



# High sensitivity protein separations using a new size exclusion chromatography microcolumn for use in conjunction with MS

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# Introduction

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Recently, Tosoh Corporation introduced TSKgel SuperSW3000 columns in 1mm and 2mm ID microbore column format. Size exclusion chromatography (SEC) in an aqueous mobile phase is a powerful tool for analyzing biological polymers like peptides, proteins and DNA and TSKgel SW series SEC columns are routinely utilized for analyzing such biological samples. The ability to detect very small amounts of proteins is one requirement in proteomics. These new TSKgel SuperSW3000 microbore columns with increased resolution, excellent sensitivity and high recovery were developed to analyze trace amounts of proteins.



# Preliminary Results

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The TSKgel SuperSW3000, 4 $\mu$ m micro-columns were characterized by analyzing protein resolution, detection sensitivity, sample capacity, and column efficiency in comparison to conventional column sizes. A 5-fold increase in peak height of a standard protein mixture was obtained when using a 2mm ID x 30cm TSKgel SuperSW3000 column, compared to a 4.6mm ID x 30cm column. The same improvement in sensitivity was also evident when analyzing aggregate-containing IgG samples. Linear calibration curves confirmed that nonspecific adsorption on the stationary phase was minimal. The detection limit of IgG was 18ng using the 1mm ID TSKgel SuperSW3000 column while still being able to detect small amounts of IgG aggregates. Unlike larger particle size columns, four micron SuperSW3000 columns showed a smaller drop in efficiency when increasing flow rate. As with 1mm ID columns, we found that reducing the injection volume of IgG solution from 10 $\mu$ L to 1 $\mu$ L greatly improved efficiency of the 2mm ID column, although at constant injection volume, efficiency did not vary with IgG concentration in the range of 1-5g/L. Results showed that trace analysis of biological components was possible when the TSKgel SuperSW3000 1mm ID column was utilized with an off-line SELDI/TOF/MS.



# Experimental Conditions

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## Columns

- TSKgel SuperSW3000 from Tosoh
  - Internal diameter: 1mm, 2mm & 4.6mm
  - Length: 30cm
  - Particle size: ca. 4 $\mu$ m
  - Exclusion limit: 5 x 10<sup>5</sup> Dalton
  - Matrix: Diol-bonded silica gel
- KW803-2E, Shodex
- Superdex™ 200 PC, GE Healthcare

## Instrumentation

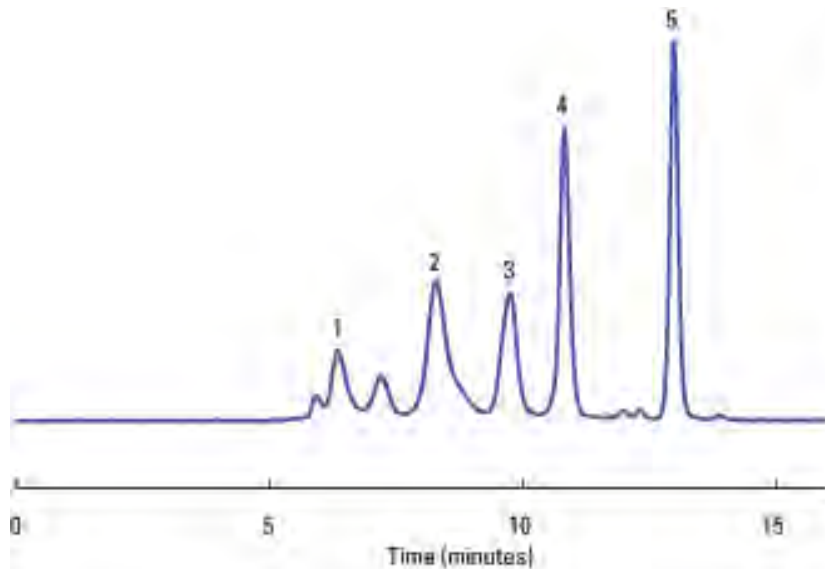
- Pump: DP-8020 (Tosoh)
- Detector: UV-8020 (Tosoh)
- UV cell: 2 $\mu$ L (for 2mm & 4.6mm ID, Tosoh)
- UV cell: 35nL (for 1mm ID, LC Packings, Netherlands)
- Sample injector: Model 7520 (Rheodyne)
- Tubing (injector to column):  
0.05mm ID x 20cm Fused Silica
- Data processing: LC-8020 model 2 (Tosoh)

## Sample

Proteins and enzymes were purchased from Sigma (USA). The antibody was a gift from the Tosoh Research Center (Kanagawa, Japan).



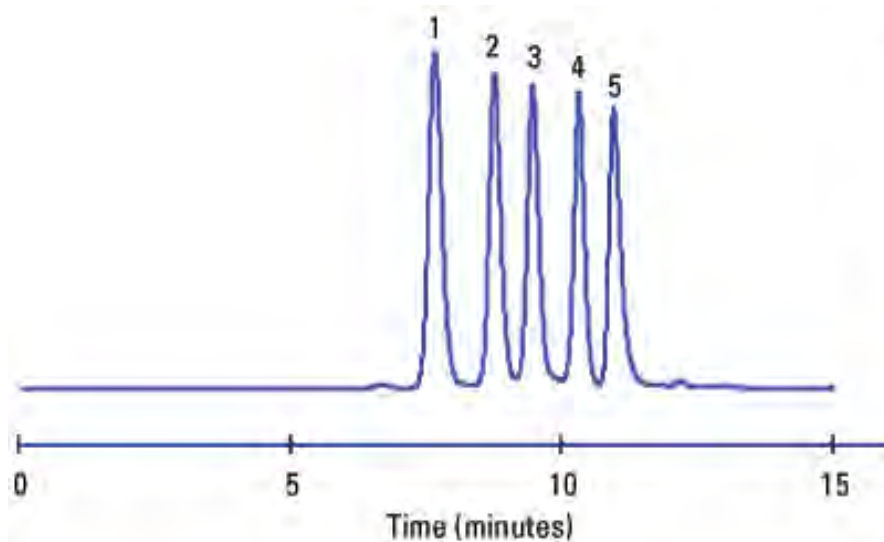
# Figure 1: Separation of Standard Proteins on a 1mm ID TSKgel SuperSW3000 Column



**Column:** TSKgel SuperSW3000, 1mm ID x 30cm  
**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)  
**Flow rate:** 16 $\mu\text{L}/\text{min}$   
**Detection:** UV@280nm  
**Temperature:** 25°C  
**Injection volume:** 0.2 $\mu\text{L}$   
**Samples:**  
1. thyroglobulin (1.0mg/mL)  
2.  $\gamma$ -globulin (2.0mg/mL)  
3. ovalbumin (2.0mg/mL)  
4. ribonuclease A (3.0mg/mL)  
5. p-aminobenzoic acid (0.02mg/mL)



## Figure 2: Separation of Standard Proteins on a 1mm ID TSKgel SuperSW3000 Column

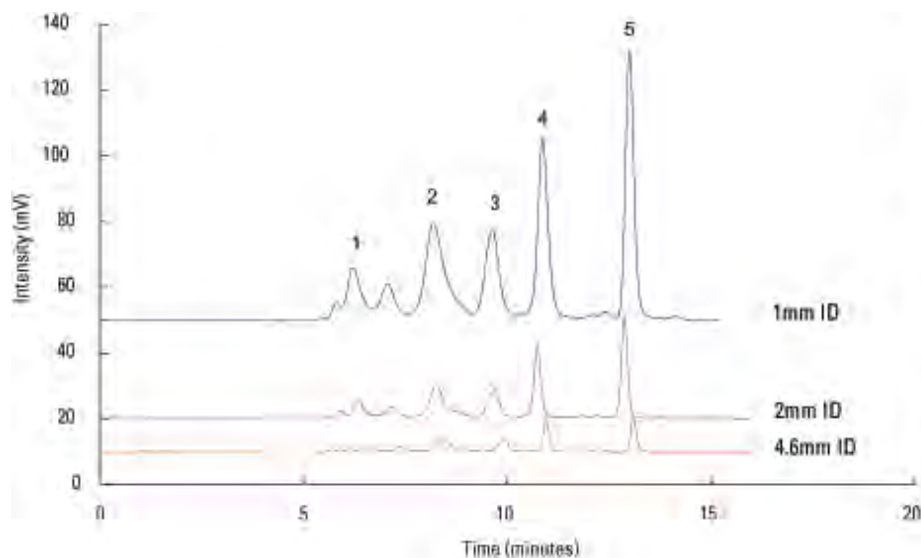


Column: TSKgel SuperSW3000, 1mm ID x 30cm  
Eluent: 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)  
Flow rate: 16 $\mu\text{L}/\text{min}$   
Detection: UV@280nm  
Temperature: 25°C  
Injection volume: 0.2 $\mu\text{L}$   
Samples: 1. glutamate dehydrogenase  
2. lactate dehydrogenase  
3. enolase  
4. myokinase  
5. cytochrome C

MW Marker 1 vial/100 $\mu\text{L}$ , Oriental Yeast Co.



# Figure 3: Sensitivity Comparison on TSKgel SuperSW3000 Columns



**Columns:** TSKgel SuperSW3000, 4.6mm ID x 30cm  
TSKgel SuperSW3000, 2mm ID x 30cm  
TSKgel SuperSW3000, 1mm ID x 30cm

**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)

**Flow rate:** 0.350mL/min (4.6mm ID)  
0.065mL/min (2mm ID)  
0.016mL/min (1mm ID)

**Detection:** UV@280nm

**Detector cell volume:** 2 $\mu\text{L}$  (4.6 and 2mm ID)  
35nL (1mm ID)

**Temperature:** 25°C

**Injection volume:** 1 $\mu\text{L}$

**Samples:** 1. thyroglobulin (1.0mg/mL)  
2.  $\gamma$ -globulin (2.0mg/mL)  
3. ovalbumin (2.0mg/mL)  
4. ribonuclease A (3.0mg/mL)  
5. p-aminobenzoic acid (0.02mg/mL)



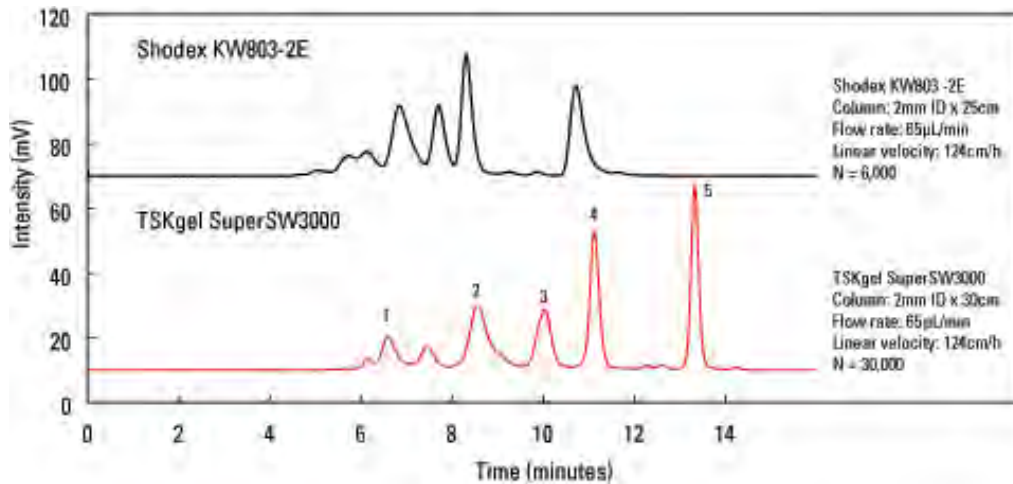
## Table 1: Comparison of 2mm ID TSKgel SuperSW3000 With Other SEC Columns

	Dimensions (mm ID x cm)	Particle size	Material	Exclusion limit
TSKgel SuperSW3000	2 x 30	4 $\mu$ m	Diol-bonded silica gel	5 x 10 <sup>5</sup> Da
KW803-2E	2 x 25	5 $\mu$ m	Silica gel	7 x 10 <sup>5</sup> Da
Superdex™ 200 PC	3.2 x 30	13 $\mu$ m	Dextran & agarose	6 x 10 <sup>5</sup> Da





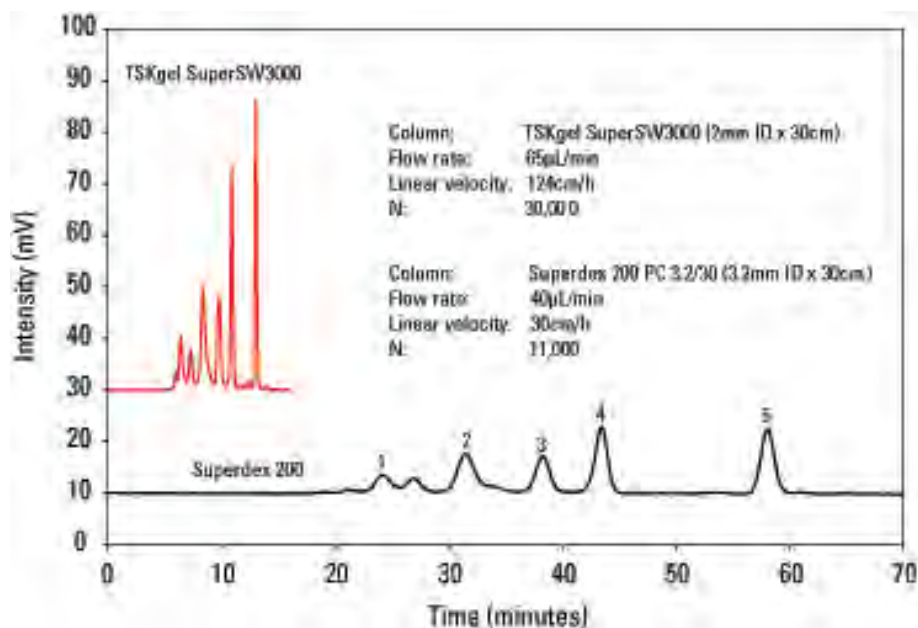
# Figure 4: Separation of Standard Proteins on Commercial GFC Columns



**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)  
**Detection:** UV@280nm  
**Temperature:** 25°C  
**Injection volume:** 0.2µL  
**Samples:**  
1. thyroglobulin (1.0mg/mL)  
2.  $\gamma$ -globulin (2.0mg/mL)  
3. ovalbumin (2.0mg/mL)  
4. ribonuclease A (3.0mg/mL)  
5. p-aminobenzoic acid (0.02mg/mL)



# Figure 5: Separation of Standard Proteins on Commercial GFC Columns



**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)

**Detection:** UV@280nm

**Temperature:** 25°C

**Injection volume:** 0.2µL

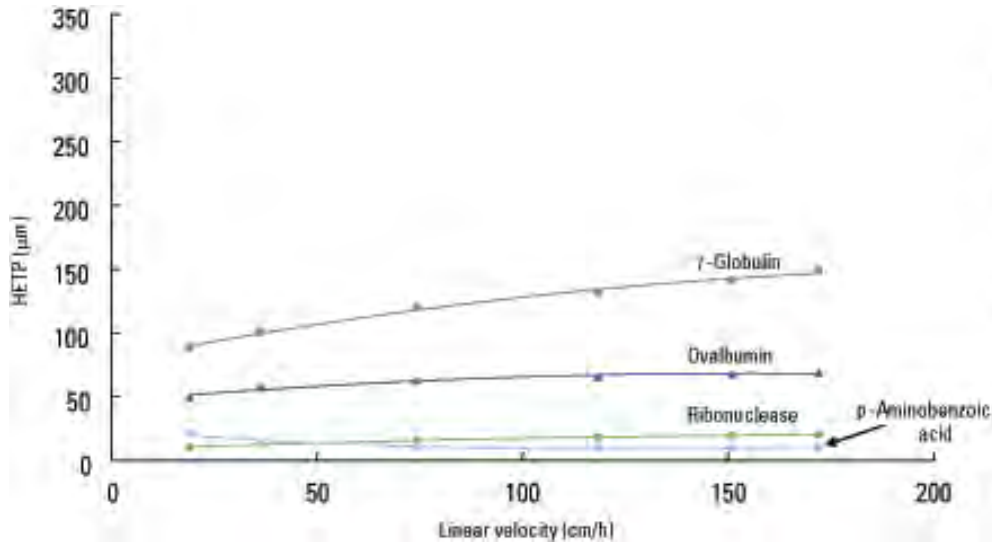
**Samples:**

1. thyroglobulin (1.0mg/mL)
2.  $\gamma$ -globulin (2.0mg/mL)
3. ovalbumin (2.0mg/mL)
4. ribonuclease A (3.0mg/mL)
5. p-aminobenzoic acid (0.02mg/mL)

Note: both columns were operated at their recommended flow rates.



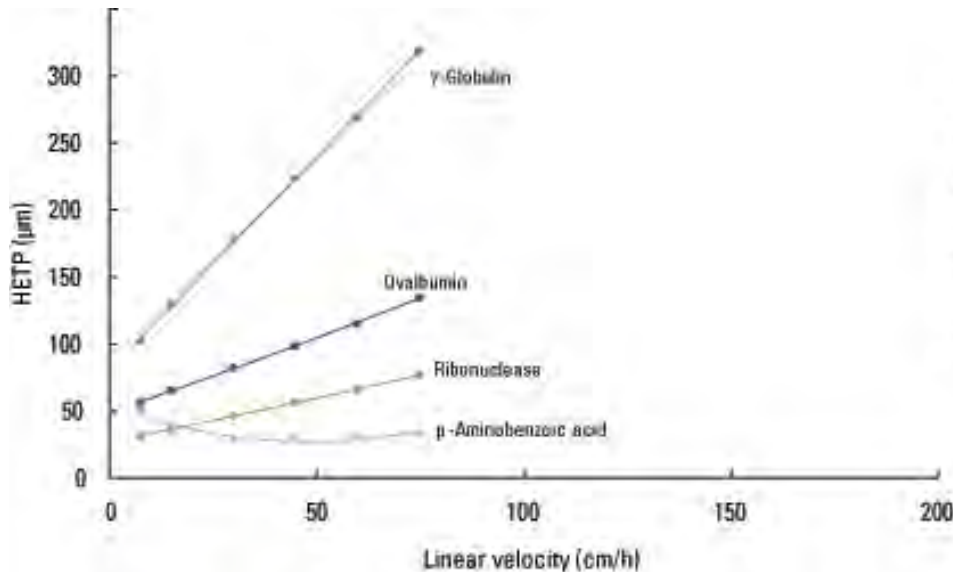
# Figure 6: Relationship Between Column Efficiency and Linear Velocity



Column: TSKgel SuperSW3000, 2mm ID x 30cm  
Eluent: 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)  
Flow rate: 16 $\mu\text{L}/\text{min}$   
Detection: UV@280nm  
Temperature: 25°C  
Injection volume: 0.2 $\mu\text{L}$   
Samples:  $\gamma$ -globulin (2.0mg/mL)  
ovalbumin (2.0mg/mL)  
ribonuclease A (3.0mg/mL)  
p-aminobenzoic acid (0.02mg/mL)



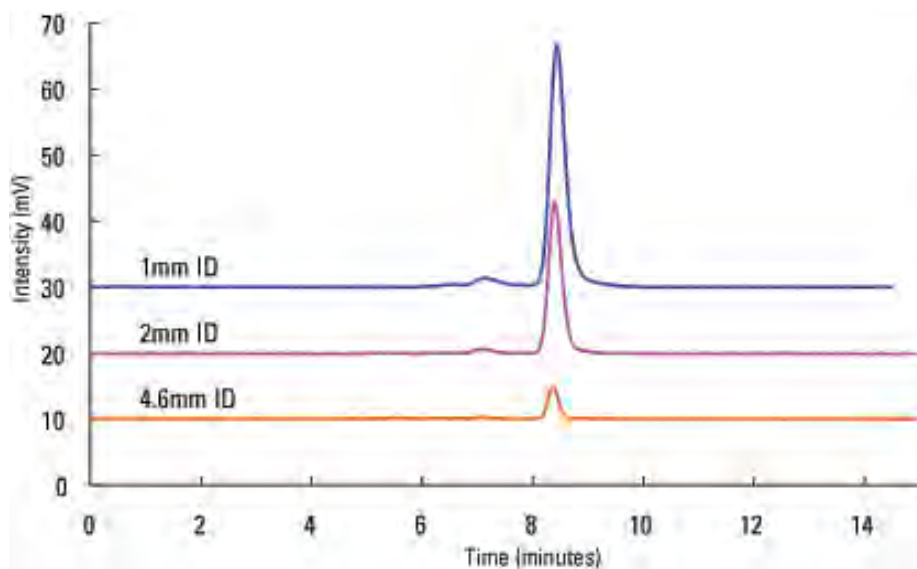
# Figure 7: Relationship Between Column Efficiency and Linear Velocity



Column: Superdex 200 PC 3.2/30, 3.2mm ID x 30cm  
Eluent: 0.1mol/L phosphate buffer + 0.1mol/L Na<sub>2</sub>SO<sub>4</sub> + 0.05% NaN<sub>3</sub> (pH 6.7)  
Flow rate: 16µL/min  
Detection: UV@280nm  
Temperature: 25°C  
Injection volume: 0.2µL  
Samples: γ-globulin (2.0mg/mL)  
ovalbumin (2.0mg/mL)  
ribonuclease A (3.0mg/mL)  
p-aminobenzoic acid (0.02mg/mL)



## Figure 8: Separation of IgG on TSKgel SuperSW3000 Columns



**Columns:** TSKgel SuperSW3000, 4.6mm ID x 30cm  
TSKgel SuperSW3000, 2mm ID x 30cm  
TSKgel SuperSW3000, 1mm ID x 30cm

**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)

**Flow rate:** 0.350mL/min (4.6mm ID)  
0.065mL/min (2mm ID)  
0.016mL/min (1mm ID)

**Detection:** UV@280nm

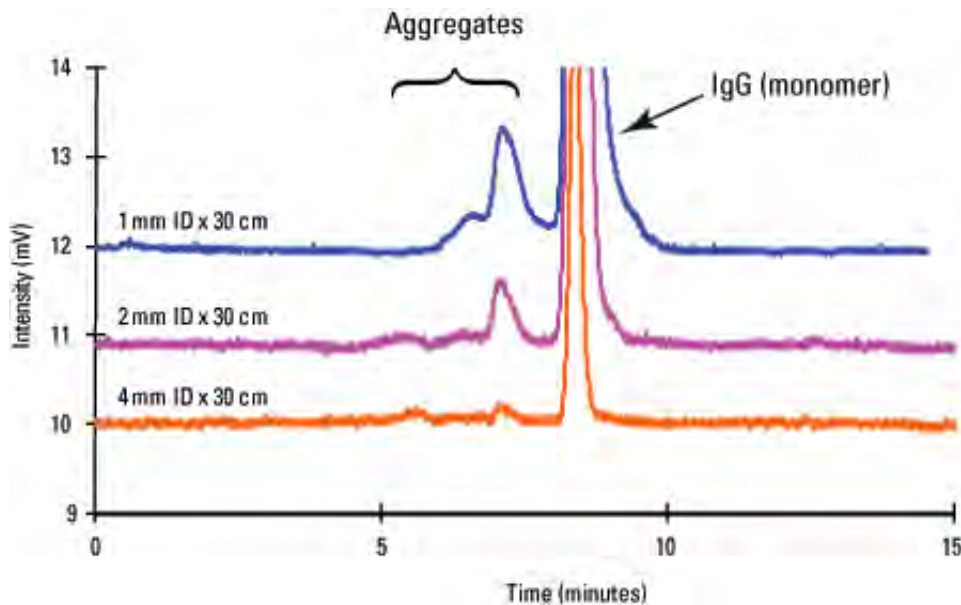
**Temperature:** 25°C

**Injection volume:** 1 $\mu$ L

**Sample:** IgG (mouse, MAb, 1mg/mL)



# Figure 9: Separation of IgG Sample on TSKgel SuperSW3000 Columns



**Columns:** TSKgel SuperSW3000, 4.6mm ID x 30cm  
TSKgel SuperSW3000, 2mm ID x 30cm  
TSKgel SuperSW3000, 1mm ID x 30cm

**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L Na<sub>2</sub>SO<sub>4</sub> + 0.05% NaN<sub>3</sub> (pH 6.7)

**Flow rate:** 0.350mL/min (4.6mm ID)  
0.065mL/min (2mm ID)  
0.016mL/min (1mm ID)

**Detection:** UV@280nm

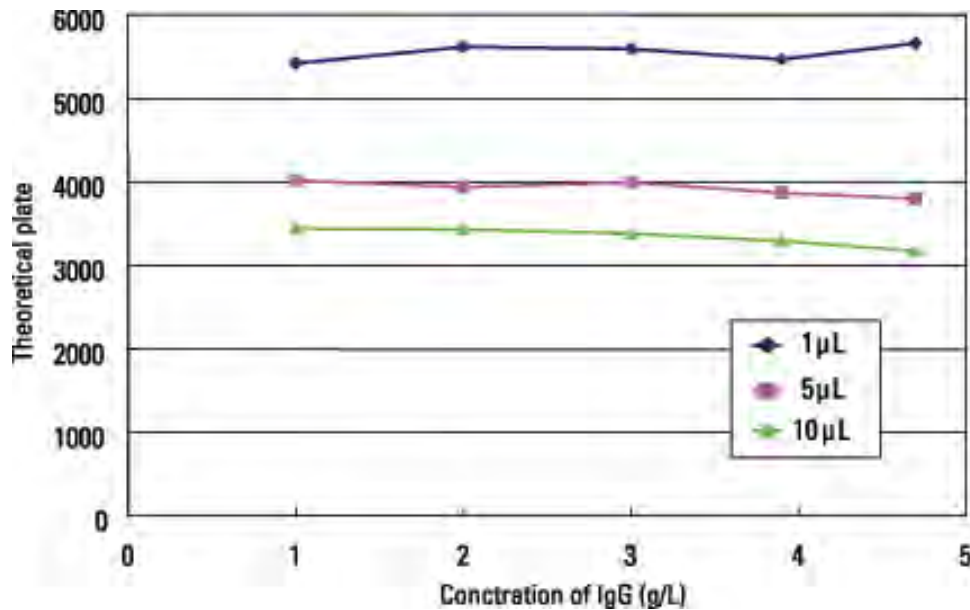
**Temperature:** 25°C

**Injection volume:** 1µL

**Sample:** IgG (mouse, MAb, 1mg/mL)



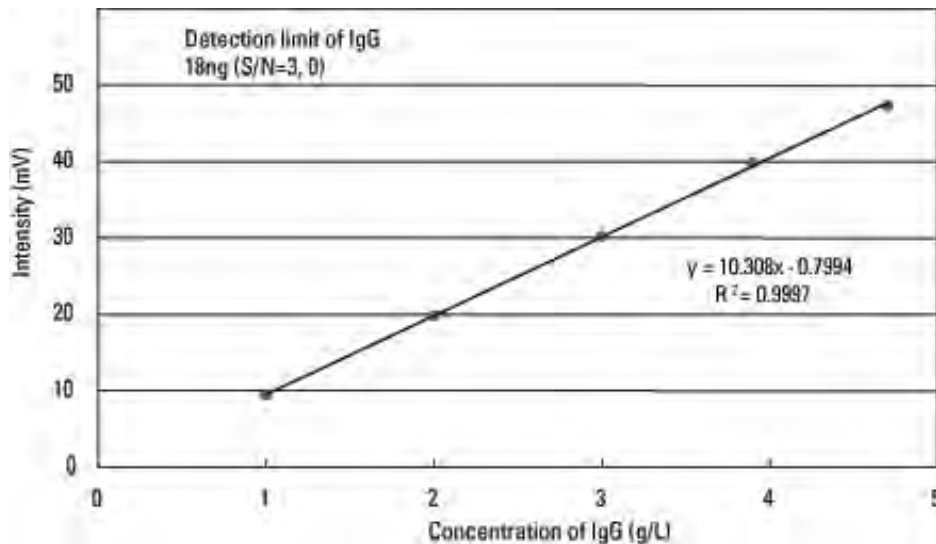
# Figure 10: IgG Loading Study on a 2mm ID TSKgel SuperSW3000 Column



**Column:** TSKgel SuperSW3000, 2mm ID x 30cm  
**Eluent:** 0.1mol/L phosphate buffer + 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$  (pH 6.7)  
**Flow rate:** 0.350mL/min (4.6mm ID)  
0.065mL/min (2mm ID)  
0.016mL/min (1mm ID)  
**Detection:** UV@280nm  
**Temperature:** 25°C  
**Sample:** IgG (mouse, MAb)



# Figure 11: Calibration Curve of IgG on a 1mm ID TSKgel SuperSW3000 Column

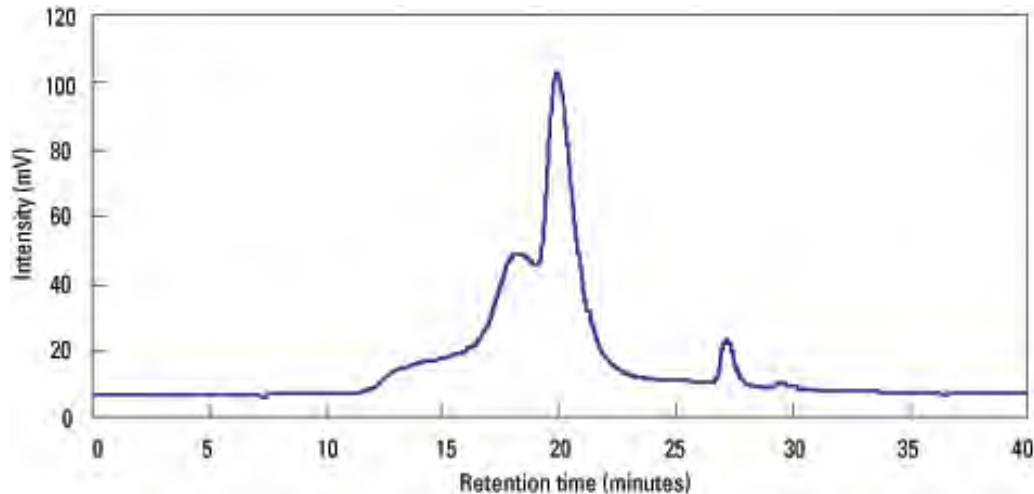


Columns: TSKgel SuperSW3000, 1mm ID x 30cm  
Eluent: 0.1mol/L phosphate buffer + 0.1mol/L Na<sub>2</sub>SO<sub>4</sub> + 0.05% NaN<sub>3</sub> (pH 6.7)  
Flow rate: 0.016mL/min  
Detection: UV@280nm  
Temperature: 25°C  
Injection volume: 0.2µL  
Samples: IgG (mouse, MAb)





# Figure 12a: Separation of Human Serum Proteins on a 1mm ID TSKgel SuperSW3000 Column



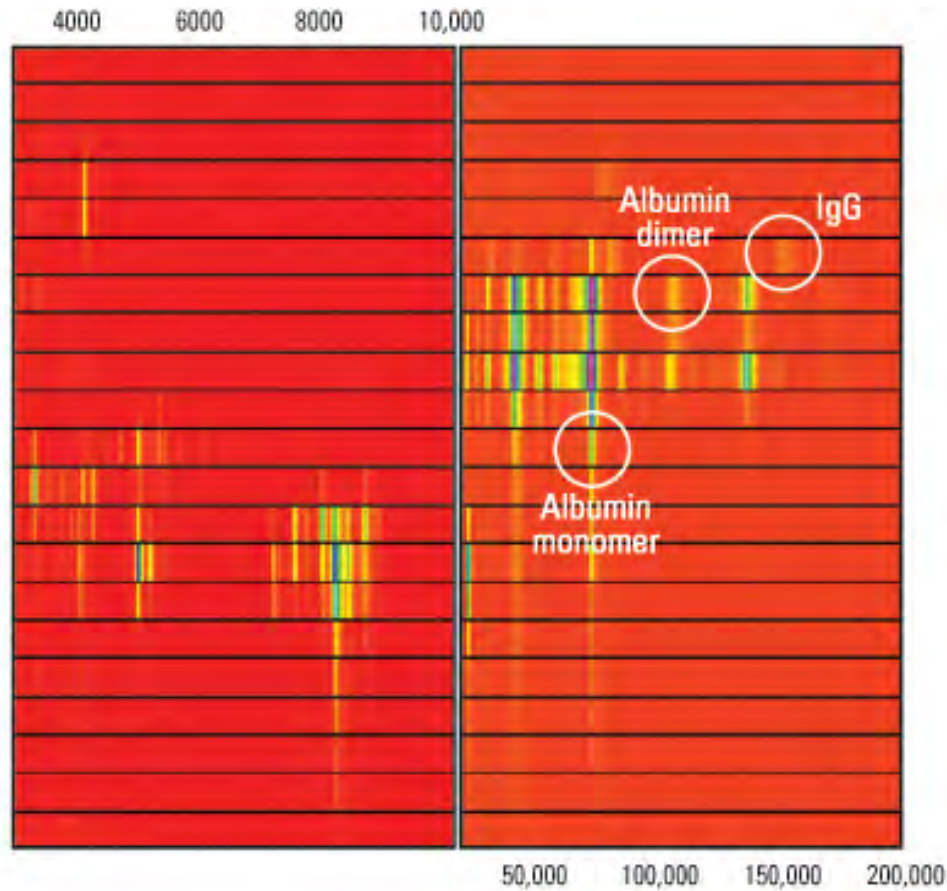
Column: TSKgel SuperSW3000, 1mm ID x 30cm  
Eluent: 50mmol/L  $\text{NaH}_2\text{PO}_4$  + 0.5mol/L NaCl (pH 7.0)  
Flow rate: 8 $\mu\text{L}/\text{min}$   
Detection: UV@280nm  
Temperature: ambient  
Sample: human serum (x 10), 1 $\mu\text{L}$

Fraction (1mL) was directly loaded to SELDI chip H50.  
The chip was washed and desalted then applied to MS.

This data is courtesy of Dr. Majima, Protenova.



# Figure 12b: Separation of Human Serum Proteins on a 1mm ID TSKgel SuperSW3000 Column



Fraction of interest analyzed by off-line SELDI/TOF/MS to establish presence of BSA aggregates and IgG.



# Conclusions

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- (1) TSKgel SuperSW3000 microbore columns (1mm ID and 2mm ID) showed high resolution power for biological samples similar to what can be obtained on 4.6mm ID conventional TSKgel SuperSW3000 columns.
- (2) Despite the high concentration of IgG (ca. 5mg/mL), good separation was achieved.
- (3) High sensitivity analysis could be achieved on the microbore columns. Linear calibration curves confirmed that nonspecific adsorption on the stationary phase was minimal. The detection limit of IgG was 18ng using a 1mm ID column while still being able to detect small amounts of IgG aggregates.
- (4) The results showed that TSKgel SuperSW3000 microbore columns are an excellent choice for the rapid separation of proteins and enzymes at micro scale and are a great fit for the trace analysis of biological components by LC/MS.

**January, 2013**

**Message for Sigma-Aldrich/Supelco customers**

**Sigma-Aldrich Corporation is a non-exclusive distributor of TSKgel columns and TOYOPEARL bulk resins (< 1 L) in North, South and Central America, Europe, Africa and the Middle East.**

**For the availability of TSKgel columns in Asia and Australia, please consult the website of Tosoh Corporation.**