# Collagens for Cell Culture

<table>
<thead>
<tr>
<th>Product Number</th>
<th>Description</th>
<th>Source</th>
<th>Storage</th>
<th>Target Cells For Attachment</th>
<th>Concentration For Use</th>
<th>Refs</th>
</tr>
</thead>
<tbody>
<tr>
<td>C1809</td>
<td>COLLAGEN TYPE I Acid soluble powder</td>
<td>kangaroo tail</td>
<td>2-8 °C</td>
<td>muscle cells, hepatocytes, spinal ganglion, embryonic lung cells, schwann cells. Mediate the attachment of many cell types</td>
<td>6-10 µg/cm²</td>
<td>33</td>
</tr>
<tr>
<td>C7661</td>
<td></td>
<td>rat tail</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C9791</td>
<td>COLLAGEN TYPE I 0.1% Solution Sterile-filtered (Not suitable for 3D gel formation)</td>
<td>calf skin</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C8919</td>
<td>COLLAGEN TYPE I 0.1% Solution Sterile-filtered (Not suitable for 3D gel formation)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C9301</td>
<td>COLLAGEN TYPE II Powder</td>
<td>chicken sternal cartilage</td>
<td>-20 °C</td>
<td>chondrocytes</td>
<td></td>
<td>18,19</td>
</tr>
<tr>
<td>C0543</td>
<td>COLLAGEN TYPE IV Powder</td>
<td>Engelbreth-Holm-Swarm mouse sarcoma</td>
<td>-20 °C; store solubilized product at 2-8 °C</td>
<td>epithelial cells, endothelial cells, muscle cells, nerve cells</td>
<td></td>
<td>5,12,13, 14,15, 16,17</td>
</tr>
<tr>
<td>C5533</td>
<td>COLLAGEN TYPE IV Lyophilized</td>
<td>human placenta</td>
<td>-20 °C</td>
<td></td>
<td></td>
<td>34</td>
</tr>
</tbody>
</table>

## Product Use

- **Collagen Type I (Product Nos. C1809, C7661, C9791, and C8919)**

  Optimal conditions for attachment must be determined for each cell line and application.

  1. Add collagen to 0.1 M acetic acid to obtain 0.1% (w/v) collagen solution. Stir at room temperature 1-3 hours until dissolved. (C8919 is prepared as a 0.1% solution, step 1 is not necessary for this product.)

  2. We recommend transferring the collagen solution to a glass bottle with a screw cap and carefully layering chloroform at the bottom. The amount of chloroform to use should be ~10% of the volume of collagen solution. DO NOT SHAKE OR STIR. Allow to stand overnight at 2–8 °C. Aseptically remove the top layer containing the collagen solution. We do not recommend sterilizing the collagen solution by membrane filtration. We have found substantial protein loss by this method. (C8919 is a sterile solution, step 2 is not necessary for this product.)

  3. Dilute desired volume (according to surface area to be treated) of sterile stock solution in step 2 or C8919 10-fold to a working concentration of 0.01% for coating surfaces.

  4. Coat dishes with 6-10 µg/cm². Allow the protein to bind for several hours at room temperature or 37 °C, or overnight at 2–8 °C.

  5. Remove excess fluid from the coated surface, and allow it to dry overnight. If the collagen solution is not sterile, the dried, coated surface can be sterilized easily by overnight exposure to UV light in a sterile tissue culture hood.

  6. Rinse with sterile tissue culture grade water or a balanced salt solution before introducing cells and medium.

- **Collagen Type II and Type IV (Product Nos. C9301, C0543, and C5533)**

  Optimal conditions for attachment must be determined for each cell line and application.

  1. Collagen Types II and IV may be reconstituted to a concentration of 0.5-2.0 mg/ml in 0.25% acetic acid. **Dissolve for several hours at 2–8 °C, occasionally swirling.**

  2. Coating of tissue culture plastic dishes may be performed by air drying the above protein solution or by preincubating the same solution overnight at 2–8 °C (or several hours at 37 °C) without air drying.

  3. Dried coated dishes can be sterilized overnight by exposure to UV light in a sterile tissue culture hood or by rinsing with 70% ethanol. Alternatively, the collagen solution may be sterilized by dialysis in a 0.25% acetic acid and 0.5% chloroform solution.
References: