StableCell™ Trypsin Solution
10×, 5.0 g porcine trypsin and 2 g EDTA • 4Na per liter of 0.9% sodium chloride
BioReagent, sterile-filtered, suitable for cell culture

Catalog Number T2610
Storage Temperature 2–8 °C

EC 3.4.21.4
Synonyms: Tryptase, Tryptar, Cocoonase, Parenzyme, Parenzymol

Product Description
Molecular mass: 23.4 kDa
Extinction Coefficient: ε1% = 15.0 (280 nm)

StableCell™ Trypsin 10× Solution (5.0 g/L of porcine trypsin and 2.0 g/L of EDTA • 4Na in 0.9% sodium chloride, plus a proprietary protein stabilizing agent) is sterile-filtered and cell culture tested.

Trypsin consists of a single chain polypeptide of 223 amino acid residues. Trypsin is produced by the removal of the N-terminus tyrosine from trypsinogen which is cleaved at the Lys6–Ile7 peptide bond. The amino acid sequence of trypsin is crosslinked by 6 disulfide bridges. This native form of trypsin is referred to as β-trypsin. Autoenzymatic cleavage of β-trypsin results in α-trypsin and is held together by disulfide bridges. Trypsin is a member of the serine protease family. The active site amino acid residues of trypsin include His46 and Ser193.1,3

Trypsin will cleave peptides on the C-terminal side of lysine and arginine amino acid residues. The rate of hydrolysis is slower if an acidic residue is on either side of the cleavage site and no cleavage occurs if a proline residue is on the carboxylic side of the cleavage site. The pH optimum of trypsin is 7–9.2 Trypsin will also hydrolyze ester and amide linkages of synthetic peptides such as benzoyl-L-arginineethyl ester (BAEE), p-toluensulfonfonyl-L-arginine methyl ester (TAME), tosyl-L-arginine methyl ester, Nα-benzoyl-L-arginine p-nitroanilide (BAPNA), L-lysyl-p-nitroanilide, and benzoyl-L-arginamide.2–5

Assuming the pH and temperature are the same and using a molar extinction coefficient of 808 at 254 nm for BAEE, the following conversions are valid:

1 BAEE μM Unit = 200 BAEE Units
1 TAME μM Unit = 0.27 BAEE μM Units
1 BAEE μM Unit = 3.64 TAME Units
1 TAME μM Unit = 55 BAEE A253 Units
1 BAEE A253 Unit = 0.018 TAME μM Unit
1 TAME μM Unit = 180 TAME A247 Units
1 TAME A247 Unit = 0.33 BAEE Units

A USP Unit = ΔA253 of 0.003 per minute
1 NF Unit = 3.3 A253 BAEE Units.

Note: These activity conversions were determined using bovine trypsin; however, they are thought to be similar for porcine trypsin.

Serine protease inhibitors that will inhibit trypsin include DFP (diisopropyl fluorophosphate), TLCK (Nα-p-tosyl-L-lysine chloromethyl ketone), PMSF (phenylmethylsulfonyl fluoride), APMSF(4-amidinophenylmethylsulfonyl fluoride), AEBSEF (4-(2-aminoethyl)benzenesulfonyl fluoride), apritinin, leupeptin, α2-macroglobulin, α1-antitrypsin, p-aminobenzamidine, benzamidine (reversible), soybean trypsin inhibitor, lima bean inhibitor, bovine pancreas trypsin inhibitor, chicken egg white inhibitor, and turkey egg white inhibitor.1,7

Precautions and Disclaimer
This product is for R&D use only, not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability
Recommended storage is 2–8 °C upon arrival. During stability studies, data showed this product retains ≥90% of its activity when stored at 37 °C for up to 8 weeks.
Procedure

StableCell™ Trypsin Solution may be used to remove adherent cells from a culture surface. Cells are most commonly removed from the culture substrate by treatment with trypsin or trypsin • EDTA. StableCell™ Trypsin Solutions can range from 0.05% to 0.5%. The reasons for the range of concentrations are as follows:

1. Differences in trypsin activity or potency,
2. Different incubation times, and
3. Different cell lines.

Incubating cells with too high a trypsin concentration for too long a time period will damage cell membranes and kill the cells. If unsure about the concentration of trypsin to use, use a low concentration. There can be lot-to-lot variation in dissociation times which is to be expected since the enzymatic activity of each lot will differ. If trypsin is being solubilized or diluted from a concentrated solution, it is important to use a buffered salt solution that contains no Ca\(^{2+}\) or Mg\(^{2+}\), such as Hank’s Balanced Salt Solution, Modified (Catalog No. H9394). Adjust the pH of trypsin solution to 7.4–7.6.

1. Remove medium from culture vessel by aspiration and wash the monolayer with Ca\(^{2+}\) and Mg\(^{2+}\)-free salt solution to remove all traces of serum. Remove salt solution by aspiration.

2. Dispense enough StableCell™ Trypsin Solution into culture vessel(s) to completely cover the monolayer of cells and place in 37°C incubator for ~2 minutes.

3. Remove the StableCell™ Trypsin Solution by aspiration and return closed culture vessel(s) to incubator. The coated cells are allowed to incubate until cells detach from the surface. Progress can be checked by examination with an inverted microscope.

   Note: The time required to remove cells from the culture surface is dependent on cell type, population density, serum concentration in the growth medium, potency of trypsin, and time since last subculture. Trypsin can cause cellular damage, thus time of exposure should be kept to a minimum.

4. When trypsinization process is complete the cells will be in suspension and appear rounded.

5. It is advisable to add serum or medium containing serum to the cell suspension as soon as possible to inhibit further tryptic activity which may damage cells. Soybean trypsin inhibitor (Catalog No. T6414) can also be added at an equimolar concentration to inhibit the trypsin that is present. Soybean trypsin inhibitor is used when culturing in serum-free conditions.

6. Cells can be resuspended by gently pipetting the cell suspension to break up the clumps. Further dilution can be made, if required, for cell counts and/or subculturing.

References


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