

**Enzymatic Assay of  $\beta$ -HYDROXYSTEROID DEHYDROGENASE  
(EC 1.1.1.51)**

**PRINCIPLE:**

Testosterone +  $\beta$ -NAD  $\xrightarrow[\text{Dehydrogenase}]{\beta\text{-Hydroxysteroid}}$  5-a-Androsterone-3, 17-dione +  $\beta$ -NADH

Abbreviations used:

$\beta$ -NAD = Nicotinamide Adenine Dinucleotide, Oxidized Form

$\beta$ -NADH = Nicotinamide Adenine Dinucleotide, Reduced Form

**CONDITIONS:** T = 25°C, pH = 8.9, A<sub>340nm</sub>, Light path = 1 cm

**METHOD:** Continuous Spectrophotometric Rate Determination

**REAGENTS:**

- A. 100 mM Sodium Pyrophosphate Buffer, pH 8.9 at 25°C  
(Prepare 100 ml in deionized water using Tetrasodium Pyrophosphate Decahydrate, Prod. No. P-9146. Adjust to pH 8.9 at 25°C with 1 M HCL.)
- B. 6 mM  $\beta$ -Nicotinamide Adenine Dinucleotide, Oxidized Form, Solution ( $\beta$ -NAD)  
(Prepare 5 ml in deionized water using  $\beta$ -Nicotinamide Adenine Dinucleotide, Oxidized Form, Prod. No. N-7004.)
- C. 0.015% (w/v) Testosterone Substrate Solution  
(Prepare 5 ml in Reagent F using Testosterone, Prod. No. T-1500.)
- D. 10 mM Potassium Phosphate Buffer, pH 7.2 at 25°C  
(Prepare 50 ml in deionized water using Potassium Phosphate, Monobasic, Prod. No. P-5379. Adjust to pH 7.2 with 1 M KOH.)
- E.  $\beta$ -Hydroxysteroid Dehydrogenase Enzyme Solution  
(Immediately before use, prepare a solution containing 0.3 units/ml in cold Reagent D.)

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**REAGENTS:** (continued)

F. Absolute Methanol (MeOH)  
(Use Methanol, Absolute, Stock No. 17-5.)

**PROCEDURE:**

Pipette (in milliliters) the following reagents into a suitable cuvettes:

	<u>Test</u>	<u>Blank</u>
Reagent A (Buffer)	0.60	0.60
Reagent B ( $\beta$ -NAD)	0.20	0.20
Reagent E (Enzyme Solution)	0.10	0.10
Deionized Water	2.00	2.00

Mix by inversion and equilibrate to 25°C. Monitor the  $A_{340nm}$  until constant, using a suitably thermostatted spectrophotometer. Then add:

Reaction C (Testosterone)	0.10	-----
Reagent F (MeOH)	-----	0.10

Immediately mix by inversion and record the increase in  $A_{340nm}$  for approximately 5 minutes. Obtain the  $r A_{340nm}/\text{minute}$  using the maximum linear rate for both the Test and Blank.

**CALCULATIONS:**

$$\text{Units/mg enzyme} = \frac{(r A_{340nm}/\text{min Test} - ?A_{340nm}/\text{min Blank})}{(6.22) (\text{mg enzyme/ml RM})}$$

6.22 = Millimolar extinction coefficient of  $\beta$ -NADH at 340nm

RM = Reaction Mix

**UNIT DEFINITION:**

One unit will oxidize 1.0  $\mu\text{mole}$  of testosterone per minute at pH 8.9 at 25°C, in the presence of  $\beta$ -NAD<sup>+</sup>.

**FINAL ASSAY CONCENTRATION:**

In a 3.0 ml reaction mix, the final concentrations are 20 mM sodium pyrophosphate, 0.4 mM  $\beta$ -NAD, 0.3 mM sodium phosphate, 0.0005% testosterone, and 0.03 units  $\beta$ -hydroxysteroid dehydrogenase.

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**NOTES:**

1. All product and stock numbers, unless otherwise indicated, are Sigma product and stock numbers.

**This procedure is for informational purposes. For a current copy of Sigma's quality control procedure contact our Technical Service Department.**