



## Product Information

### PRECAST AGAROSE GELS

Sigma Product Nos. **P 5472, P 5597, P 5722, P 5847, P 6097, P 5972**

### PRODUCT DESCRIPTION

Precast agarose gels are cast using a 1X TBE (0.089 M Tris borate, pH approx. 8.3, containing 2 mM EDTA) buffer system. Ethidium bromide (0.5 µg/ml) is included in the gel for easy visualization. The gels are suitable for separating nucleic acids, giving sharp DNA bands and low background fluorescence. DNase activity and DNA binding are not detected.

Agarose Concentration	8 Well	20 well	24 well
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1.0%	P 5472	P 5722	P 5972
4.0%	P 5597	P 5847	P 6097

### GEL DESCRIPTIONS

Tray dimension: 6.8 cm x 10.2 cm  
 Gel dimension: 6.0 cm x 9.5 cm  
 Gel thickness: 5.5 mm  
 Sample format: Gels are cast with 8, 20 or 24 wells that will each accommodate up to 15 µl sample volume.

**STORAGE:** Store flat at room temperature (18 °C – 25 °C). **DO NOT FREEZE.** Limit exposure to light.

**PRECAUTIONS:** Ethidium bromide is a mutagen. Use gloves when handling gels containing the dye. Avoid skin and eye exposure to UV light. Refer to Material Safety Data Sheet.

### COMPATIBILITY:

Unit manufacturer	8- and 24-well portrait formats
Bio-Rad Mini-Sub®	YES
Hoefer Minnie the Gel-Cicle®	YES
Life Technologies Horizon® 58	YES

Unit manufacturer	20-well landscape formats
IBI Model QSH	YES
Mupid®	YES

### DYE MIGRATION

Agarose Type and Concentration	Bromophenol Blue	Xylene Cyanol
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<b>1X TBE Buffer</b>		
1.0% Agarose	400 bp	4,100 bp
4.0% Agarose	20 bp	250 bp

### INSTRUCTIONS FOR USE:

Sigma's Precast Agarose mini-gels require less than 5 minutes to set up.

1. Peel the paper backing from the adhesive strips on the bottom of the tray.
2. Peel off the lid. **Leave the gel in the tray.**
3. Press the tray directly onto the chamber platform. Align the wells so the DNA samples will run straight.
4. Pour 1X TBE electrophoresis buffer in the chamber to a **depth of 5 mm** OVER the flange of the tray.
5. Load the DNA sample (≤15 µl volume).
6. Electrophorese the gels at no more than 10 V/cm for 30 minutes. Lower voltages for longer times are acceptable.
7. For DNA fragments ≥5 kb, use 1 to 5 V/cm and increase the run time.
8. Remove the gel from the tray to photograph/document and/or destain.

12/02