

## Product Information

### Lectin from Concanavalin A (*Canavalia ensiformis*) Peroxidase Labeled

Product Number **L6397**  
Storage Temperature 0 °C

#### Product Description

Concanavalin A (Con A) is reported to have several isoelectric points possibly corresponding to different isoforms. The pI values are reported as 4.5, 4.7, 5.05, and 5.5.<sup>1</sup>

Lectins are proteins or glycoproteins of non-immune origin that agglutinate cells and/or precipitate complex carbohydrates. Lectins are capable of binding glycoproteins even in presence of various detergents.<sup>2</sup> The agglutination activity of these highly specific carbohydrate-binding molecules is usually inhibited by a simple monosaccharide, but for some lectins, di, tri, and even polysaccharides are required.

Lectins are isolated from a wide variety of natural sources, including seeds, plant roots and bark, fungi, bacteria, seaweed and sponges, mollusks, fish eggs, body fluids of invertebrates and lower vertebrates, and from mammalian cell membranes. The precise physiological role of lectins in nature is still unknown, but they have proved to be very valuable in a wide variety of applications *in vitro*, including:

1. blood grouping and erythrocyte polyagglutination studies.
2. mitogenic stimulation of lymphocytes.
3. lymphocyte subpopulation studies.
4. fractionation of cells and other particles.
5. histochemical studies of normal and pathological conditions.

Sigma offers a range of lectins suitable for the above applications. Most Sigma lectins are highly purified by affinity chromatography, but some are offered as purified or partially purified lectins, suitable for specific applications.

Many of the lectins are available conjugated to (conjugation does not alter the specificity of the lectin):

1. fluorochromes (for detection by fluorimetry).
2. enzymes (for enzyme-linked assays).
3. insoluble matrices (for use as affinity media).

Please refer to the table for general information on the most common lectins.

This product is labeled with horseradish peroxidase (Product No. P8375). The peroxidase label allows use of this lectin in blotting procedures for the identification of sugar side-chains on proteins.

#### Procedure

A general procedure for probing sugar side chains on immobilized proteins is as follows:

1. Proteins are first separated by SDS-PAGE and transferred to nitrocellulose.
2. Excess binding sites are blocked by incubation in PBS containing 2% (v/v) TWEEN<sup>®</sup> 20 for 2 minutes at 20 °C.
3. Rinse the blot twice in PBS.
4. Incubate with 1 to 5 µg/ml of lectin-peroxidase in PBS containing 0.05% (v/v) TWEEN 20, with 1 mM CaCl<sub>2</sub>, 1 mM MnCl<sub>2</sub>, and 1 mM MgCl<sub>2</sub> for 16 hours at 20 °C.
5. Remove surplus lectin by rinsing in PBS.
6. Peroxidase activity can be detected using standard HRP substrates.

#### Precautions and Disclaimer

For Laboratory Use Only. Not for drug, household or other uses.

#### Preparation Instructions

The product is soluble in water (1mg/ml), yielding a hazy to clear, colorless to red or tan solution.

#### Storage/Stability

Aggregation is thought to occur in the presence of high concentrations of 2-mercaptoethanol.

Solutions of Concanavalin A are thought to be completely denatured after heating at 80 °C for five minutes.<sup>3</sup>

Lectin	MW (kDa)	Subunits	Specificity		Mitogenic Activity
			Blood Group	Sugar	
<i>Abrus precatorius</i>			–		+
Agglutinin	134	4		gal	
Abrin A (toxin)	60	2		gal	
Abrin B (toxin)	63.8	2( $\alpha\beta$ )		gal	
<i>Agarius bisporus</i>	58.5	–	–	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	
<i>Anguilla anguilla</i>	40	2	H	$\alpha$ -L-Fuc	
<i>Arachis hypogaea</i>	120	4	T	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	
<i>Artocarpus integrifolia</i>	42	4	T	$\alpha$ -gal $\rightarrow$ OME	+
<i>Bandeiraea simplicifolia</i>					
BS-I	114	4	A, B	$\alpha$ -gal, $\alpha$ -galNAc	
BS-I-A <sub>4</sub>	114	4	A	$\alpha$ -galNAc	
BS-I-B <sub>4</sub>	114	4	B	$\alpha$ -gal	
BS-II	113	4	acq, B, Tk, T	glcNAc	
<i>Bauhinia purpurea</i>	195	4	–	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	+
<i>Caragana arborescens</i>	60; 120 <sup>a</sup>	2/4	–	galNAc	
<i>Cicer arietinum</i>	44	2	–	fetuin	
<i>Codium fragile</i>	60	4	–	galNAc	
<i>Concanavalin A</i>	102	4	–	$\alpha$ -man, $\alpha$ -glc	+
<i>Succinyl-Concanavalin A</i>	51	2	–	$\alpha$ -man, $\alpha$ -glc	+ <sup>b</sup>
<i>Cytisus scoparius</i>	–	–	–	galNAc, gal	
<i>Datura stramonium</i>	86	2( $\alpha\beta$ )	–	(glcNAc) <sub>2</sub>	
<i>Dolichos biflorus</i>	140	4	A <sub>1</sub>	$\alpha$ -galNAc	
<i>Erythrina corallodendron</i>	60	2	–	$\beta$ -gal(1 $\rightarrow$ 4)glcNAc	+
<i>Erythrina cristagalli</i>	56.8	2( $\alpha\beta$ )	–	$\beta$ -gal(1 $\rightarrow$ 4)glcNAc	
<i>Euonymus europaeus</i>	166	4( $\alpha\beta$ )	B, H	$\alpha$ -gal(1 $\rightarrow$ 3)gal	+
<i>Galanthus nivalis</i>	52	4	(h)	non-reduc. $\alpha$ -man	
<i>Glycine max</i>	110	4	–	galNAc	+ <sup>c</sup>
<i>Helix aspersa</i>	79	–	A	galNAc	
<i>Helix pomatia</i>	79	6	A	galNAc	
<i>Lathyrus odoratus</i>	40-43	4( $\alpha\beta$ )	–	$\alpha$ -man	+
<i>Lens culinaris</i>	49	2	–	$\alpha$ -man	+
<i>Limulus polyphemus</i>	400	18	–	NeuNAc	
Bacterial agglutinin	–	–	–	galNAc, glcNAc	
<i>Lycopersicon esculentum</i>	71	–	–	(glcNAc) <sub>3</sub>	
<i>Maackia amurensis</i>	130	2( $\alpha\beta$ )	O	sialic acid	+
<i>Maclura pomifera</i>	40-43	2( $\alpha\beta$ )	–	$\alpha$ -gal, $\alpha$ -galNAc	
<i>Momordica charantia</i>	115-129	4( $\alpha\beta$ )	–	gal, galNAc	
<i>Naja mocambique mocambique</i>	–	–	–	–	
<i>Naja naja kaouthia</i>	–	–	–	–	
<i>Narcissus pseudonarcissus</i>	26	2	(h)	$\alpha$ -D-man	
<i>Perseu americana</i>	–	–	–	–	
<i>Phaseolus coccineus</i>	112	4	–	–	
<i>Phaseolus limensis</i>	247(II)	8	A	galNAc	+
	124(III)	4			
<i>Phaseolus vulgaris</i>					
PHA-E	128	4	–	oligosaccharide	+
PHA-L	128	4	–	oligosaccharide	+
PHA-P					
PHA-M					

----- Table continued on next page -----

Lectin	MW (kDa)	Subunits	Specificity		Mitogenic Activity
			Blood Group	Sugar	
<i>Phytolacca americana</i>	32	–	–	(glcNAc) <sub>3</sub>	+
<i>Pisum sativum</i>	49	4(αβ)	–	α-man	+
<i>Pseudomonas aeruginosa PA-I</i>	13-13.7	–	–	gal	+ <sup>c</sup>
<i>Psophocarpus tetragonolobus</i>	35	1	–	galNAc, gal	
<i>Ptilota plumosa</i>	65; 170	–	B	α-gal	
<i>Ricinus communis</i>					
Toxin, RCA <sub>60</sub>	60	2	–	galNAc, β-gal	
Toxin, RCA <sub>120</sub>	120	4	–	β-gal	
<i>Sambucus nigra</i>	140	4(αβ)	–	αNeuNAC(2→6)gal	+ <sup>c</sup>
				galNAc	
<i>Solanum tuberosum</i>	50; 100 <sup>a</sup>	1, 2	–	(glcNAc) <sub>3</sub>	
<i>Sophora japonica</i>	133	4	A, B	β-galNAc	
<i>Tetragonolobus purpureas</i>	120(A)	4	H	α-L-fuc	
	58(BA)	2	H	α-L-fuc	
	117(C)	4	H	α-L-fuc	
<i>Triticum vulgare</i>	36	2	–	(glcNAc) <sub>2</sub> , NeuNAC	+
<i>Ulex europaeus</i>					
UEA I	68	–	H	α-L-fuc	
UEA II	68	–	–	(glcNAc) <sub>2</sub>	
<i>Vicia faba</i>	50	4(αβ)	–	man, glc	+
<i>Vicia sativa</i>	40	4(αβ)	–	glc, man	+
<i>Vicia villosa</i>	139	4	A <sub>1</sub> +T <sub>n</sub>	galNAc	
A <sub>4</sub>	134	4	A <sub>1</sub>	galNAc	
B <sub>4</sub>	143	4	T <sub>n</sub>	galNAc	
<i>Vigna radiata</i>	160	4	–	α-gal	
<i>Viscum album</i>	115	4(αβ)	–	β-gal	
<i>Wisteria floribunda</i>	68	2	–	galNAc	

<sup>a</sup> Concentration-dependent molecular weight

<sup>b</sup> Non-agglutinating and mitogenic

<sup>c</sup> Mitogenic for neuraminidase-treated lymphocytes

## References

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