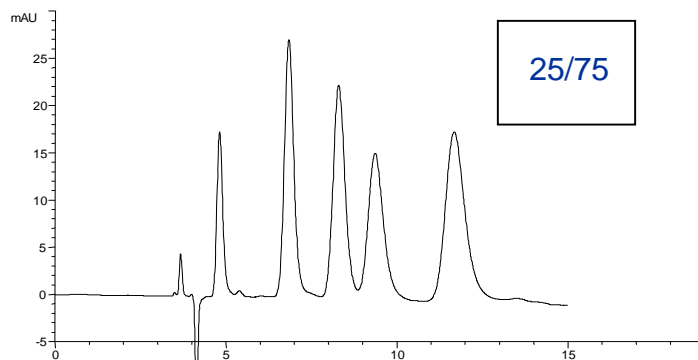
Best result



Optimization: [ACN] Effects – 5 Angiotensins II



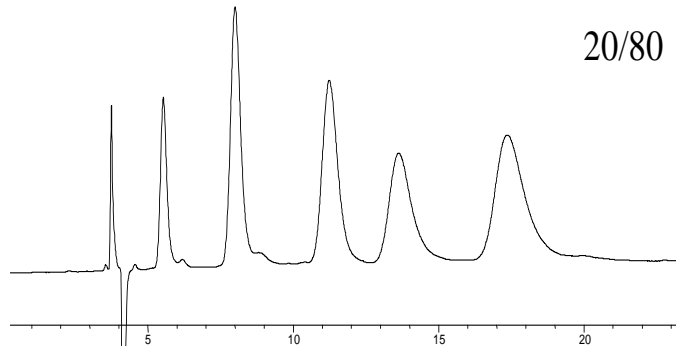
Column: **CHIROBIOTIC T2, 250x4.6mm**
Mobile phase: **ACN/0.1% HCOOH**
Temperature: **35°C**
Flow rate: **0.8 mL/min**
UV: **220 nm**



Optimization: [ACN] Effects

5 Angiotensins II

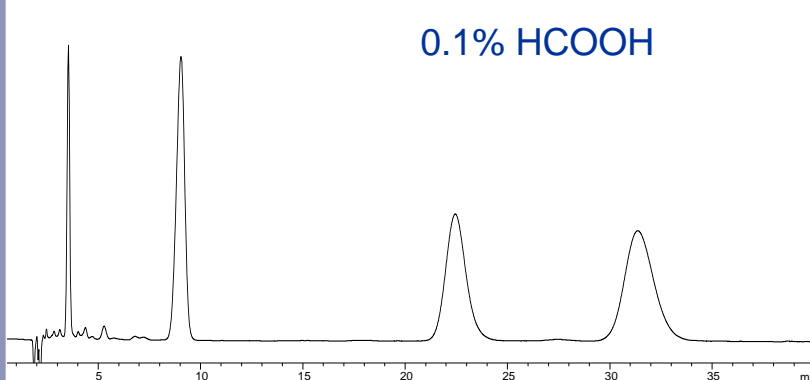
Best result



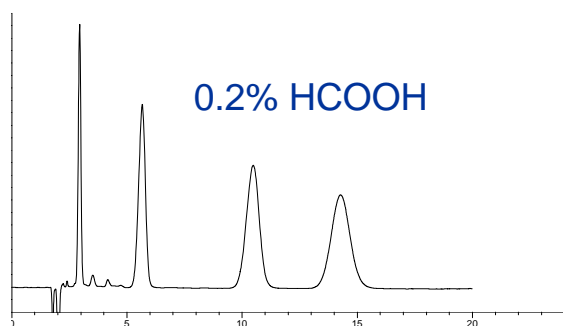
Column: CHIROBIOTIC T2, 250x4.6mm
Mobile phase: 20/80: ACN/0.1% HCOOH
Flow rate: 0.8 mL/min
Temperature: 35°C
UV: 220 nm

Peak 1: 5.53 min Glu-Gly-Val-Tyr-Val-His-Pro-Val
Peak 2: 8.00 min Asp-Arg-Val-Tyr-Val-His-Pro-Phe
Peak 3: 11.24 min Sar-Arg-Val-Tyr-Ile-His-Pro-Thr
Peak 4: 13.63 min Asp-Arg-Val-Tyr-Ile-His-Pro-Phe
Peak 5: 17.36 min Sar-Arg-Val-Tyr-Ile-His-Pro-Phe

Optimization: [Buffer] Effects 4 Bradykinins



Column: **CHIROBIOTIC TAG, 250x4.6mm**
Mobile phase: **50/50: ACN/HCOOH**
Flow rate: **1.5 mL/min**
Temperature: **5°C**
UV: **220 nm**

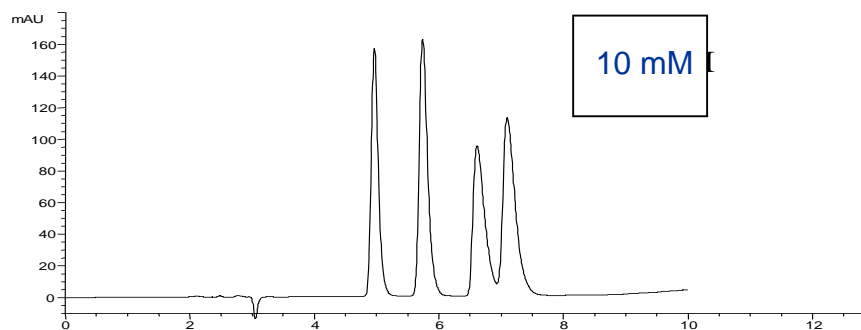
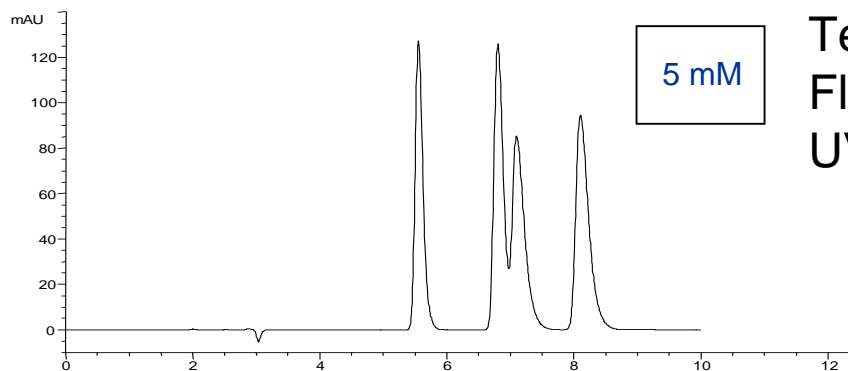


- Peak 1:** 2.95min,
Pro-Pro-Gly-Phe-Ser-Pro
- Peak 2:** 5.67 min,
Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg
- Peak 3:** 10.48 min,
Arg-Pro-Gly-Phe-Ser-Pro-Phe-Arg
- Peak 4:** 14.28 min,
Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg

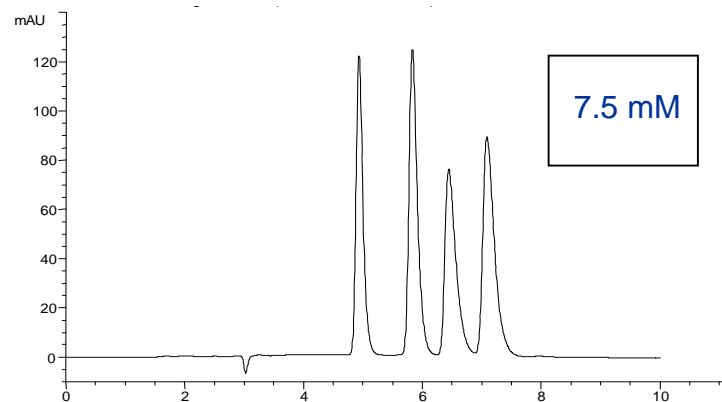
Optimization: [Buffer] Effects Di-peptide

Sample: DL-Ala-DL-Val

Column: CHIROBIOTIC T, 250x4.6mm
Mobile phase: 50/50: ACN/NH₄OAc, pH 4.1
Temperature: 25°C
Flow rate: 1 mL/min
UV: 220 nm



Best result



Optimization: Temperature Effects

Di-peptide

Sample: DL-Ala-DL-Val

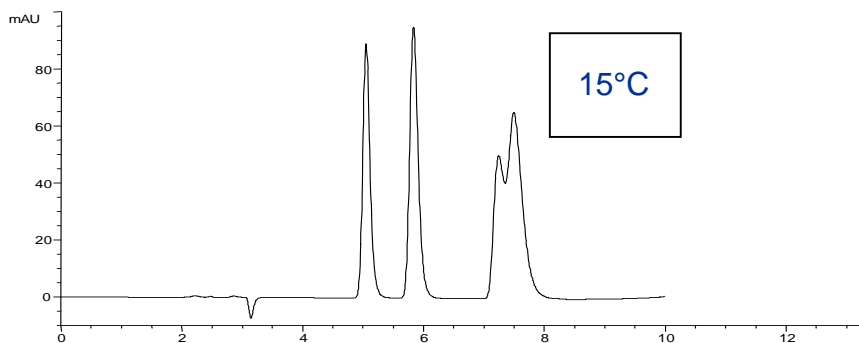
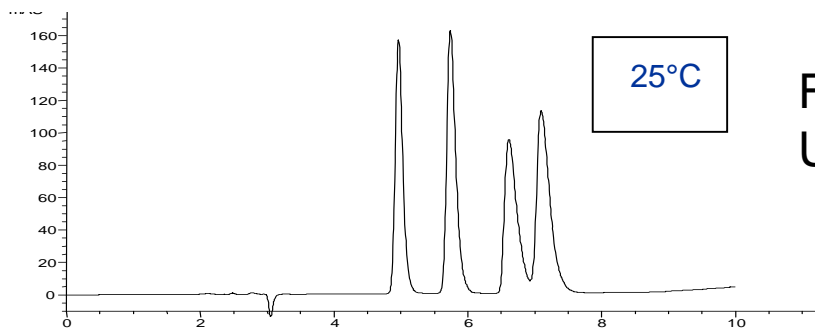
Column: CHIROBIOTIC T, 250x4.6mm

Mobile phase: 50/50: ACN/10mM

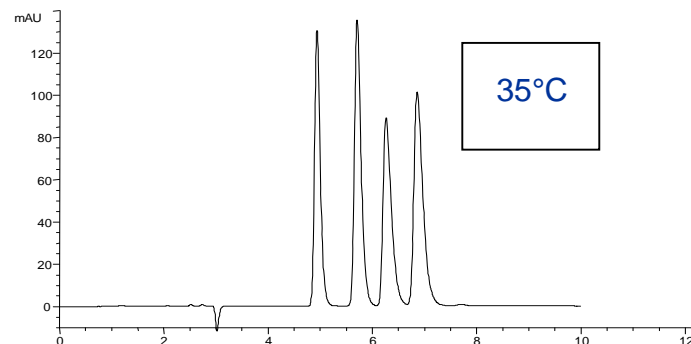
NH₄OAc, pH 4.1

Flow rate: 1mL/min

UV: 220 nm

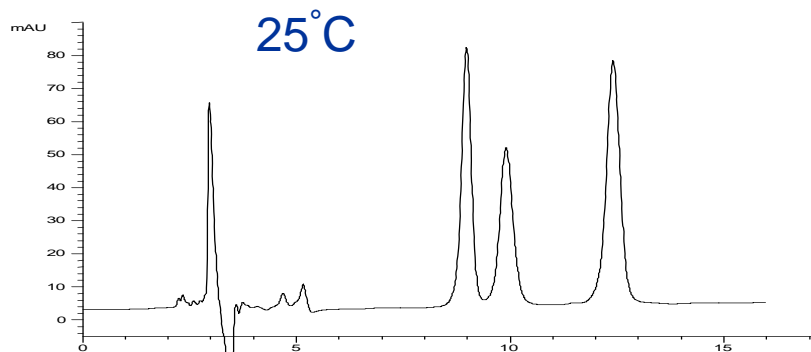


Best result

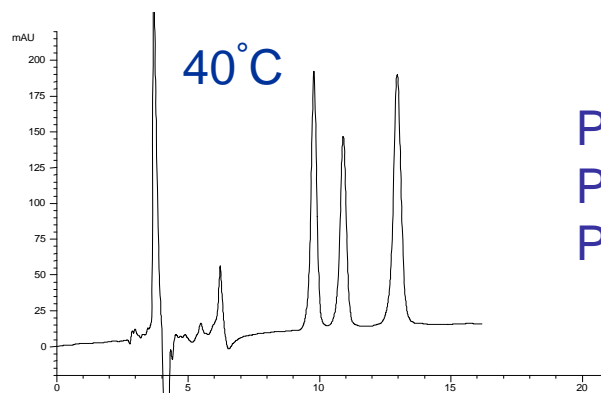


Optimization: Temperature Effects

3 β -Casomorphins



Column: CHIROBIOTIC T, 250x4.6mm
Mobile phase: 75/25: ACN/20mM
NH₄Formate
Flow rate: 1 mL/min
UV: 230 nm

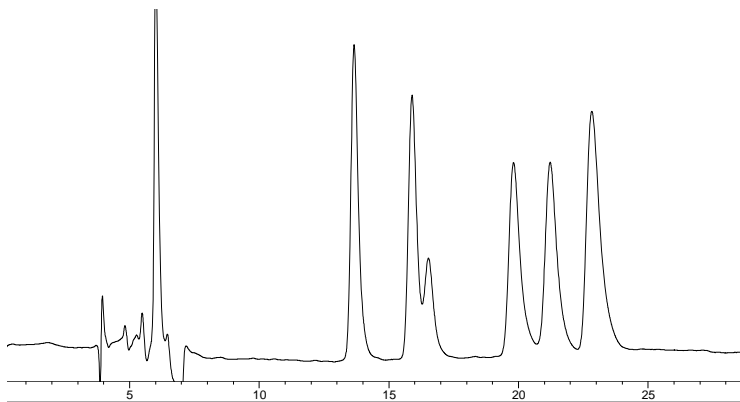


Peak 1: 8.98 min Tyr-D-Ala-Phe-**D-Ala**-Tyr-NH₂
Peak 2: 9.91 min Tyr-D-Ala-Phe-**D-Pro**-Tyr-NH₂
Peak 3: 12.41 min Tyr-D-Ala-Phe-**Hyp**-Tyr-NH₂

Complementary Effect: T/T2

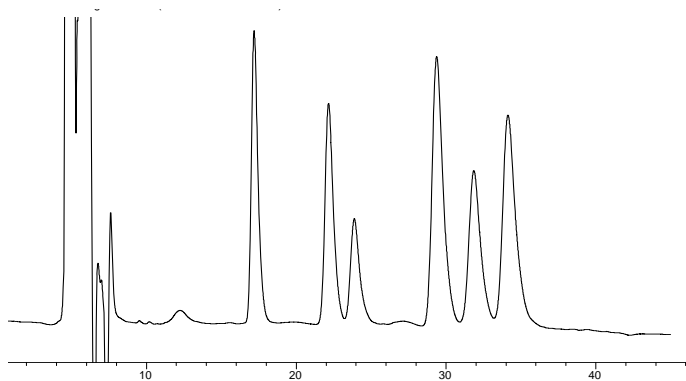
6 Enkephalins

CHIROBIOTIC T, 250x4.6mm



Mobile phase: 75/25: ACN/ 5mM
NH₄Formate, pH 3.3
Temperature: 25°C
Flow rate: 0.5mL/min
UV: 220nm

CHIROBIOTIC T2, 250x4.6mm

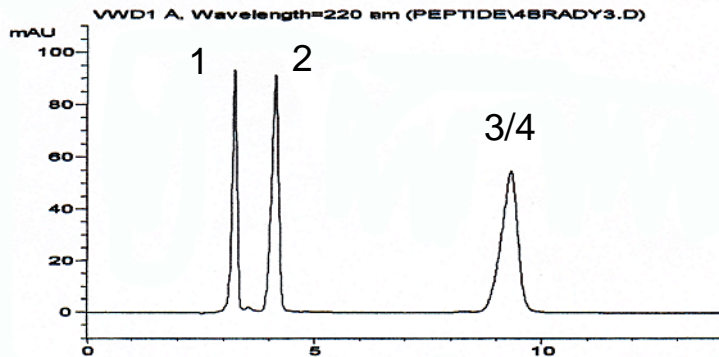


Best result

Column:
CHIROBIOTIC T2, 250x4.6mm

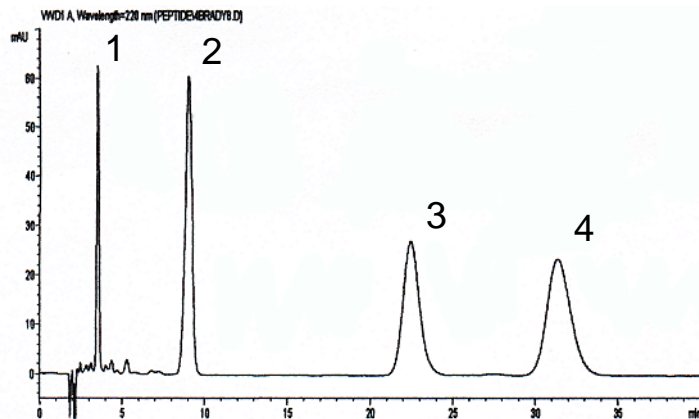
Complementary Effect: T2/TAG 4 Bradykinins

CHIROBIOTIC T2, 250x4.6mm



Mobile phase: 40/60: ACN/0.1% HCOOH
Flow rate: 1.5 mL/min
Temperature: 35°C
UV: 220 nm

CHIROBIOTIC TAG, 250x4.6mm

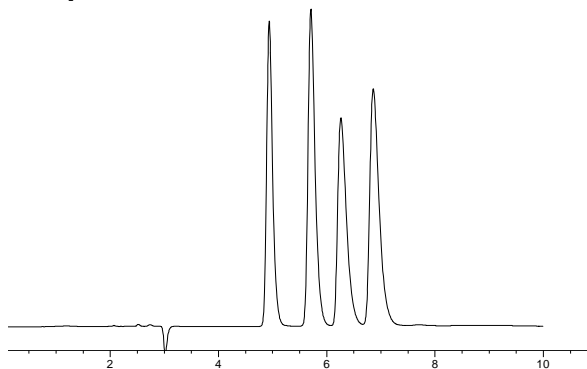


Best result

Column:
CHIROBIOTIC TAG, 250x4.6mm

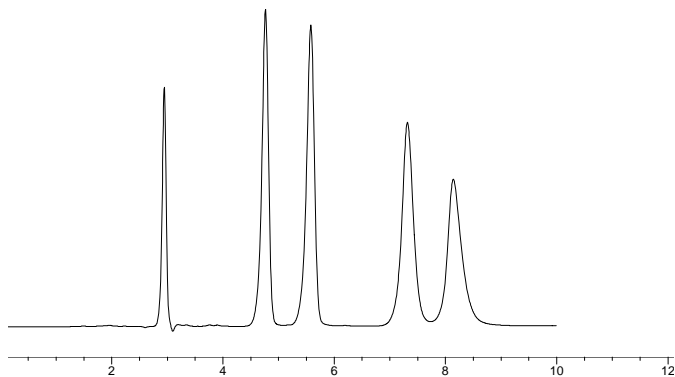
Best Conditions: Di-peptides

Sample: DL-Ala-DL-Val



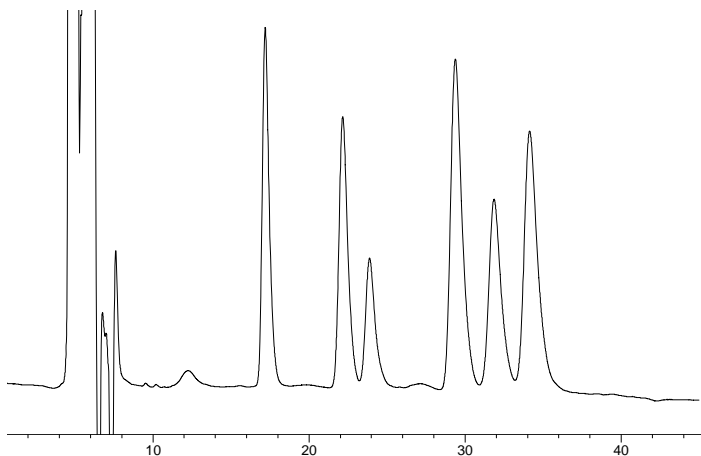
Column: **CHIROBIOTIC T, 250x4.6mm**
Mobile phase: 50/50: ACN/10mM
NH₄OAc, pH 4.1
Temperature: 35°C
Flow rate: 1 mL/min
UV: 220 nm

Sample: DL-Ala-DL-Phe



Column: **CHIROBIOTIC T, 250x4.6mm**
Mobile phase: 50/50: ACN/5mM
NH₄OAc, pH 4.1
Temperature: 25°C
Flow rate: 1 mL/min
UV: 220 nm

Application: 6 Enkephalins



Column: **CHIROBIOTIC T2, 250x4.6mm**
Mobile phase: 75/25: ACN/5mM
NH₄Formate, pH 3.3
Temperature: 25°C,
Flow rate: 0.5 mL/min
UV: 220 nm

- **Single Amino Acid Analogues:**

- Peak 2 and 3
- Peak 4 and 5
- Peak 5 and 6

- **Chiral Amino Acid Analogues:**

- Peak 1 and 3
- Peak 3 and 6

Peak 1: 17.06 min Tyr-D-Ala-Gly-Phe-D-Leu

Peak 2: 21.96 min Tyr-D-Ala-Gly-Phe-Met

Peak 3: 23.59 min Tyr-D-Ala-Gly-Phe-Leu

Peak 4: 29.10 min Tyr-Gly-Gly-Phe-Met

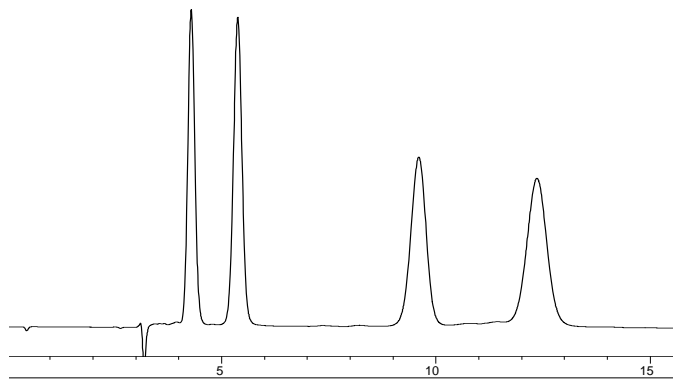
Peak 5: 31.47 min Tyr-Gly-Gly-Phe-Leu

Peak 6: 33.74 min Tyr-Ala-Gly-Phe-Leu

Application.

Luteinising Hormone-Releasing Hormone

Sample: LHRH

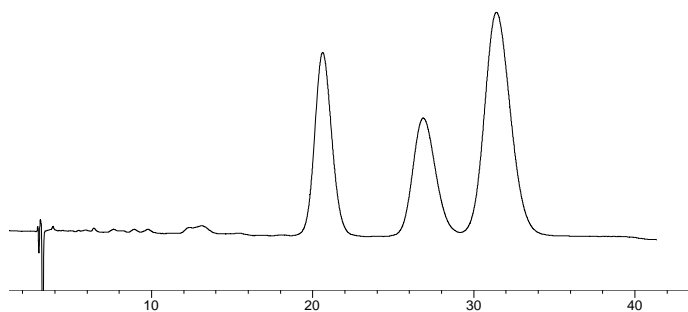


Column: CHIROBIOTIC T2, 250x4.6mm
Mobile phase: 40/60: ACN/0.1% HCOOH
Flow rate: 1.0 mL/min
Temperature: 35°C
UV: 220 nm

Peak 1: 4.29 min pGlu-**D-Phe**-Trp-Ser-Tyr-D-Ala-Leu-Arg-Pro-Gly-NH₂
Peak 2: 5.38 min pGlu-His-Trp-Ser-Tyr-**D-Ala**-Leu-Arg-Pro-Gly-NH₂
Peak 3: 9.60 min pGlu-His-Trp-Ser-Tyr-**D-Lys**-Leu-Arg-Pro-Gly-NH₂
Peak 4: 12.36 min His-Trp-Ser-Tyr-**Gly**-Leu-Arg-Pro-Gly-NH₂

Application:

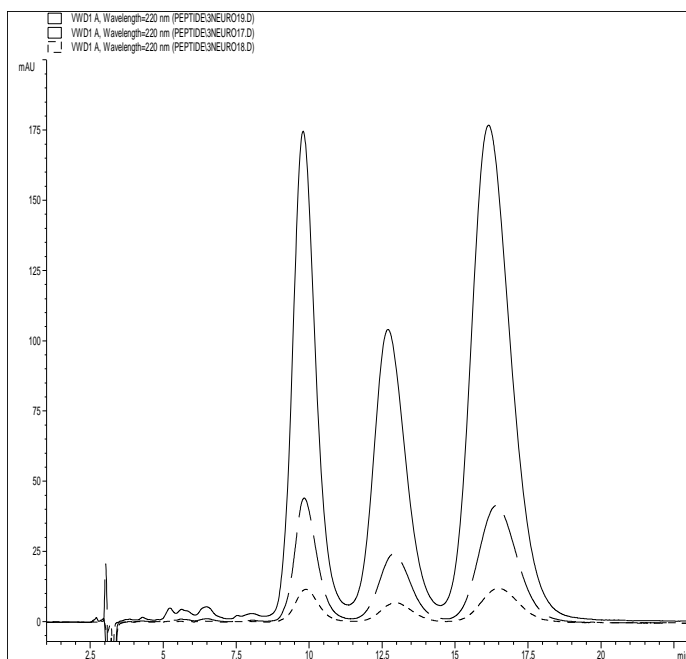
3 Neurotensins (13 AA residues)



Column: CHIROBIOTIC T2, 250x4.6mm
Mobile phase: 25/75: ACN/0.1% HCOOH
Flow rate: 1.0 mL/min
Temperature: 35°C
UV: 220 nm

Peak 1: 20.63 min; pGlu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-**Tyr**-Ile-Leu
Peak 2: 26.86 min; pGlu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-**Phe**-Ile-Leu
Peak 3: 31.41 min; pGlu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-**D-Trp**-Ile-Leu

No Overloading Issue ex: 3 Neurotensins

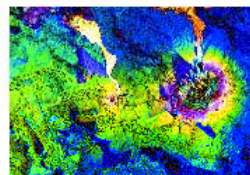
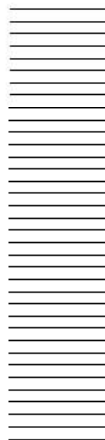
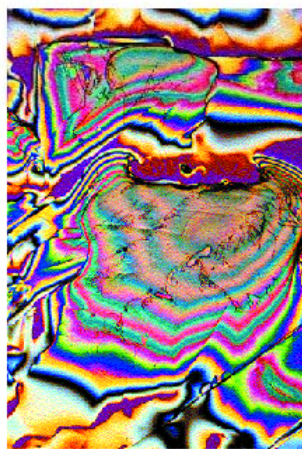
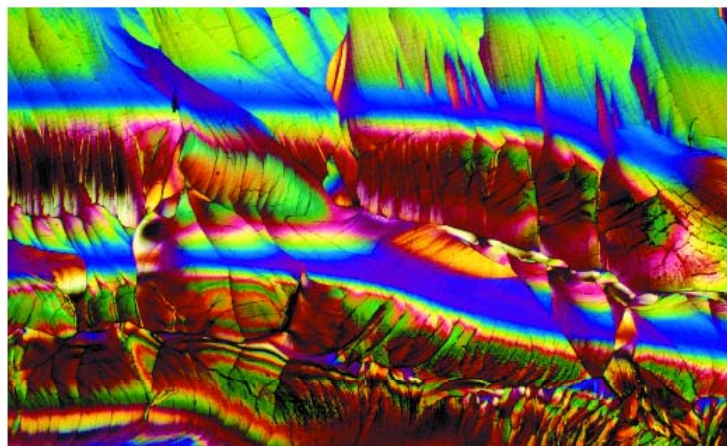


Column: **CHIROBIOTIC T2, 250x4.6mm**
Mobile phase: 20/80: ACN/0.2% HCOOH
Temperature: 35°C
Flow rate: 1mL/min
UV: 220nm
Load: Each peptide contains:
— 2µg
- - - 0.5µg
..... 0.14µg

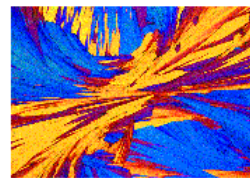
CONCLUSIONS

- Using ACN/buffer as the mobile phase, the retention of peptides was usually found to be the shortest when the % ACN was in the 40-60% range.
- Higher buffer concentrations were found to correlate with lower retentions.
- Some elution order changes were observed between CHIROBIOTIC T and T2.
- Mobile phase compositions are LC-MS compatible utilizing APCI or ESI mode.
- The methods reported are flexible in composition and operating conditions allowing an orthogonal platform to be developed, if necessary.
- Sample capacity is higher than traditional C₁₈ phases.
- Peptide families with up to 13 amino acid units have been successfully separated for sequence variations/deletions, even chiral analogues.
- Complementary effects among CHIROBIOTIC T/T2/TAG.

AMINO ACID and PEPTIDE CHIRAL SEPARATIONS HANDBOOK



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