

HPLC Carbohydrate Column Selection Guide

Because carbohydrates exhibit a significant degree of chemical and physical similarity, they are more difficult to analyze than most other classes of compounds. No single HPLC column or method is capable of separating all carbohydrates. Advantages, limitations, and applications for SUPELCOGEL and SUPELCOSIL columns specifically prepared for various carbohydrate analyses are summarized here.

Key Words:

- carbohydrates ● sugars ● saccharides
- monosaccharides ● disaccharides ● oligosaccharides

Carbohydrate analyses are a challenge for the chromatographer. Diversity among compounds classified as carbohydrates (Table 1) is far greater than among other classes of biochemicals. The potential for complexity in the structure of carbohydrate molecules is summarized by the fact that three amino acids can combine in a total of six different ways, while three monosaccharides can form more than 1000 distinct trisaccharides. To further complicate the situation, chemical and physical properties among monosaccharides, disaccharides, and trisaccharides differ only slightly. Hence, HPLC separations of carbohydrates depend on differences in conformation, configuration, and bonding mode, and are more difficult than analyses of other classes of compounds. No single HPLC column or method is capable of separating all carbohydrates. For this reason, we offer a selection of columns specifically prepared for various carbohydrate analyses. Advantages and limitations of each column are summarized in this bulletin. Table 2 summarizes applications for these columns; Table 3 shows retention times for an extensive number of carbohydrates, under typical operating conditions.

Table 1. Types of Carbohydrates

Monosaccharides

Pentoses (arabinose, ribose)

Hexaoses (glucose, galactose, fructose)

Disaccharides (sucrose, lactose, maltose)

Trisaccharides (raffinose, melezitose)

Tetrasaccharides (stachyose)

Oligosaccharides (DP3 to DP10*)

Polysaccharides (100–1000 units)

Polyhydric Alcohols/Sugar Alcohols

(mannitol, sorbitol, polyols)

*DP (degree of polymerization) = number of monosaccharide units in the molecule.

Detection

Because the UV wavelengths required to detect carbohydrates also will be strongly absorbed by impurities in the samples (Figure A), refractive index (RI) detection is most commonly used in analyses of carbohydrates. The major advantage of RI detection is the

universal response over a fairly wide linear range of analyte concentrations. Major disadvantages are poor sensitivity and baseline instability caused by temperature, solvent, or pressure changes (RI detection is impractical with gradient elution). Both positive and negative peaks can be present in a chromatogram. The abundance of carbohydrates in food and other samples, however, makes RI detection a suitable tool for these analyses.

Figure A. Refractive Index Affords Superior Detection for Carbohydrates

Column: SUPELCOSIL LC-NH₂, 25cm x 4.6mm, 5µm particles
 Cat. No.: 58338
 Mobile Phase: acetonitrile:water, 75:25
 Flow Rate: 1.5mL/min
 Temp.: ambient
 Inj.: 20µL, 10mg/mL each analyte in acetonitrile:water, 25:75

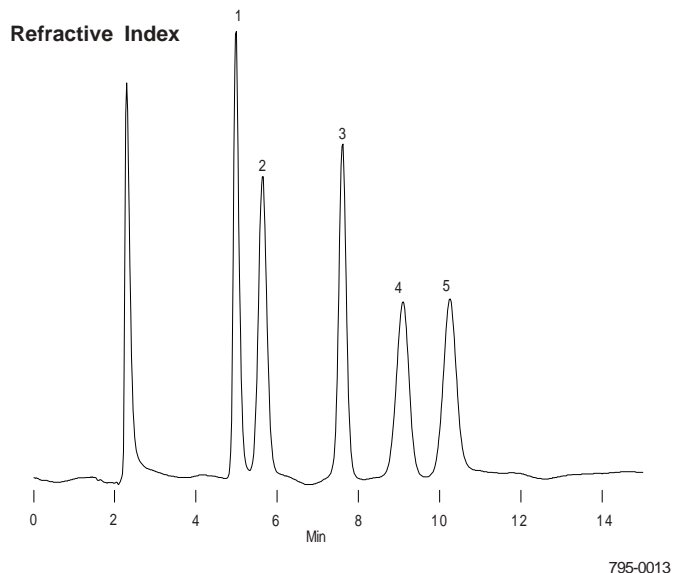
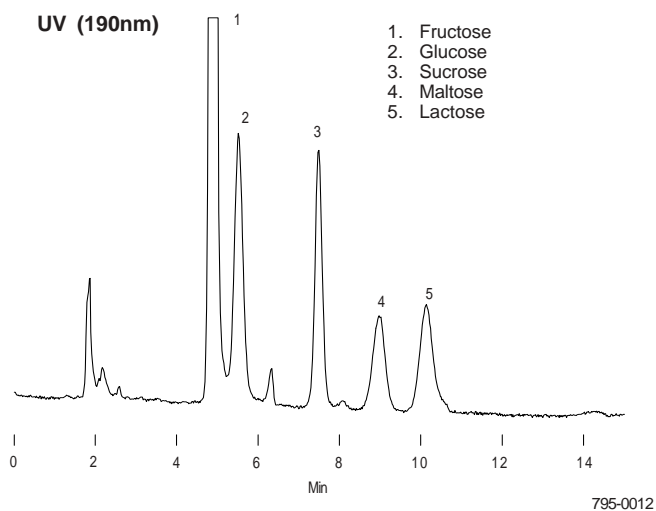


Table 2. Carbohydrate Column Applications and Operating Parameters

SUPELCOGEL™ columns are resin-based; SUPELCO SIL™ LC-NH₂ column is silica-based.

| Column | Application | Form | Typical Mobile Phase | Maximum Flow (mL/min) | Sample pH Range | Max. Temp. (°C) |
|--------------------------------|---|--------------------|---|-----------------------|-----------------|-----------------|
| SUPELCOGEL K | beet sugar, cane sugar, molasses, corn syrup | potassium | 10mM K ₂ HPO ₄ | 1.5 | 1-13 | 90 |
| SUPELCOGEL Pb | monosaccharides, xylose/galactose/mannose | lead | deionized water | 1.5 | 1-13 | 90 |
| SUPELCOGEL Ca | high fructose corn syrup, monosaccharides, sugar alcohols, oligosaccharides | calcium | deionized water | 1.5 | 1-13 | 90 |
| SUPELCOGEL C-610H | organic acids, acid/carbohydrate/ alcohol mixes | hydrogen | 0.1% H ₃ PO ₄ or H ₂ SO ₄ | 1.5 | 1-13 | 60 |
| SUPELCOGEL H | organic acids, acid/carbohydrate/ alcohol mixes | hydrogen | 0.1% H ₃ PO ₄ or H ₂ SO ₄ | 1.5 | 1-13 | 90 |
| SUPELCOGEL C-611 | mono-, di-, and trisaccharides, galactose/mannose | 2 divalent cations | 10 ⁻⁴ N NaOH | 1.5 | 1-13 | 85 |
| SUPELCOGEL Ag | oligosaccharides, glycerol/ethanol, beer, corn syrup, hydrolyzed starch | silver | deionized water | 1.5 | 1-13 | 90 |
| SUPELCO SIL LC-NH ₂ | mono-, di-, some trisaccharides | aminopropyl silica | 75% CH ₃ CN in water | 2.0 | 2-7 | 70 |

Table 3. Typical Retention Times on Supelco Carbohydrate Columns

| Cat. No.: | SUPELCOGEL Columns | | | | | | | | SUPELCO SIL LC-NH ₂ |
|---------------------|-----------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------|-------------------------|--------------------------------------|-------------------|--------------------------------|
| | Ca 59305-U | C-610H 59320-U | H 59304-U | H 59346 | Pb 59343 | C-611 59310-U | K 59342 | Ag2 59315 | 58338 |
| Dimens. (mm): | 300 x 7.8 | 300 x 7.8 | 300 x 7.8 | 250 x 4.6 | 300 x 7.8 | 300 x 7.8 | 300 x 7.8 | 300 x 7.8 | 250 x 4.6 |
| Temp.: | 80°C | 30°C | 30°C | 30°C | 85°C | 60°C | 85°C | 85°C | ambient |
| Mobile Phase: | DH ₂ O | 0.1% H ₃ PO ₄ | 0.1% H ₃ PO ₄ | 0.1% H ₃ PO ₄ | DH ₂ O | 10 ⁻⁴ N NaOH | 15mM K ₂ HPO ₄ | DH ₂ O | ACN: DH ₂ O (3:1) |
| Flow Rate (mL/min): | 0.5 | 0.5 | 0.5 | 0.17 | 0.5 | 0.5 | 0.5 | 0.5 | 1.0 |
| Det.: | refractive index | | | | | | | | |
| Compound | Retention Times (min) | | | | | | | | |
| Arabinose | 15.3 | 13.9 | 14.3 | 13.8 | 19.2 | 19.6 | 16.8 | 17.1 | 7.5 |
| Arabitol | 19.8 | 14.1 | 14.9 | 14.3 | 32.3 | 22.8 | 13.5 | 16.0 | 7.2 |
| Betaine | ND | ND | ND | ND | NR | ND | 13.0 | ND | ND |
| Dulcitol | 22.3 | 13.4 | 14.2 | 13.7 | 43.4 | 25.7 | 12.9 | 15.9 | 9.0 |
| Erythritol | 17.7 | 15.0 | 15.6 | 14.8 | 24.5 | 20.2 | 14.0 | 16.1 | 5.9 |
| Ethanol | 19.4 | 25.6 | ND | ND | ND | 21.0 | ND | 18.4 | NR |
| Fructose | 14.9 | 13.1 | 13.3 | 12.9 | 20.8 | 20.7 | 15.2 | 16.0 | 8.3 |
| Galactose | 13.4 | 12.9 | 13.0 | 12.6 | 17.6 | 17.6 | 15.1 | 15.8 | 10.3 |
| Glucose | 12.0 | 12.1 | 11.9 | 11.7 | 14.9 | 15.8 | 14.0 | 14.6 | 9.8 |
| Glycerol | 18.7 | 16.8 | 17.6 | 16.6 | 23.8 | 20.9 | 15.2 | 17.1 | NR |
| Inositol | 14.9 | 12.6 | 12.7 | 12.4 | 24.5 | 20.1 | 15.7 | 17.4 | ND |
| Isomaltose | 9.6 | 10.3 | ND | ND | ND | 13.8 | ND | 11.6 | 19.4 |
| Isomaltotriose | 8.5 | 9.5 | ND | ND | ND | 12.6 | ND | 9.8 | NR |
| Lactitol | ND | ND | 11.1 | 11.0 | 26.5 | ND | 10.6 | ND | ND |
| Lactose | 10.2 | 10.8 | 10.2 | 10.2 | 13.5 | 14.3 | 10.9 | 11.8 | 19.5 |
| Maltitol | 13.6 | 11.0 | 10.7 | 10.7 | 23.8 | 17.7 | 10.2 | 15.0 | 15.5 |
| Maltoheptaose | 7.5 | 8.8 | 7.6 | 7.9 | 9.2 | 11.6 | 7.2 | 7.3 | NR |
| Maltohexaose | 7.7 | 8.9 | 7.7 | 8.1 | 9.7 | 12.0 | 7.4 | 7.6 | NR |
| Maltopentaose | 7.9 | 9.1 | 7.9 | 8.2 | 10.5 | 12.6 | 7.8 | 8.1 | NR |
| Maltose | 9.8 | 10.5 | 9.9 | 9.9 | 13.0 | 14.2 | 10.7 | 11.5 | 17.4 |
| Maltotetraose | 8.3 | 9.3 | 8.2 | 8.5 | 11.2 | 13.2 | 8.4 | 8.8 | NR |
| Maltotriose | 8.8 | 9.7 | 8.8 | 9.0 | 12.0 | 13.6 | 9.2 | 9.8 | 31.0 |
| Mannitol | 19.2 | 13.2 | 13.7 | 13.2 | 32.5 | 22.1 | 12.6 | 15.2 | 9.2 |
| Mannose | 13.7 | 12.8 | 12.9 | 12.5 | 19.8 | 18.9 | 15.6 | 15.9 | 9.1 |
| Melezitose | 8.7 | 9.7 | 8.8 | 9.0 | 10.8 | 12.4 | 8.6 | 9.3 | 24.5 |
| Psicose | 22.5 | 13.4 | 14.5 | 13.9 | 36.5 | 32.9 | 15.5 | 17.2 | 6.6 |
| Raffinose | 8.7 | 9.7 | 8.7 | 8.9 | 11.2 | 12.6 | 8.7 | 9.6 | 29.7 |
| Ribitol | 16.7 | 13.7 | 14.2 | 13.6 | 25.1 | 19.5 | 13.1 | 15.3 | ND |
| Ribose | 24.3 | 14.2 | 15.8 | 15.0 | 40.7 | 34.6 | 17.7 | 19.1 | 6.0 |
| Sorbitol | 23.4 | 13.4 | 14.4 | 13.9 | 46.9 | 28.3 | 13.3 | 16.3 | 9.0 |
| Stachyose | 8.1 | 9.3 | 8.1 | 8.4 | 10.4 | 11.9 | 7.9 | 8.5 | 67.3 |
| Sucrose | 9.8 | 10.6 | 9.9 | 9.9 | 12.2 | 13.6 | 10.1 | 11.2 | 14.0 |
| Xylitol | 23.3 | 14.4 | 15.7 | 15.0 | 42.1 | 28.0 | 14.2 | 17.1 | 7.3 |
| Xylose | 13.2 | 12.8 | 12.8 | 12.6 | 16.1 | 17.2 | 15.3 | 15.6 | 6.8 |

NR - not recommended

ND - no data available

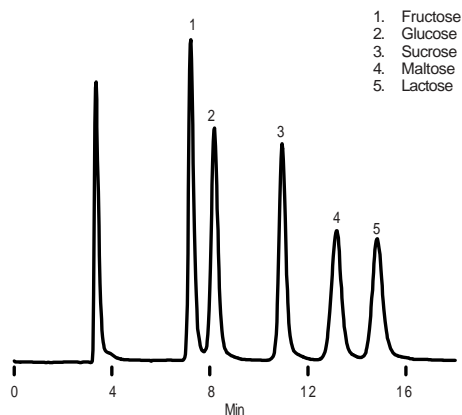
For optimal separations, allow at least 1 minute between compounds.

SUPELCO SIL LC-NH₂ Column

This column, containing a silica-based, aminopropyl-modified phase packing, provides good separations of a variety of mono-, di-, and some trisaccharides found in foods. Separation is in order of increasing molecular weight, with monosaccharides eluting first (Figures B and C). Oligosaccharides are strongly retained and should not be analyzed on this column. Sample preparation is minimal. The sugars do not require derivatization and most samples require only dilution with deionized water, extraction of the sugars, and filtration (0.45µm filter). Many juices and soft drinks can be analyzed undiluted, after degassing and filtration. Mobile phases typically contain acetonitrile and water in various proportions (75:25 or 80:20 mixtures are recommended). Note that the disaccharides sucrose and lactose are best separated on the SUPELCO SIL LC-NH₂ column.

Figure B. Amino Column Test Mix

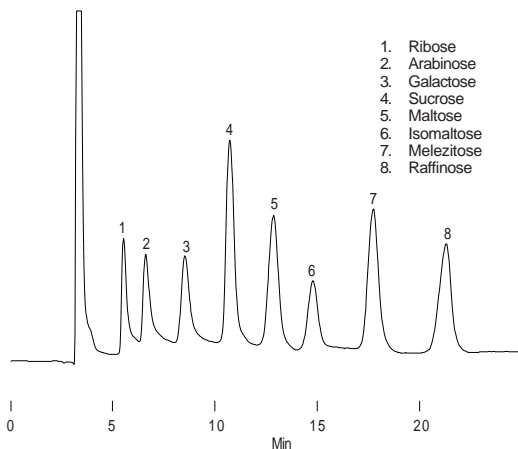
Column: **SUPELCO SIL LC-NH₂, 25cm x 4.6mm, 5µm particles**
Cat. No.: **58338**
Mobile Phase: acetonitrile:water, 75:25
Flow Rate: 1.0mL/min
Temp.: ambient
Det.: RI
Inj.: 10µL, 10mg/mL each analyte in acetonitrile:water, 25:75



795-0014

Figure C. Carbohydrate Standards on an Amino Column

Column: **SUPELCO SIL LC-NH₂, 25cm x 4.6mm, 5µm particles**
Cat. No.: **58338**
Mobile Phase: acetonitrile:water, 75:25
Flow Rate: 1.0mL/min
Temp.: ambient
Det.: RI
Inj.: 10µL, 10mg/mL each analyte in water



795-0015

Figure B shows an analysis of the test mix used to evaluate the performance of SUPELCO SIL LC-NH₂ columns. The mix is available (Cat. No. 58424) for routine monitoring of the column in the laboratory. Figure C shows the variety of sugars that can be separated by using a SUPELCO SIL LC-NH₂ column. Resolution between each pair of analytes is good.

SUPELCO GEL Columns

In contrast to their elution order on SUPELCO SIL LC-NH₂ columns, carbohydrates elute in descending order of molecular size, monosaccharides last, from the resin-based SUPELCO GEL columns described below. The pores in the resins exclude polysaccharides and larger oligosaccharides, which elute first. Smaller di- and monosaccharides enter the pores, interact with the counterions, and are more strongly retained. Figures D and E show comparable separations on five columns. In addition to the differences in elution order and resolution, note the negative peak, a common characteristic of refractive index detection, in both figures. Also note that in Figure E the disaccharides sucrose and lactose are separated only on the SUPELCO SIL LC-NH₂ column. Disaccharide separations are difficult to obtain with resin-based column packings.

SUPELCO GEL Ca Column

The SUPELCO GEL Ca column contains a polystyrene-divinylbenzene cross-linked resin in the calcium form. It separates oligo-, tri-, and disaccharides by class, using a mixed size exclusion/ion exchange mode, with the largest molecules eluting first. Its true separating power, however, is in its chromatography of monosaccharides and sugar alcohols – a variety of monosaccharides can be separated, using only water as the mobile phase. The column can be operated at low temperatures, but separations are best at elevated temperatures. Figures F and G show a separation of carbohydrate standards and an analysis of high fructose corn syrup, respectively, on a SUPELCO GEL Ca column. Both figures were obtained using a temperature of 80°C and a mobile phase consisting solely of deionized water.

SUPELCO GEL C-611 Column

The unique polystyrene-divinylbenzene resin-based packing in this column contains two divalent cations, strontium and barium, rather than one. As with the other SUPELCO GEL columns, the separation mechanism is a combination of size exclusion and ion exchange modes. Disaccharides elute before monosaccharides, usually as a single peak. Sugar alcohols are strongly retained and elute with the monosaccharides. The column is compatible with many inorganic salts and can be used with dilute bases. Separations normally are carried out using a very weak basic solution, such as 10⁻⁴ N sodium hydroxide, at 60°C. Analyses on a SUPELCO GEL C-611 column are temperature dependent. Figure I shows that resolution is improved as the temperature is increased.

SUPELCO GEL Ag Columns

These columns contain a silver-form styrene-divinylbenzene resin. Larger analytes elute before smaller analytes. The Ag1 column was developed specifically for separating the oligosaccharides in beer and corn syrup, and provides rapid separations of oligomers up to DP7. It will separate ethanol and glycerol. The Ag2 column is better suited for larger oligosaccharides, resolving up to and including DP12. Hydrolysis products and oligosaccharides can be analyzed on either an Ag1 or an Ag2 column. Figure J shows the carbohydrate profile of a domestic beer; Figures K and L show separations of corn syrup on SUPELCO GEL Ag1 and Ag2 columns, respectively.

Figure D. Carbohydrate Standards

Flow Rate (SUPELCO SILLC-NH₂ column): 1.5mL/min
 Inj.: 10µL, 2.5mg/mL each analyte in water
 Other conditions listed in Table 3

1. Maltotriose
2. Maltose
3. Glucose
4. Xylose
5. Arabinose
6. Ribitol
7. Arabitol
8. Xylitol

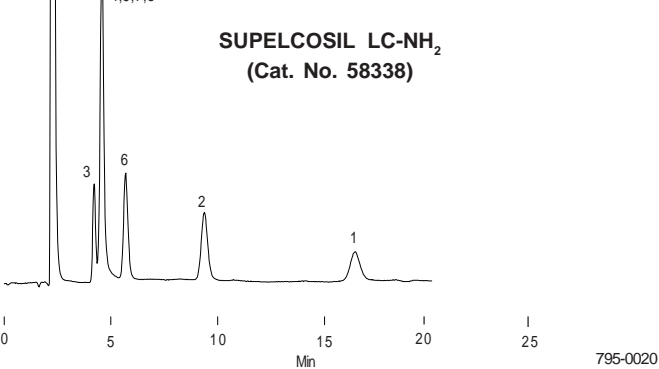
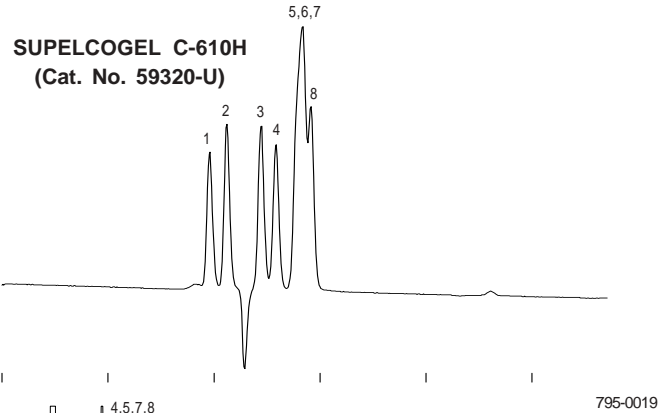
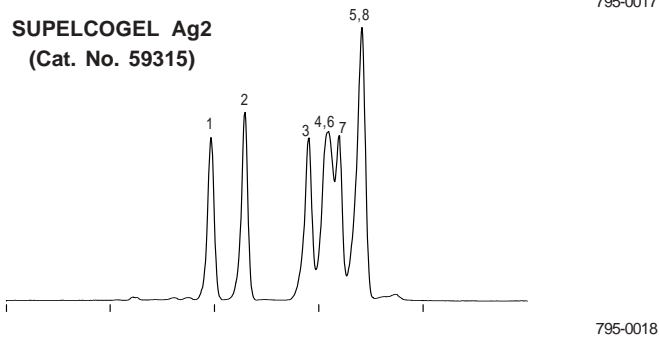
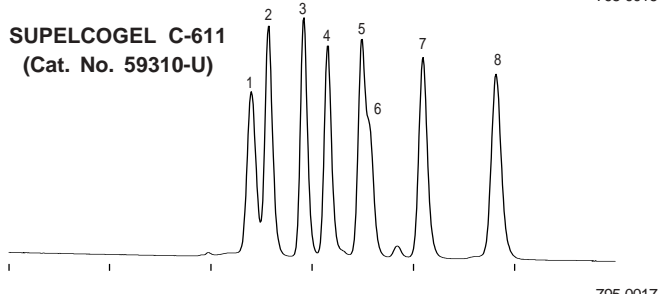
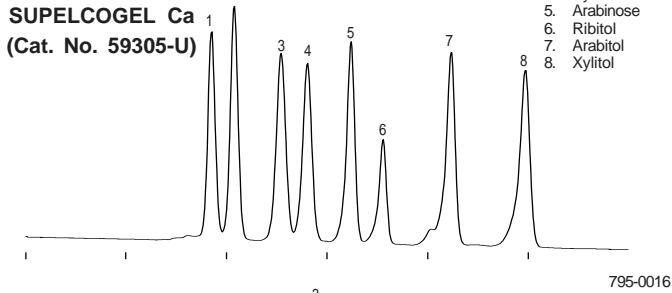


Figure E. Carbohydrates in Fruit Yogurt

Flow Rate (SUPELCO SILLC-NH₂ column): 1.5mL/min
 Inj.: 10µL of 10g yogurt/100mL DI water, filtered (0.20µm filter)
 Other conditions listed in Table 3

1. Oligosaccharides
2. Sucrose
3. Lactose
4. Glucose
5. Fructose

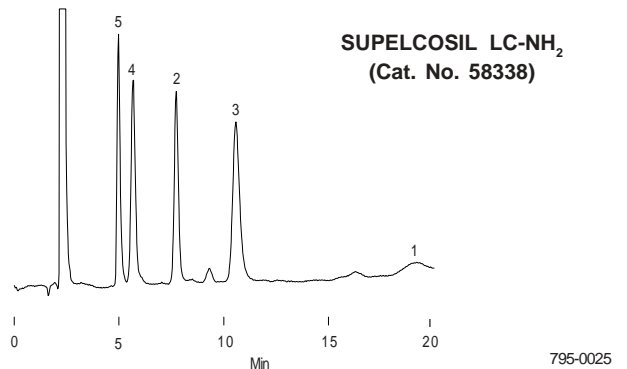
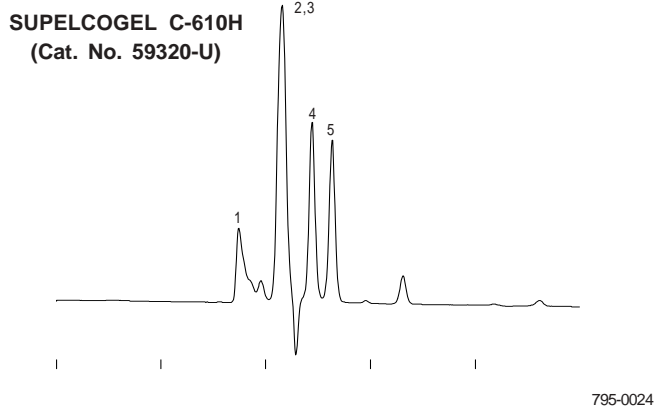
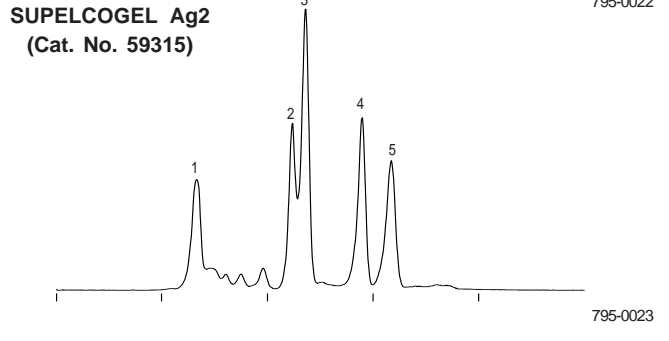
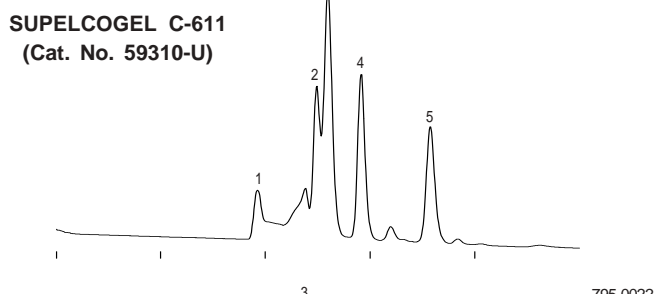
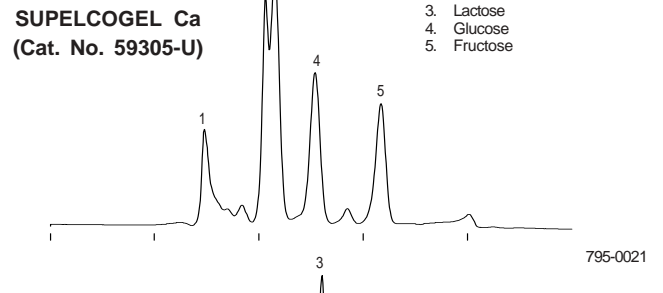
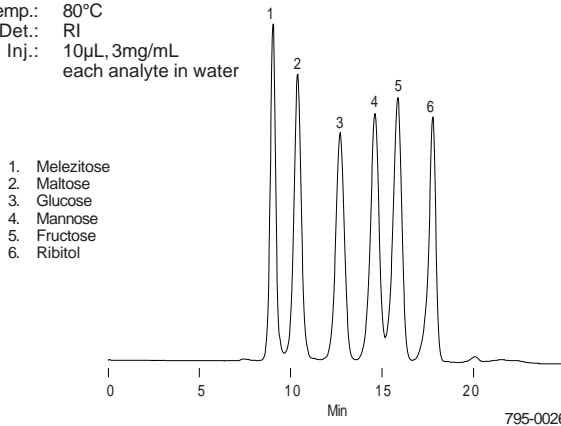


Figure F. Carbohydrate Standards on a SUPELCOGEL Ca Column

Column: **SUPELCOGEL Ca, 30cm x 7.8mm**
 Cat. No.: **59305-U**
 Mobile Phase: water
 Flow Rate: 0.5mL/min
 Temp.: 80°C
 Det.: RI
 Inj.: 10µL, 3mg/mL each analyte in water

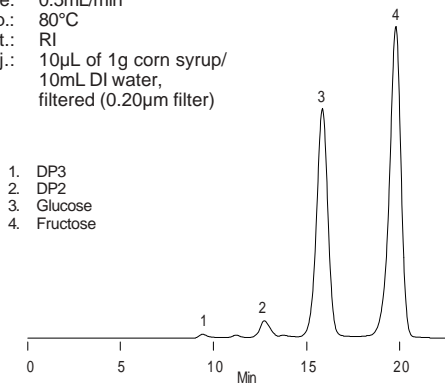


1. Melezitose
2. Maltose
3. Glucose
4. Mannose
5. Fructose
6. Ribitol

795-0026

Figure G. High Fructose Corn Syrup

Column: **SUPELCOGEL Ca, 30cm x 7.8mm**
 Cat. No.: **59305-U**
 Mobile Phase: water
 Flow Rate: 0.5mL/min
 Temp.: 80°C
 Det.: RI
 Inj.: 10µL of 1g corn syrup/
 10mL DI water,
 filtered (0.20µm filter)

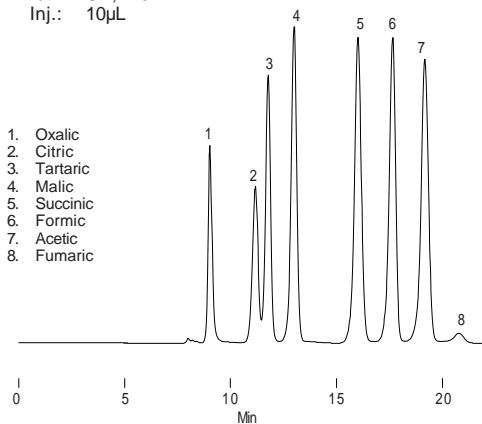


1. DP3
2. DP2
3. Glucose
4. Fructose

795-0027

Figure H. Organic Acid Standards

Column: **SUPELCOGEL C-610H, 30cm x 7.8mm**
 Cat. No.: **59320-U**
 Mobile Phase: 0.1% phosphoric acid
 Flow Rate: 0.5mL/min
 Temp.: 30°C
 Det.: UV, 210nm
 Inj.: 10µL



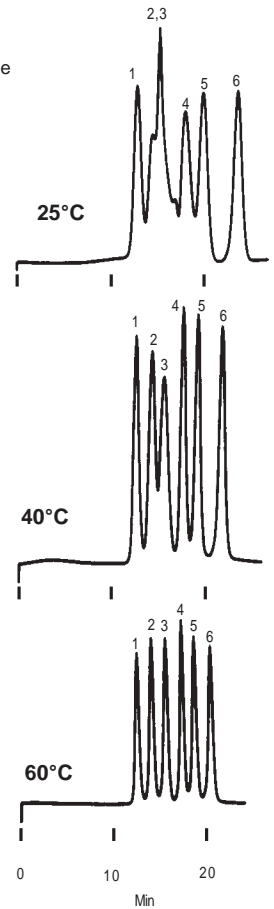
1. Oxalic
2. Citric
3. Tartaric
4. Malic
5. Succinic
6. Formic
7. Acetic
8. Fumaric

794-0231

Figure I. Temperature Affects Carbohydrate Analyses on SUPELCOGEL Columns

Column: **SUPELCOGEL C-611, 30cm x 7.8mm**
 Cat. No.: **59310-U**
 Mobile Phase: 10⁻⁴ N sodium hydroxide
 Flow Rate: 0.5mL/min
 Det.: RI
 Inj.: 10µL

1. Raffinose
2. Maltose
3. Glucose
4. Galactose
5. Mannose
6. Fructose

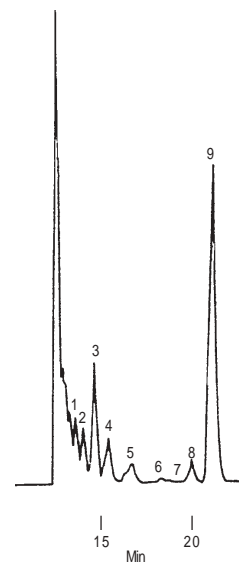


713-0519

Figure J. Domestic Beer

Column: **SUPELCOGEL Ag1, 30cm x 7.8mm**
 Cat. No.: **59318-U**
 Mobile Phase: water
 Flow Rate: 0.3mL/min
 Temp.: 90°C
 Det.: RI
 Inj.: 20µL beer

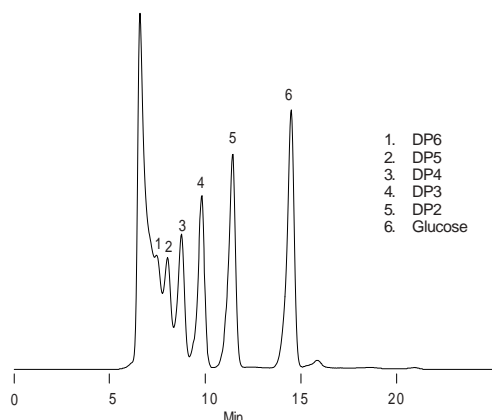
1. DP6
2. DP5
3. DP4
4. Maltotriose
5. Maltose
6. Glucose
7. Fructose
8. Glycerol
9. Ethanol



G000994

Figure K. Dark Corn Syrup

Column: **SUPELCOGEL Ag1, 30cm x 7.8mm**
Cat. No.: **59318-U**
Mobile Phase: water
Flow Rate: 0.5mL/min
Temp.: 90°C
Det.: RI
Inj.: 10µL of 1g syrup/10mL DI water, filtered (0.20µm filter)



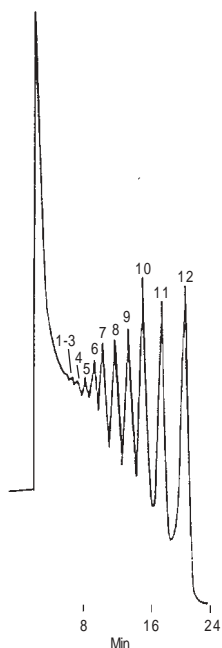
- 1. DP6
- 2. DP5
- 3. DP4
- 4. DP3
- 5. DP2
- 6. Glucose

795-0029

Figure L. Corn Syrup

Column: **SUPELCOGEL Ag2, 30mm x 7.8mm**
Cat. No.: **59315**
Mobile Phase: water
Flow Rate: 0.4mL/min
Temp.: 90°C
Det.: RI
Inj.: 20µL

- 1-3. DP12-10
- 4. DP9
- 5. DP8
- 6. DP7
- 7. DP6
- 8. DP5
- 9. DP4
- 10. Maltotriose
- 11. Maltose
- 12. Glucose



G000995

SUPELCOGEL C-610H Column

This column contains a polystyrene resin in the hydrogen form. It is ideal for separating mixtures of organic acids (Figure H), fermentation products (e.g., alcohols), and carbohydrates. Such mixtures commonly occur in fruits, vegetables, and beverages. Larger and acidic analytes elute before smaller analytes. The column is stable between pH 1 and 13, but results are best at low pH. A simple mobile phase containing 0.1% H₃PO₄ is suitable for a wide variety of analyses.

SUPELCOGEL K Column

A SUPELCOGEL K column separates raffinose, sucrose, glucose, fructose, and betaine, a trimethylammonium zwitterionic compound found in beet and cane sugars and widely distributed in other plants.

SUPELCOGEL Pb Column

The lead-form resin in SUPELCOGEL Pb columns provides the highest resolution and best selectivity for monosaccharides. SUPELCOGEL Pb columns provide excellent separation of xylose, galactose, and mannose, which are not completely resolved on calcium-form resin columns.

SUPELCOGEL H Columns

SUPELCOGEL H columns have the same particle composition, retention mechanism, performance, sensitivity, and applications as SUPELCOGEL C-610H columns. However, particle improvements have made it possible to pack the SUPELCOGEL H packing material efficiently into conventional 4.6mm ID columns, to improve detection and reduce solvent consumption relative to 7.8mm ID columns.

Making a Decision

To choose the best column for your particular carbohydrates analysis, we suggest you find the compounds of interest in Table 3 and note their retention times on each column. Using this information, select the column that will separate the compounds of interest with at least 1 minute between any pair.

Guard Columns

Although filtration removes particulate matter from a sample, food, beverages, and other samples often contain soluble components that can be strongly retained by the analytical columns described here. We strongly recommend using a guard column to protect the analytical column from these potentially damaging sample components. We offer 5cm x 4.6mm Supelguard™ guard columns that are compatible with our SUPELCOGEL resin-based analytical columns and a 2cm x 4.6mm Supelguard guard column for use with the SUPELCOGEL LC-NH₂ silica-based column. Guard columns are listed under Ordering Information. Note that SUPELCOGEL Ca columns and SUPELCOGEL C-611 columns use the same guard column (Supelguard Ca).

Suggested Reading

- Knight, P. *Biotechnology*, **7**: 35 (1989).
- Parriot, D. *A Practical Guide to HPLC Detection* (Chapter 2) Academic Press (1992).
- Lehninger, A. *Biochemistry* (Chapter 10) Worth Publishers (1975).
- Nollet, L. *Food Analysis by HPLC* (Chapter 8) Marcel Dekker (1992).

Ordering Information:

SUPELCOGEL and SUPELCOFIL Carbohydrate Columns and Guard Columns

| Column | Length (cm) | ID (mm) | Cat. No. | Supelguard Guard Column | Cat. No. |
|-------------------------------|-------------|---------|----------|--|----------------|
| SUPELCOGEL K | 30 | 7.8 | 59342 | K | 59344 |
| SUPELCOGEL Pb | 30 | 7.8 | 59343 | Pb | 59345 |
| SUPELCOGEL Ca | 30 | 7.8 | 59305-U | Ca | 59306-U |
| SUPELCOGEL C-610H | 30 | 7.8 | 59320-U | H | 59319 |
| SUPELCOGEL H | 30 | 7.8 | 59304-U | H | 59319 |
| SUPELCOGEL H | 25 | 4.6 | 59346 | H | 59319 |
| SUPELCOGEL C-611 | 30 | 7.8 | 59310-U | Ca | 59306-U |
| SUPELCOGEL Ag1 | 30 | 7.8 | 59318-U | Ag1 | 59317-U |
| SUPELCOGEL Ag2 | 30 | 7.8 | 59315 | Ag2 | 59316 |
| SUPELCOFIL LC-NH ₂ | 25 | 4.6 | 58338 | LC-NH ₂ (kit) LC-NH ₂ (pk. 2) | 59558 59568 |

Carbohydrate/Organic Acid/Sugar Alcohol Reference Standards

Prepared, tested, and packaged using rigorous manufacturing procedures.

| Description | CAS No. | Qty. | Cat. No. | Description | CAS No. | Qty. | Cat. No. |
|------------------------------|-------------|-------|----------|--------------------------------------|----------|-------|----------|
| Monosaccharides | | | | d-(+)Arabitol | 488-82-4 | 500mg | 46919-U |
| d-(-)Arabinose | 28697-53-2 | 500mg | 47246-U | Dulcitol (Galactitol) | 608-66-2 | 500mg | 46920-U |
| d-(-)Fructose | 57-48-7 | 500mg | 47247 | iso-Erythritol | 149-32-6 | 500mg | 46921 |
| d-(+)Galactose | 59-23-4 | 500mg | 47248 | Glycerol | 56-81-5 | 500mg | 46922 |
| d-(+)Glucose (mixed anomers) | 50-99-7 | 500mg | 47249 | Maltitol | 585-88-6 | 500mg | 46923-U |
| d-(+)Mannose (mixed anomers) | 3458-28-4 | 500mg | 47250 | D-Mannitol | 69-65-8 | 500mg | 46924-U |
| D-Psicose (mixed anomers) | 551-68-8 | 100mg | 47251 | Ribitol (Adonitol) | 488-81-3 | 500mg | 46925-U |
| d-(-)Ribose | 50-69-1 | 500mg | 47252 | D-Sorbitol | 50-70-4 | 500mg | 46926-U |
| d-(+)Xylose | 58-86-6 | 500mg | 47253 | Xylitol | 87-99-0 | 500mg | 46927 |
| Disaccharides | | | | Kits | | | |
| α-Lactose | 5989-81-1 | 500mg | 47287-U | Description | | | |
| Maltose | 6363-53-7 | 500mg | 47288 | Monosaccharides Kit: | | | |
| Sucrose | 57-50-1 | 500mg | 47289 | each monosaccharide standard listed | | | 47267 |
| Oligosaccharides | | | | Disaccharides Kit: | | | |
| Maltoheptaose (DP7) | 34620-78-5 | 100mg | 47872 | each disaccharide standard listed | | | 47268-U |
| Maltohexaose (DP6) | 34620-77-4 | 100mg | 47873 | Oligosaccharides Kit: | | | |
| Maltopentaose (DP5) | 34620-76-3 | 100mg | 47876 | each oligosaccharide standard listed | | | 47265 |
| Maltotetraose (DP4) | 34612-38-9 | 100mg | 47877 | Organic Acids Kit: | | | |
| Stachyose (DP4) | 10094-58-3 | 100mg | 47879 | each organic acid standard listed | | | 47264 |
| Maltotriose (DP3) | 1109-28-0 | 100mg | 47878 | Sugar Alcohols Kit: | | | |
| d-(+)Melezitose (DP3) | 10030-67-8 | 100mg | 47882-U | each sugar alcohol standard listed | | | 47266 |
| d-(+)Raffinose (DP3) | 17629-30-0 | 100mg | 47883 | | | | |
| Isomaltotriose (DP3) | 3371-50-4 | 100mg | 47884 | | | | |
| Organic Acids | | | | | | | |
| Acetic acid | 64-19-7 | 500mg | 46928 | | | | |
| Adipic acid | 124-04-9 | 500mg | 46929 | | | | |
| L-Ascorbic acid | 50-81-7 | 500mg | 46930-U | | | | |
| Benzoic acid | 65-85-0 | 500mg | 46931 | | | | |
| Butyric acid | 107-92-6 | 500mg | 46932 | | | | |
| Citric acid | 77-92-9 | 500mg | 46933 | | | | |
| Formic acid | 64-18-6 | 500mg | 46934-U | | | | |
| Fumaric acid | 110-17-8 | 500mg | 46948 | | | | |
| Isobutyric acid | 79-31-2 | 500mg | 46935 | | | | |
| D,L-Isocitric acid | 1637-73-6 | 100mg | 46936 | | | | |
| L-(+)Lactic acid | 79-33-4 | 100mg | 46937 | | | | |
| Maleic acid | 110-16-7 | 500mg | 46939 | | | | |
| D-Malic acid | 636-61-3 | 100mg | 46940-U | | | | |
| Malonic acid | 141-82-2 | 500mg | 46938 | | | | |
| Oxalic acid | 144-62-7 | 500mg | 46941-U | | | | |
| Phytic acid | 123408-98-0 | 500mg | 46942-U | | | | |
| Propionic acid | 79-09-4 | 500mg | 46943-U | | | | |
| (-)Quinic acid | 77-95-2 | 500mg | 46944-U | | | | |
| Shikimic acid | 138-59-0 | 100mg | 46945-U | | | | |
| Succinic acid | 110-15-6 | 500mg | 46946-U | | | | |
| D-Tartaric acid | 147-71-7 | 500mg | 46947-U | | | | |

Sugar Alcohols

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