



## Product Information

### ANTI-FLAG<sup>®</sup> M2 Monoclonal Antibody

Product Code **F 3165**  
Storage Temperature  $-20\text{ }^{\circ}\text{C}$

#### Product Description

ANTI-FLAG<sup>®</sup> M2 monoclonal antibody is a purified IgG1 monoclonal antibody, isolated from murine ascites fluid, that binds to FLAG fusion proteins.<sup>1</sup> Unlike ANTI-FLAG M1 antibody, the M2 antibody will recognize the FLAG sequence at the N-terminus, Met-N-terminus, C-terminus, or at an internal site of FLAG fusion proteins. ANTI-FLAG M2 monoclonal antibody is useful for identification and capture of FLAG fusion proteins by common immunological procedures such as Western blots and immunoprecipitation. It is also useful for affinity purification of FLAG fusion proteins when bound to a solid support.

M2 binding is not calcium dependent.

ANTI-FLAG M2 antibody is supplied in 10 mM sodium phosphate, 150 mM NaCl, pH 7.4, containing 0.02% sodium azide.

#### Storage

Store the undiluted antibody at  $-20\text{ }^{\circ}\text{C}$  in working aliquots. Repeated freezing and thawing is not recommended.

Note: Over time, small amounts of purified antibodies can precipitate from solution due to intermolecular hydrophobic interactions. If a precipitate is observed in this product, briefly centrifuge the vial to pellet the precipitate. Withdraw the desired volume of antibody solution from the clear supernatant for use. This should not alter the performance of the purified antibody in Western blot or immunoprecipitation applications.

#### Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

#### Preparation Instructions

Dilute the ANTI-FLAG M2 monoclonal antibody solution from 0.5 to 10  $\mu\text{g/ml}$  in Tris buffered saline (50 mM Tris, 0.138 M NaCl, 2.7 mM KCl, pH 8.0) (TBS) with 3% nonfat dry milk (Product Code T 8793). Adjust the antibody concentration to maximize detection sensitivity and to minimize background.

#### Procedure

Improved Western Blot Method for Detecting FLAG Fusion Proteins using ANTI-FLAG M2 Antibody.

1. Separate FLAG-tagged proteins from sample lysates using a standard sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) protocol. Load 2.5 to 10  $\mu\text{g}$  of total lysate protein per lane.
2. Transfer proteins from the gel to an Immobilon<sup>™</sup>-P or other polyvinylidene difluoride (PVDF) membrane. Nitrocellulose membranes can be used, but typically result in less sensitivity.
3. Wash the blot in at least 0.5  $\text{ml/cm}^2$  of Milli-Q<sup>™</sup> water for 2-3 minutes with mild agitation.
4. Block the blot with at least 0.5  $\text{ml/cm}^2$  of TBS with 3% nonfat dry milk (Product Code T 8793 or 50 mM Tris, 0.138 M NaCl, 2.7 mM KCl, pH 8.0, containing 30 mg/ml nonfat dry milk) for 30 minutes at room temperature with agitation (about 50-60 rpm).
5. Remove the blocking agent and wash once with 0.5  $\text{ml/cm}^2$  of TBS (Product No. T 6664).
6. Add ANTI-FLAG M2 antibody to a final concentration of 10  $\mu\text{g/ml}$  to the blot in at least 0.5  $\text{ml/cm}^2$  of TBS with 3% nonfat dry milk and incubate at room temperature for 30 minutes.  
Note: Using less M2 ANTI-FLAG antibody may help to reduce background and cross-reactivity. See the Troubleshooting Guide.
7. Remove the ANTI-FLAG M2 antibody solution and wash once with at least 0.5  $\text{ml/cm}^2$  of TBS.

8. Add rabbit anti-mouse IgG, peroxidase conjugate (Product No. A 9044 or equivalent) to at least 0.5 ml/cm<sup>2</sup> of TBS with 3% nonfat dry milk. Use the concentrations listed in the table below. These concentrations are recommended starting concentrations for the antibodies used in the Western blot. Incubate the blots with shaking at room temperature for 30 minutes.

ANTI-FLAG M2 primary antibody (µg/ml)	Substrate	Secondary antibody concentration
0.5–10	ECL+™	1:80,000
0.5–10	ECL™	1:10,000

9. Wash the blot eight times for a total of 20 minutes in 50 mM Tris, 0.138 M NaCl, 2.7 mM KCl, pH 8.0, plus 0.05% TWEEN® 20 (TBS- TWEEN 20, Product Code T 9039).
10. Develop the blots with the appropriate substrate for 5 minutes.
11. Expose BioMax™ light film to the blot. Exposure times range from 30 seconds to 10 minutes. It is best to do a quick exposure of 10 to 30 seconds to determine what exposure time is needed. If the signal is too intense even at the short exposure times let the signal decay from 1 to 8 hours or longer if necessary and then re-expose the film.

#### Immunofluorescence

ANTI-FLAG M2 may be used in immunofluorescent procedures. A typical concentration for use is 20 µg/ml.<sup>2</sup>

#### **Product Profile**

Antigenic binding site:

N-Asp-Tyr-Lys-Asp-Asp-Asp-Lys-C

Specificity: ANTI-FLAG M2 monoclonal antibody detects a single band of protein on a Western blot from an *E. coli* crude cell lysate.

Sensitivity: ANTI-FLAG M2 monoclonal antibody detects 2 ng of FLAG-BAP fusion protein on a dot blot using chemiluminescent detection. In order to obtain best results, it is recommended that each individual user determine working dilution by titration assay.

#### **References**

1. Brizzard, B.L., *et al.*, Immunoaffinity purification of FLAG epitope-tagged bacterial alkaline phosphatase using a novel monoclonal antibody and peptide elution. *BioTechniques*, **16**, 730-735 (1994)
2. Ciaccia, A.V., and Price, E.M., IBI FLAG Epitope, **1**, 4-5 (1992)
3. Bjerrum, O.J., and Heegaard. N.H.H., CRC Handbook of Immunoblotting of Proteins, Volume I, Technical Descriptions, CRC Press, (1988) p. 229-236
4. Dunbar, B.S. (ed.) Protein Blotting: A Practical Approach, IRL Press, NY, p. 67-70 (1994)
5. Fortin, A., *et al.*, A 56- to 54-kilodalton *non grata* signal in immunoblot analysis using the horseradish peroxidase chemiluminescence system. *Biochem. Cell Biol.*, **72**, 239-243 (1994)
6. Harlow, E., and Lane, D., Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988, Product Code A2926
7. Pampori, N.A., *et al.*, Dilution of the chemiluminescence reagents reduces the background noise on western blots. *BioTechniques*, **18**, 589-590 (1995)

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## Troubleshooting Guide

Problem	Possible Cause	Solution
Fusion protein not detected	Protein not expressed	Verify nucleic acid sequence of FLAG in vector construct. If sequence is present, attempt to optimize expression.
	Target protein poorly represented in sample	Positive controls should always be included. If the positive control works, the sample may not contain the FLAG fusion protein of interest or it may be present at concentrations too low to detect. Immunoprecipitation with ANTI-FLAG M2 Affinity Gel (Product Code A 1205) may be required for low FLAG fusion protein concentrations. Positive controls available from Sigma: <ul style="list-style-type: none"> <li>• Amino-Terminal FLAG-BAP fusion protein, Product Code P 7582</li> <li>• Carboxy-Terminal FLAG-BAP fusion protein, Product Code P 7457</li> <li>• Amino-Terminal Met-FLAG-BAP fusion protein, Product Code P 5975</li> </ul>
	Defective detection reagents	Run appropriate controls to ensure performance. Use 10 ng/lane of a control FLAG-BAP-fusion protein as a positive control. If no signal is obtained with the control, repeat the procedure using a newer lot of antibody-HRP conjugate and freshly prepared reagents.
	Inadequate exposure time using chemiluminescent system	If no signal is seen, expose for longer times. Sigma recommends trying 30 second to 10 minute exposure times.
	Inappropriate film	Switch to film designated for chemiluminescent detection such as Kodak BioMax Light.
	No target protein present on membrane	Verify transfer by visualizing proteins on the membrane using a Ponceau S solution (Product Code P 7170). If possible, a positive control should always be run to insure components are functioning. Prestained protein markers (e.g. Product Code C3 437 and P 1677) may also be used to verify complete transfer.
	Antigen is covered by blocking reagent due to overblocking	Masking of a signal can occur if the blocking reagent (such as casein or gelatin blocking buffers, Product Nos. C 7594 or G 7663, respectively) is used at too high a concentration. A dilution of 1:1 to 1:3 may be done to decrease the concentration. If the problem persists, use TBS with 3% non-fat dry milk (Product No T 8793).
Antibody concentration not optimal	Determine optimal working dilution for ANTI-FLAG antibody by titration. Consider using more antibody if no signal or weak signal is detected. Also, antibody used at too high a concentration can also cause inhibition of signal especially in chemiluminescent detection systems.	
Cross-reactivity	Cellular extract concentration is too high	2.5 to 10 µg per lane of total lysate protein is usually enough to obtain a good signal. Load less cellular extract or serially dilute the cell extract to obtain the optimal signal to noise ratio.
	ANTI-FLAG M2 antibody concentration is too high	Dilute ANTI-FLAG M2 antibody from 0.1 to 0.5 µg/ml. Use TBS with 3% non-fat dry milk as diluent.
	Secondary antibody concentration is too high	Sigma recommends initial dilutions of 1:10,000 for ECL and 1:80,000 for ECL+. Further dilutions may be necessary.
	ANTI-FLAG M2 antibody cross-reacts with naturally occurring FLAG-like epitopes	Increasing the temperature to 37 °C during the blocking, binding and wash steps may reduce cross-reactivity. Lysates from mock-transfected controls (transfected with plasmid without insert DNA) will help distinguish the FLAG-fusion proteins from other cross-reacting proteins.

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