

## Product Information

### Lectin

#### from *Phaseolus vulgaris* Erythrohemagglutinin PHA-E

Product Number **L 8629**  
Storage Temperature 2-8 °C

#### Product Description

PHA-E has an approximate MW of 128 kDa. This product is affinity purified to remove additional protein content. Purity is determined by SDS-PAGE analysis. This product is extensively dialyzed against water prior to lyophilization to render it essentially salt free.

Lectin PHA-E is specifically reactive against erythrocytes. This product is essentially pure isolectin E4.

Lectins are proteins or glycoproteins of non-immune origin that agglutinate cells and/or precipitate complex carbohydrates. Lectins are capable of binding glycoproteins even in presence of various detergents.<sup>1</sup> The agglutination activity of these highly specific carbohydrate-binding molecules is usually inhibited by a simple monosaccharide, but for some lectins, di, tri, and even polysaccharides are required. Lectin PHA-E has not been proven to be specific for any particular oligosaccharide. Lectin PHA-E can promote agglutination of human erythrocytes at 8 µg/ml, but its activity is not inhibited by any monosaccharide such as lactose, galactose, glucose, mannose, etc.<sup>2,3</sup>

Lectins are isolated from a wide variety of natural sources, including seeds, plant roots and bark, fungi, bacteria, seaweed and sponges, mollusks, fish eggs, body fluids of invertebrates and lower vertebrates, and from mammalian cell membranes. The precise physiological role of lectins in nature is still unknown, but they have proved to be very valuable in a wide variety of applications *in vitro*, including:

1. blood grouping and erythrocyte polyagglutination studies.
2. mitogenic stimulation of lymphocytes.
3. lymphocyte subpopulation studies.
4. fractionation of cells and other particles.
5. histochemical studies of normal and pathological conditions.

Sigma offers a range of lectins suitable for the above applications. Most Sigma lectins are highly purified by affinity chromatography, but some are offered as purified or partially purified lectins, suitable for specific applications.

Many of the lectins are available conjugated to (conjugation does not alter the specificity of the lectin):

1. fluorochromes (for detection by fluorimetry).
2. enzymes (for enzyme-linked assays).
3. insoluble matrices (for use as affinity media).

Please refer to the table for general information on the most common lectins.

#### Procedure

This product has been tested for agglutination using washed red blood cells.

For washing the cells:

1. Transfer whole blood to graduated conical tubes. Use between 5-10 ml of whole blood.
2. Dilute the blood to 10 ml with phosphate buffered saline, pH 6.8.
3. Centrifuge the sample at medium speed for 5 minutes in a clinical centrifuge to force the red blood cells to the bottom.
4. Gently remove the plasma/buffer layer with a pipette.
5. Repeat steps 2-4 three times or until the upper layer is clear.
6. Measure the volume that the red blood cells occupy in the bottom of the tube. Dilute the cells 25-fold with phosphate buffered saline, pH 6.8.

The Agglutination Test is performed as follows:

1. Serially dilute the lectin as follows. Put 50 µl of phosphate buffered saline, pH 6.8 in each of the wells to be used. Add 50 µl of a 1 mg/ml lectin solution to the first well. Remove 50 µl of diluted lectin from the first well and add to the buffer in the next well. Repeat the serial dilutions as desired.

2. Carefully layer 50  $\mu$ l of washed red blood cells on top of the 50  $\mu$ l lectin solution.
3. When all the cell suspensions have been added, agitate to mix.
4. Allow to sit for 1 hour at room temperature and then read the results.

**Precautions and Disclaimer**

For Laboratory Use Only. Not for drug, household or other uses.

**Preparation Instructions**

This lectin is soluble in phosphate buffered saline, pH 6.8 (1 mg/ml).

**Storage/Stability**

Aggregation is thought to occur in the presence of high concentrations of 2-mercaptoethanol.

Lectin	MW (kDa)	Subunits	Specificity		Mitogenic Activity
			Blood Group	Sugar	
<i>Abrus precatorius</i>			–		+
Agglutinin	134	4		gal	
Abrin A (toxin)	60	2		gal	
Abrin B (toxin)	63.8	2( $\alpha\beta$ )		gal	
<i>Agarius bisporus</i>	58.5	–	–	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	
<i>Anguilla anguilla</i>	40	2	H	$\alpha$ -L-Fuc	
<i>Arachis hypogaea</i>	120	4	T	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	
<i>Artocarpus integrifolia</i>	42	4	T	$\alpha$ -gal $\rightarrow$ OME	+
<i>Bandeiraea simplicifolia</i>					
BS-I	114	4	A, B	$\alpha$ -gal, $\alpha$ -galNAc	
BS-I-A <sub>4</sub>	114	4	A	$\alpha$ -galNAc	
BS-I-B <sub>4</sub>	114	4	B	$\alpha$ -gal	
BS-II	113	4	acq, B, Tk, T	glcNAc	
<i>Bauhinia purpurea</i>	195	4	–	$\beta$ -gal(1 $\rightarrow$ 3)galNAc	+
<i>Caragana arborescens</i>	60; 120 <sup>a</sup>	2/4	–	galNAc	
<i>Cicer arietinum</i>	44	2	–	fetuin	
<i>Codium fragile</i>	60	4	–	galNAc	
<i>Concanavalin A</i>	102	4	–	$\alpha$ -man, $\alpha$ -glc	+
<i>Succinyl-Concanavalin A</i>	51	2	–	$\alpha$ -man, $\alpha$ -glc	+ <sup>b</sup>
<i>Cytisus scoparius</i>	–	–	–	galNAc, gal	
<i>Datura stramonium</i>	86	2( $\alpha\beta$ )	–	(glcNAc) <sub>2</sub>	
<i>Dolichos biflorus</i>	140	4	A <sub>1</sub>	$\alpha$ -galNAc	
<i>Erythrina corallodendron</i>	60	2	–	$\beta$ -gal(1 $\rightarrow$ 4)glcNAc	+
<i>Erythrina cristagalli</i>	56.8	2( $\alpha\beta$ )	–	$\beta$ -gal(1 $\rightarrow$ 4)glcNAc	
<i>Euonymus europaeus</i>	166	4( $\alpha\beta$ )	B, H	$\alpha$ -gal(1 $\rightarrow$ 3)gal	+
<i>Galanthus nivalis</i>	52	4	(h)	non-reduc. $\alpha$ -man	
<i>Glycine max</i>	110	4	–	galNAc	+ <sup>c</sup>
<i>Helix aspersa</i>	79	–	A	galNAc	
<i>Helix pomatia</i>	79	6	A	galNAc	
<i>Lathyrus odoratus</i>	40-43	4( $\alpha\beta$ )	–	$\alpha$ -man	+
<i>Lens culinaris</i>	49	2	–	$\alpha$ -man	+
<i>Limulus polyphemus</i>	400	18	–	NeuNAc	
Bacterial agglutinin	–	–	–	galNAc, glcNAc	
<i>Lycopersicon esculentum</i>	71	–	–	(glcNAc) <sub>3</sub>	
<i>Maackia amurensis</i>	130	2( $\alpha\beta$ )	O	sialic acid	+
<i>Maclura pomifera</i>	40-43	2( $\alpha\beta$ )	–	$\alpha$ -gal, $\alpha$ -galNAc	
<i>Momordica charantia</i>	115-129	4( $\alpha\beta$ )	–	gal, galNAc	
<i>Naja mocambique mocambique</i>	–	–	–	–	
<i>Naja naja kaouthia</i>	–	–	–	–	
<i>Narcissus pseudonarcissus</i>	26	2	(h)	$\alpha$ -D-man	
<i>Perseu americana</i>	–	–	–	–	
<i>Phaseolus coccineus</i>	112	4	–	–	
<i>Phaseolus limensis</i>	247(II)	8	A	galNAc	+
	124(III)	4			
<i>Phaseolus vulgaris</i>					
PHA-E	128	4	–	oligosaccharide	+
PHA-L	128	4	–	oligosaccharide	+
PHA-P					
PHA-M					

----- Table continued on next page -----

Lectin	MW (kDa)	Subunits	Specificity		Mitogenic Activity
			Blood Group	Sugar	
<i>Phytolacca americana</i>	32	–	–	(glcNAc) <sub>3</sub>	+
<i>Pisum sativum</i>	49	4(αβ)	–	α-man	+
<i>Pseudomonas aeruginosa PA-I</i>	13-13.7	–	–	gal	+ <sup>c</sup>
<i>Psophocarpus tetragonolobus</i>	35	1	–	galNAc, gal	
<i>Ptilota plumosa</i>	65; 170	–	B	α-gal	
<i>Ricinus communis</i>					
Toxin, RCA <sub>60</sub>	60	2	–	galNAc, β-gal	
Toxin, RCA <sub>120</sub>	120	4	–	β-gal	
<i>Sambucus nigra</i>	140	4(αβ)	–	αNeuNAC(2→6)gal galNAc	+ <sup>c</sup>
<i>Solanum tuberosum</i>	50; 100 <sup>a</sup>	1, 2	–	(glcNAc) <sub>3</sub>	
<i>Sophora japonica</i>	133	4	A, B	β-galNAc	
<i>Tetragonolobus purpureas</i>	120(A)	4	H	α-L-fuc	
	58(BA)	2	H	α-L-fuc	
	117(C)	4	H	α-L-fuc	
<i>Triticum vulgare</i>	36	2	–	(glcNAc) <sub>2</sub> , NeuNAc	+
<i>Ulex europaeus</i>					
UEA I	68	–	H	α-L-fuc	
UEA II	68	–	–	(glcNAc) <sub>2</sub>	
<i>Vicia faba</i>	50	4(αβ)	–	man, glc	+
<i>Vicia sativa</i>	40	4(αβ)	–	glc, man	+
<i>Vicia villosa</i>	139	4	A <sub>1</sub> +T <sub>n</sub>	galNAc	
A <sub>4</sub>	134	4	A <sub>1</sub>	galNAc	
B <sub>4</sub>	143	4	T <sub>n</sub>	galNAc	
<i>Vigna radiata</i>	160	4	–	α-gal	
<i>Viscum album</i>	115	4(αβ)	–	β-gal	
<i>Wisteria floribunda</i>	68	2	–	galNAc	

<sup>a</sup> Concentration-dependent molecular weight

<sup>b</sup> Non-agglutinating and mitogenic

<sup>c</sup> Mitogenic for neuraminidase-treated lymphocytes

## References

1. Rueben, L., et al., Activities of lectins and their immobilized derivatives in detergent solutions. Implications on the use of lectin affinity chromatography for the purification of membrane glycoproteins. *Biochemistry*, **16**, 1787-1794 (1977).
2. Chripeels, M. J. and Raikhel, N.V. *Lectin Reviews*, vol. 1, pp. 183-197, (New York 1992).
3. Schumacher, U., et al., *Lectin Reviews*, vol. 1, pp. 195-201, (New York 1992).

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