

## User Guide

# MILLIPLEX® Mouse Th17 Magnetic Bead Panel

## 96-Well Plate Assay

**MTH17MAG-47K**  
**MT17MAG47K-PX25**  
**MT17MAG47PMX25BK**

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## Introduction

CD4+T-helper cells are major players in adaptive immunity. They can be divided into the major subsets Th1, Th2, Th17 and Treg based on their expression profile of transcription factors and secreted cytokines. Th1 cells, induced by IL-12 and enhanced by IFN $\gamma$ , secrete IFN $\gamma$  for activating macrophages. Th2 cells, induced by IL-4, produce mainly IL-4, IL-5, IL-13, IL-25/IL-17E and IL-31 and play a role in allergy. Treg cells, induced by TGF $\beta$ , produce the anti-inflammatory cytokine IL-10 and also TGF $\beta$  and function to control T-cell responses to prevent autoimmune reactions. Th17 cells, a more recently discovered subset characterized by the production of IL-17, are induced by TGF $\beta$ , IL-6 and IL-21 in mice or TGF $\beta$  combined with IL-23 and IL-1 $\beta$  in humans. Activated Th17 cells secrete IL-17A, IL-17F, IL-6, IL-21, IL-22 and TNF $\alpha$  which promote tissue inflammation.

Th17 cells are involved in the clearance of extracellular bacteria and fungi. They are abundant in the intestinal lamina propria and function as a barrier against invading pathogens.

Excessive amounts of Th17 cells have been implicated in the pathogenesis of several autoimmune diseases such as multiple sclerosis, psoriasis, autoimmune uveitis, juvenile diabetes, rheumatoid arthritis, and Crohn's disease. In addition, Th17 cells play an important role in cancer development and progression, potentially playing a role in both promoting and inhibiting tumor growth.

The MILLIPLEX<sup>®</sup> portfolio offer the broadest selection of analytes across a wide range of disease states and species. Once the analytes of interest have been identified, you can rely on the quality that we build into each kit to produce results you can trust. In addition to the assay characteristics listed in the protocol, other performance criteria evaluated during the verification process include: cross-reactivity, dilution linearity, kit stability, and sample behavior (for example, detectability and stability).

Each MILLIPLEX<sup>®</sup> panel and kit includes:

- Quality controls (QCs) provided to qualify assay performance
- Comparison of standard (calibrator) and QC lots to a reference lot to ensure lot-to-lot consistency
- Optimized serum matrix to mimic native analyte environment
- Detection antibody cocktails designed to yield consistent analyte profiles within panel

In addition, each panel and kit meets stringent manufacturing criteria to ensure batch-to-batch reproducibility. The MILLIPLEX<sup>®</sup> Mouse Th17 Magnetic Bead Panel thus enables you to focus on the therapeutic potential of the modulation of CD4+T-helper cell cytokines. Coupled with the Luminex<sup>®</sup> xMAP<sup>®</sup> platform in a magnetic bead format, you receive the advantage of ideal speed and sensitivity, allowing quantitative multiplex detection of dozens of analytes simultaneously, which can dramatically improve productivity.

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The MILLIPLEX® Mouse Th17 Magnetic Bead Panel is part of the most versatile system available for CD4+T-helper cell cytokine research. From our single to multiplex biomarker solutions, we partner with you to design, develop, analytically verify and build the most comprehensive library available for protein detection and quantitation.

MILLIPLEX® products offer you:

- The ability to select a 25-plex premixed option or
- The ability to choose any combination of analytes from our panel of 25 analytes to design a custom kit that better meets your needs.
- A convenient “all-in-one” box format that gives you the assurance that you will have all the necessary reagents you need to run your assay.

The MILLIPLEX® Mouse Th17 Magnetic Bead Panel is a 25-plex kit to be used for the simultaneous quantification of any or all of the following murine cytokines in serum, plasma, tissue/cell lysate and culture supernatant samples: IL-1 $\beta$ , IL-2, IL-4, IL-5, IL-6, IL-10, IL-12p70, IL-13, IL-15, IL-17A, IL-25/IL-17E, IL-17F, IL-21, IL-22, IL-23, IL-27, IL-28B, IL-31, IL-33, CD40L, GM-CSF, IFN $\gamma$ , MIP-3 $\alpha$ , TNF $\alpha$ , TNF $\beta$ .

**This kit is for research purposes only. Not for use in diagnostic procedures. Please read the entire protocol before use. It is important to use same assay incubation conditions throughout your study.**

## Principle

MILLIPLEX® products are based on the Luminex® xMAP® technology — one of the fastest growing and most respected multiplex technologies offering applications throughout the life-sciences and capable of performing a variety of bioassays including immunoassays on the surface of fluorescent-coded magnetic beads known as MagPlex®-C microspheres.

- Luminex® products use proprietary techniques to internally color-code microspheres with two fluorescent dyes. Through precise concentrations of these dyes, distinctly colored bead sets of 500-5.6  $\mu$ m polystyrene microspheres or 80-6.45  $\mu$ m magnetic microspheres can be created, each of which is coated with a specific capture antibody.
- After an analyte from a test sample is captured by the bead, a biotinylated detection antibody is introduced.
- The reaction mixture is then incubated with Streptavidin-PE conjugate, the reporter molecule, to complete the reaction on the surface of each microsphere.

- The following Luminex® instruments can be used to acquire and analyze data using two detection methods:
  - The Luminex® analyzers, Luminex® 200™, FLEXMAP 3D® and xMAP® INTELLIFLEX, are flow cytometry-based instruments that integrate key xMAP® detection components, such as lasers, optics, advanced fluidics and high-speed digital signal processors.
  - The Luminex® analyzer (MAGPIX®), a CCD-based instrument that integrates key xMAP® capture and detection components with the speed and efficiency of magnetic beads.
- Each individual microsphere is identified and the result of its bioassay is quantified based on fluorescent reporter signals. We combine the streamlined data acquisition power of Luminex® xPONENT® acquisition software with sophisticated analysis capabilities of the new MILLIFLEX® Analyst 5.1, integrating data acquisition and analysis seamlessly with all Luminex® instruments.
- xMAP® INTELLIFLEX runs on INTELLIFLEX software for instrument control, run setup and generating high quality data with flexible output options. Data can be exported in xPONENT® style CSV files for compatibility with many existing analytical applications, or in the new, customizable INTELLIFLEX file format. The INTELLIFLEX file format is intended for flexibility and simplicity, allowing the user to freely select which data points to include and to reduce the time to analysis.

The capability of adding multiple conjugated beads to each sample results in the ability to obtain multiple results from each sample. Open-architecture xMAP® technology enables multiplexing of many types of bioassays reducing time, labor and costs over traditional methods.

## Storage Conditions Upon Receipt

- Recommended storage for kit components is 2–8 °C.
- For long-term storage, freeze reconstituted standards and controls at  $\leq -20$  °C. Avoid multiple ( $> 2$ ) freeze/thaw cycles.
- **DO NOT FREEZE** Antibody-Immobilized Beads, Detection Antibody, and Streptavidin-Phycoerythrin.

## Reagents Supplied

Store all reagents at 2–8 °C

Reagents Supplied	Volume	Quantity	Cat. No.
Mouse Th17 Standard	Lyophilized	1 vial	MTH17-8047
Mouse Th17 Quality Controls 1 and 2	Lyophilized	2 vials	MTH17-6047
Serum Matrix*	Lyophilized	1 vial	MXMSM
Set of one 96-Well Plate with 2 sealers	-	1 set	-
Assay Buffer	30 mL	1 bottle	L-AB
10X Wash Buffer**	60 mL	1 bottle	L-WB
Mouse Th17 Detection Antibodies	3.2 mL	1 bottle	MTH17-1047
Streptavidin-Phycoerythrin	3.2 mL	1 bottle	L-SAPE3
Mixing Bottle (not provided with premixed panel)	-	1 bottle	-

\* Contains 0.08% Sodium azide

\*\* Contains 0.05% Proclin

### Mouse Th17 Antibody-Immobilized Premixed Magnetic Beads

	Volume	Quantity	Cat. No.
Premixed 25-plex Beads	3.5 mL	1 bottle	MTH17PMX25-MAG

Included Mouse Th17 Antibody-Immobilized Magnetic Beads are dependent on customizable selection of analytes within the panel.

## Mouse Th17 Antibody-Immobilized Magnetic Beads

Bead/Analyte Name	Luminex® Magnetic Bead Region	Customizable 25 Analytes (50X concentration, 90 µL)	
		Available	Cat. No.
Anti-Mouse IL-25/IL-17E Bead	14	✓	MIL17E-MAG
Anti-Mouse GM-CSF Bead	15	✓	MGMCSF-MAG
Anti-Mouse IFN $\gamma$ Bead	19	✓	MIFNG-MAG
Anti-Mouse MIP-3 $\alpha$ /CCL20 Bead	20	✓	MTHMIP3A-MAG
Anti-Mouse IL-1 $\beta$ Bead	25	✓	MIL1B-MAG
Anti-Mouse IL-2 Bead	26	✓	MIL2-MAG
Anti-Mouse IL-4 Bead	28	✓	MIL4-MAG
Anti-Mouse IL-5 Bead	30	✓	MIL5-MAG
Anti-Mouse IL-6 Bead	34	✓	MCYIL6-MAG
Anti-Mouse IL-21 Bead	35	✓	MIL21-MAG
Anti-Mouse IL-22 Bead	37	✓	MIL22-MAG
Anti-Mouse IL-28B Bead	42	✓	MIL28B-MAG
Anti-Mouse IL-10 Bead	43	✓	MIL10-MAG
Anti-Mouse IL-23 Bead	46	✓	MIL23-MAG
Anti-Mouse IL-12p70 Bead	47	✓	MIL12P70-MAG
Anti-Mouse IL-27 Bead	48	✓	MIL27-MAG
Anti-Mouse IL-13 Bead	52	✓	MIL13-MAG
Anti-Mouse IL-15 Bead	54	✓	MIL15-MAG
Anti-Mouse IL-17A Bead	56	✓	MIL17A-MAG
Anti-Mouse IL-17F Bead	61	✓	MIL17F-MAG
Anti-Mouse IL-33 Bead	67	✓	MIL33-MAG
Anti-Mouse IL-31 Bead	72	✓	MIL31-MAG
Anti-Mouse TNF $\beta$ Bead	73	✓	MTNFB-MAG
Anti-Mouse TNF $\alpha$ Bead	77	✓	MCYTNFA-MAG
Anti-Mouse CD40L Bead	78	✓	MCD40L-MAG

## Materials Required (not included)

### Reagents

MAGPIX® Drive Fluid PLUS (Cat. No. 40-50030), xMAP® Sheath Fluid PLUS (Cat. No. 40-50021), or xMAP® Sheath Concentrate PLUS (Cat. No. 40-50023)

### Instrumentation/Materials








- Adjustable pipettes with tips capable of delivering 25 µL to 1000 µL
- Multichannel pipettes capable of delivering 5 µL to 50 µL, or 25 µL to 200 µL
- Reagent reservoirs
- Polypropylene microfuge tubes
- Rubber bands
- Aluminum foil
- Absorbent pads
- Laboratory vortex mixer
- Sonicator (Branson Ultrasonic Cleaner Model B200 or equivalent)
- Titer plate shaker (VWR® Microplate Shaker Cat. No. 12620-926 or equivalent)
- Luminex® 200™, HTS, FLEXMAP 3D®, MAGPIX® instrument with xPONENT® software, or xMAP® INTELLIFLEX instrument with INTELLIFLEX software by Luminex® Corporation
- Automatic plate washer for magnetic beads (BioTek® 405 LS and 405 TS, Cat. No. 40-094, 40-095, 40-096, 40-097 or equivalent) or Handheld Magnetic Separation Block (Cat. No. 40-285 or equivalent).

**Note:** If a plate washer or handheld magnetic separation block for magnetic beads is not available, one can use a microtiter filter plate (Cat. No. MX-PLATE) to run the assay using a vacuum filtration unit (Vacuum Manifold, Cat. No. MSVMHTS00 or equivalent with Vacuum Pump, Cat. No. WP6111560 or equivalent).

## Safety Precautions

- All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing of infectious agents.
- Sodium azide or Proclin has been added to some reagents as a preservative. Although the concentrations are low, Sodium azide and Proclin may react with lead and copper plumbing to form highly explosive metal azides. Dispose of unused contents and waste in accordance with international, federal, state, and local regulations.

## Symbol Definitions

Ingredient	Cat. No.	Label	
Mouse Th17 Detection Antibodies	MTH17-1047		<b>Warning.</b> Causes serious eye irritation. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
Mouse Th17 Quality Controls 1 & 2	MTH17-6047	 	<b>Danger.</b> Harmful if swallowed. Causes serious eye damage. Harmful to aquatic life with long lasting effects. Avoid release to the environment. Wear eye protection. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. Get medical advice/attention.
Mouse Th17 Standard	MTH17-8047	 	<b>Danger.</b> Harmful if swallowed. Causes serious eye damage. Harmful to aquatic life with long lasting effects. Avoid release to the environment. Wear eye protection. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. Get medical advice/attention.
Serum Matrix	MXMSM	No Symbol Required	Harmful to aquatic life with long lasting effects. Avoid release to the environment.
Streptavidin-Phycoerythrin	L-SAPE3		<b>Warning.</b> Causes serious eye irritation. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
Wash Buffer	L-WB		<b>Warning.</b> May cause an allergic skin reaction. Wear protective gloves. IF ON SKIN: Wash with plenty of soap and water.



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## Technical Guidelines

To obtain reliable and reproducible results, the operator should carefully read this entire manual and fully understand all aspects of each assay step before running the assay. The following notes should be reviewed and understood before the assay is set up.

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- Do not use beyond the expiration date on the label.
- Do not mix or substitute reagents with those from other lots or sources.
- The Antibody-Immobilized Beads are light sensitive and must be protected from light at all times. Cover the assay plate containing beads with opaque plate lid or aluminum foil during all incubation steps.
- It is important to allow all reagents to warm to room temperature (20-25 °C) before use in the assay.
- Incomplete washing can adversely affect the assay outcome. All washing must be performed with the Wash Buffer provided.
- After hydration, all standards and controls may be kept in their polypropylene tubes.
- The standards prepared by serial dilution must be used within 1 hour of preparation. Discard any unused standards except the standard stock which may be stored at  $\leq -20$  °C for 1 month and at  $\leq -80$  °C for greater than one month.
- If samples fall outside the dynamic range of the assay, further dilute the samples with the appropriate diluent and repeat the assay.
- Any unused mixed Antibody-Immobilized Beads may be stored in the Mixing Bottle at 2-8 °C for up to one month.
- During the preparation of the standard curve, make certain to mix the higher concentration well before making the next dilution. Use a new tip with each dilution.
- The plate should be read immediately after the assay is finished. If, however, the plate cannot be read immediately, seal the plate, cover with aluminum foil or an opaque lid, and store the plate at 2-8 °C for up to 24 hours. Prior to reading, agitate the plate on the plate shaker at room temperature for 10 minutes. Delay in reading a plate may result in decreased sensitivity for some analytes.
- The titer plate shaker should be set at a speed to provide maximum orbital mixing without splashing of liquid outside the wells. For the recommended plate shaker, this would be a setting of 5-7 which is approximately 500-800 rpm.
- Ensure that the needle probe is clean. This may be achieved by sonication and/or alcohol flushes.

- When reading the assay on the Luminex® 200™ instrument, adjust probe height according to the protocols recommended by Luminex® to the kit solid plate or to the recommended filter plates using 3 alignment discs. When reading the assay on the MAGPIX® instrument, adjust probe height according to the protocols recommended by Luminex® to the kit solid plate or to the recommended filter plates using 2 alignment discs. When reading the assay on the FLEXMAP 3D® instrument, adjust probe height according to the protocols recommended by Luminex® to the kit solid plate using 1 alignment disc.
- For the FLEXMAP 3D® instrument, when using the solid plate in the kit, the final resuspension should be with 150 µL Sheath Fluid PLUS in each well and 75 µL should be aspirated.
- For xMAP® INTELLIFLEX, adjust probe height based on the type of plate you are using, place an alignment disk or an alignment sphere in the well according to the protocol recommended by Luminex®.
- For cell culture supernatants or tissue extraction, use the culture or extraction medium as the matrix solution in background, standard curve and control wells. If samples are diluted in assay buffer, use the assay buffer as matrix.
- For serum/plasma samples that require dilution, use the Serum Matrix provided in the kit.
- For cell/tissue homogenate, the final cell or tissue homogenate should be prepared in a buffer that has a neutral pH, contains minimal detergents or strong denaturing detergents, and has an ionic strength close to physiological concentration. Avoid debris, lipids, and cell/tissue aggregates. Centrifuge samples before use.
- Vortex all reagents well before adding to plate.

## Sample Collection and Storage

### Preparation of Serum Samples

- Allow the blood to clot for at least 30 minutes before centrifugation for 10 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq -20$  °C.
- Avoid multiple ( $> 2$ ) freeze/thaw cycles.
- When using frozen samples, it is recommended to thaw the samples completely, mix well by vortexing and centrifuge prior to use in the assay to remove particulates.
- It is recommended to use neat samples in the assay.

### Preparation of Plasma Samples

- Plasma collection using EDTA as an anti-coagulant is recommended. Centrifuge for 10 minutes at 1000 x g within 30 minutes of blood collection. Remove plasma and assay immediately or aliquot and store samples at  $\leq -20$  °C.
- Avoid multiple ( $> 2$ ) freeze/thaw cycles.
- When using frozen samples, it is recommended to thaw the samples completely, mix well by vortexing and centrifuge prior to use in the assay to remove particulates.
- It is recommended to use neat samples in the assay.

### Preparation of Tissue Culture Supernatant

- Centrifuge the sample to remove debris and assay immediately or aliquot and store samples at  $\leq -20$  °C.
- Avoid multiple ( $> 2$ ) freeze/thaw cycles.
- Tissue culture supernatant may require a dilution with an appropriate control medium prior to assay. Tissue/cell extracts should be done in neutral buffers containing reagents and conditions that do not interfere with assay performance. Excess concentrations of detergent, salt, denaturants, high or low pH, etc. will negatively affect the assay. Organic solvents should be avoided. The tissue/cell extract samples should be free of particles such as cells or tissue debris.

### NOTE:

- A maximum of 25  $\mu$ L per well of neat serum or plasma can be used. Tissue culture or other media may also be used.
- All samples must be stored in polypropylene tubes. **DO NOT STORE SAMPLES IN GLASS.**
- Avoid debris, lipids and cells when using samples with gross hemolysis or lipemia.
- Care must be taken when using heparin as an anticoagulant since an excess of heparin will provide falsely high values. Use no more than 10 IU heparin per mL of blood collected.

## Preparation of Reagents for Immunoassay

### Preparation of Antibody-Immobilized Beads

- If premixed beads are used, sonicate the premixed bead bottle 30 seconds and then vortex for 1 minute before use.
- For individual vials of beads, sonicate each antibody-bead vial for 30 seconds; vortex for 1 minute. Add 60  $\mu\text{L}$  from each antibody bead vial to the Mixing Bottle and bring final volume to 3.0 mL with Assay Buffer. Vortex the mixed beads well. Unused portion may be stored at 2-8  $^{\circ}\text{C}$  for up to one month. (**Note:** Due to the composition of magnetic beads, you may notice a slight color in the bead solution. This does not affect the performance of the beads or the kit.)

Example 1: When using 20 antibody-immobilized beads, add 60  $\mu\text{L}$  from each of the 20 bead vials to the Mixing Bottle. Then add 1.8 mL Assay Buffer.

Example 2: When using 10 antibody-immobilized beads, add 60  $\mu\text{L}$  from each of the 10 bead vials to the Mixing Bottle. Then add 2.4 mL Assay Buffer.

### Preparation of Quality Controls

Before use, reconstitute Quality Control 1 and Quality Control 2 with 250  $\mu\text{L}$  deionized water. Invert the vial several times to mix and vortex. Allow the vial to sit for 5-10 minutes and then transfer the controls to appropriately labeled polypropylene microfuge tubes. Unused portion may be stored at  $\leq 20$   $^{\circ}\text{C}$  for up to one month.

### Preparation of Wash Buffer

Bring the 10X Wash Buffer to room temperature and mix to bring all salts into solution. Dilute 60 mL of 10X Wash Buffer with 540 mL deionized water. Store unused portion at 2-8  $^{\circ}\text{C}$  for up to one month.

### Preparation of Serum Matrix

**This step is required for serum or plasma samples only.**

Add 1.0 mL deionized water to the bottle containing lyophilized Serum Matrix. Mix well. Allow at least 10 minutes for complete reconstitution. Leftover reconstituted Serum Matrix should be stored at  $\leq -20$   $^{\circ}\text{C}$  for up to one month.

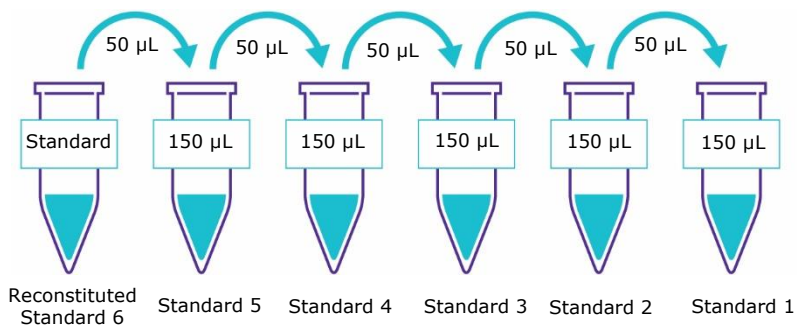
## Preparation of Mouse Th17 Standard

1. Prior to use, reconstitute the Mouse Th17 Standard with 250  $\mu\text{L}$  deionized water. Refer to table below for analyte concentrations. Invert the vial several times to mix. Vortex the vial for 10 seconds. Allow the vial to sit for 5-10 minutes and then transfer the standard to an appropriately labeled polypropylene microfuge tube. This will be used as standard 6; the unused portion may be stored at  $\leq -20$   $^{\circ}\text{C}$  for up to one month.
2. Preparation of Working Standards  
Label five polypropylene microfuge tubes as Standard 5, Standard 4, Standard 3, Standard 2 and Standard 1. Add 150  $\mu\text{L}$  of Assay Buffer to each of the five tubes. Prepare 1:4 serial dilutions by adding 50  $\mu\text{L}$  of the reconstituted Standard 6 to the Standard 5 tube, mix well and transfer 50  $\mu\text{L}$  of the Standard 5 to the Standard 4 tube, mix well and transfer 50  $\mu\text{L}$  of the Standard 4 to the Standard 3 tube, mix well and transfer 50  $\mu\text{L}$  of the Standard 3 to the Standard 2 tube, mix well and transfer 50  $\mu\text{L}$  of the Standard 2 to the Standard 1 tube. The 0 pg/mL Standard (Background) will be the Assay Buffer.

<b>Standard Number</b>	<b>Add Deionized Water (<math>\mu\text{L}</math>)</b>	<b>Add Standard (volume)</b>
Standard 6	250	0

<b>Standard Number</b>	<b>Add Assay Buffer (<math>\mu\text{L}</math>)</b>	<b>Add Standard (volume)</b>
Standard 5	150	50 $\mu\text{L}$ of Standard 6
Standard 4	150	50 $\mu\text{L}$ of Standard 5
Standard 3	150	50 $\mu\text{L}$ of Standard 4
Standard 2	150	50 $\mu\text{L}$ of Standard 3
Standard 1	150	50 $\mu\text{L}$ of Standard 2

## Preparation of Standards



<b>Standard</b>	<b>IL-17E (pg/mL)</b>	<b>GM-CSF &amp; IL-15 (pg/mL)</b>	<b>IFN<math>\gamma</math> &amp; IL-6 (pg/mL)</b>	<b>MIP-3<math>\alpha</math>, IL-31 &amp; CD40L (pg/mL)</b>	<b>IL-1<math>\beta</math> (pg/mL)</b>	<b>IL-2 (pg/mL)</b>
Standard 1	586	34	7.8	49	15	6.9
Standard 2	2344	137	31.3	195	59	23.4
Standard 3	9375	547	125	781	234	94
Standard 4	37500	2188	500	3125	938	375
Standard 5	150000	8750	2000	12500	3750	1500
Standard 6	600000	35000	8000	50000	15000	6000

<b>Standard</b>	<b>IL-4 (pg/mL)</b>	<b>IL-5 (pg/mL)</b>	<b>IL-21, IL-10 &amp; IL-12p70 (pg/mL)</b>	<b>IL-22 (pg/mL)</b>	<b>IL-28B (pg/mL)</b>	<b>IL-23 (pg/mL)</b>
Standard 1	1.5	4.9	20	2.4	127	342
Standard 2	5.9	19.5	78	9.8	508	1367
Standard 3	23	78	313	39	2031	5469
Standard 4	94	313	1250	156	8125	21875
Standard 5	375	1250	5000	625	32500	87500
Standard 6	1500	5000	20000	2500	130000	350000

<b>Standard</b>	<b>IL-27 (pg/mL)</b>	<b>IL-13 &amp; IL17A (pg/mL)</b>	<b>IL-17F (pg/mL)</b>	<b>IL-33 (pg/mL)</b>	<b>TNFβ (pg/mL)</b>	<b>TNFα (pg/mL)</b>
Standard 1	879	39	10	78	488	3.4
Standard 2	3516	156	39	313	1953	13.7
Standard 3	14063	625	156	1250	7813	55
Standard 4	56250	2500	625	5000	31250	219
Standard 5	225000	10000	2500	20000	125000	875
Standard 6	900000	40000	10000	80000	500000	3500

## Immunoassay Procedure

- Prior to beginning this assay, it is imperative to read this protocol completely and to thoroughly understand the Technical Guidelines.
  - Allow all reagents to warm to room temperature (20-25 °C) before use in the assay.
  - Diagram the placement of Standards [0 (Background), Standards 1 through 6], Controls 1 and 2, and samples on Well Map Worksheet in a vertical configuration.  
**(Note:** Most instruments will only read the 96-well plate vertically by default.) It is recommended to run the assay in duplicate.
  - If using a filter plate, set the filter plate on a plate holder at all times during reagent dispensing and incubation steps so that the bottom of the plate does not touch any surface.
1. Add 200  $\mu\text{L}$  of Wash Buffer into each well of the plate. Seal and mix on a plate shaker for 10 minutes at room temperature (20-25 °C).
  2. Decant Wash Buffer and remove the residual amount from all wells by inverting the plate and tapping it smartly onto absorbent towels several times.
  3. Add 25  $\mu\text{L}$  of each Standard or Control into the appropriate wells. Assay Buffer should be used for 0  $\text{pg/mL}$  standard (Background).
  4. Add 25  $\mu\text{L}$  of Assay Buffer to the sample wells.
  5. Add 25  $\mu\text{L}$  of appropriate matrix solution to the background, Standards, and Control wells. When assaying serum or plasma, use the Serum Matrix. When assaying tissue culture or other supernatant, use proper control culture medium as the matrix solution.
  6. Add 25  $\mu\text{L}$  of sample (neat) into the appropriate wells.
  7. Vortex Mixing Bottle and add 25  $\mu\text{L}$  of the Mixed or Premixed Beads to each well.  
**(Note:** During addition of Beads, shake bead bottle intermittently to avoid settling.)

Add 200  $\mu\text{L}$  Wash Buffer per well



Shake 10 min, RT  
Decant

- Add 25  $\mu\text{L}$  Standard or Control to appropriate wells
- Add 25  $\mu\text{L}$  Assay Buffer to background and sample wells
- Add 25  $\mu\text{L}$  appropriate matrix solution to background, Standards, and Control wells
- Add 25  $\mu\text{L}$  neat samples to sample wells
- Add 25  $\mu\text{L}$  Beads to each well



8. Seal the plate with a plate sealer. Wrap the plate with foil and incubate with agitation on a plate shaker overnight (16-18 hours) at 4 °C. Alternatively, incubate for 2 hours at room temperature (20-25 °C), but the overall performance, including sensitivity may be less than for the overnight incubation.
9. Gently remove well contents and wash plate two times following instructions listed in the Plate Washing section.
10. Add 25 µL of Detection Antibodies into each well.  
**(Note:** Allow the Detection Antibodies to warm to room temperature prior to addition.)
11. Seal, cover with foil and incubate with agitation on a plate shaker for 1 hr at room temperature (20-25 °C). **DO NOT ASPIRATE AFTER INCUBATION.**
12. Add 25 µL Streptavidin-Phycoerythrin to each well containing the 25 µL of Detection Antibodies.
13. Seal, cover with foil and incubate with agitation on a plate shaker for 30 minutes at room temperature (20-25 °C).
14. Gently Remove well contents and wash plate 2 times following instructions listed in the Plate Washing section.
15. Add 150 µL of Sheath Fluid PLUS (or Drive Fluid PLUS if using MAGPIX®) to all wells. Resuspend the beads in a plate shaker for 5 minutes.
16. Run plate on Luminex® 200™, HTS, FLEXMAP 3D®, MAGPIX® instrument with xPONENT® software or xMAP® INTELLIFLEX instrument with INTELLIFLEX software.
17. Save and analyze the Median Fluorescent Intensity (MFI) data using a 5-parameter logistic or spline-fitting method for calculating analyte concentrations in samples.  
**Note:** For diluted samples, multiply the calculated concentration by the dilution factor.



Incubate overnight at 4 °C or 2 hours at RT with shaking



Remove well contents and wash 2X with 200 µL Wash Buffer

Add 25 µL Detection Antibodies per well



Incubate 1 hr at RT (20-25°C)  
Do Not Aspirate

Add 25 µL Streptavidin-Phycoerythrin per well



Incubate for 30 minutes at RT  
Remove well contents and wash 2X with 200 µL Wash Buffer

Add 150 µL Sheath Fluid PLUS or Drive Fluid PLUS per well

Read on Luminex® 100 µL, 50 beads per bead set)

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## Plate Washing

If using a solid plate, use either a handheld magnet or magnetic plate washer.

### Solid Plate

- Handheld magnet (Cat. No. 40-285)  
Rest plate on magnet for 60 seconds to allow complete settling of magnetic beads. Remove well contents by gently decanting the plate in an appropriate waste receptacle and gently tapping on absorbent pads to remove residual liquid. Wash plate with 200  $\mu$ L of Wash Buffer by removing plate from magnet, adding Wash Buffer, shaking for 30 seconds, reattaching to magnet, letting beads settle for 60 seconds and removing well contents as previously described after each wash. Repeat wash steps as recommended in Assay Procedure.
- Magnetic plate washer (Cat. No. 40-094, 40-095, 40-096 and 40-097)  
Please refer to specific automatic plate washer manual for appropriate equipment settings. Please note that after the final aspiration, there will be approximately 25  $\mu$ L of residual wash buffer in each well. This is expected when using the BioTek<sup>®</sup> plate washer and this volume does not need to be aspirated from the plate.

If using an automatic plate washer other than BioTek<sup>®</sup> 405 LS or 405 TS, please refer to the manufacturer's recommendations for programming instructions.

### Filter Plate (Cat. No. MX-PLATE)

If using a filter plate, use a vacuum filtration manifold to remove well contents. Wash plate with 200  $\mu$ L/well of Wash Buffer, removing Wash Buffer by vacuum filtration after each wash. Repeat wash steps as recommended in the Assay Procedure.

## Equipment Settings

Luminex<sup>®</sup> 200™, HTS, FLEXMAP 3D<sup>®</sup>, MAGPIX<sup>®</sup> with xPONENT<sup>®</sup> software and xMAP<sup>®</sup> INTELLIFLEX with INTELLIFLEX software:

These specifications are for the above listed instruments and software. Luminex<sup>®</sup> instruments with other software (for example, MasterPlex<sup>®</sup>, StarStation, LiquiChip, Bio-Plex<sup>®</sup> Manager™, LABScan™100) would need to follow instrument instructions for gate settings and additional specifications from the vendors for reading Luminex<sup>®</sup> magnetic beads.

For magnetic bead assays, each instrument must be calibrated and performance verified with the indicated calibration and verification kits.

<b>Instrument</b>	<b>Calibration Kit</b>	<b>Verification Kit</b>
Luminex® 200™ and HTS	xPONENT® 3.1 compatible Calibration Kit (Cat. No. LX2R-CAL-K25)	Performance Verification Kit (Cat. No. LX2R-PVER-K25)
FLEXMAP 3D®	FLEXMAP 3D® Calibrator Kit (Cat. No. F3D-CAL-K25)	FLEXMAP 3D® Performance Verification Kit (Cat. No. F3D-PVER-K25)
xMAP® INTELLIFLEX	xMAP® INTELLIFLEX Calibration Kit (Cat. No. IFX-CAL-K20)	xMAP® INTELLIFLEX Performance Verification Kit (Cat. No. IFX-PVER-K20)
MAGPIX®	MAGPIX® Calibration Kit (Cat. No. MPX-CAL-K25)	MAGPIX® Performance Verification Kit (Cat. No. MPX-PVER-K25)

**NOTE:** When setting up a Protocol using the xPONENT® software, you must select MagPlex® as the Bead Type in the Acquisition settings.

**NOTE:** These assays cannot be run on any instruments using Luminex® IS 2.3 or Luminex® 1.7 software.

The Luminex® probe height must be adjusted to the plate provided in the kit. Please use Cat. No. MAG-PLATE, if additional plates are required for this purpose.

Events	50, per bead	
Sample Size	100 $\mu$ L	
Gate Settings	8,000 to 15,000	
Reporter Gain	Default (low PMT)	
Time Out	60 seconds	
Bead Set	Customizable 25-Plex Beads	
	IL-17E/IL-25	14
	GM-CSF	15
	IFN $\gamma$	19
	MIP-3 $\alpha$ /CCL20	20
	IL-1 $\beta$	25
	IL-2	26
	IL-4	28
	IL-5	30
	IL-6	34
	IL-21	35
	IL-22	37
	IL-28B	42
	IL-10	43
	IL-23	46
	IL-12p70	47
	IL-27	48
	IL-13	52
	IL-15	54
	IL-17A	56
	IL-17F	61
	IL-33	67
	IL-31	72
	TNF $\beta$	73
	TNF $\alpha$	77
	CD40L	78

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## Quality Controls

The ranges for each analyte in Quality Control 1 and 2 are provided on the card insert or can be located at our website [SigmaAldrich.com](http://SigmaAldrich.com) using the catalogue number as the keyword.

## Assay Characteristics

### Cross-Reactivity

There was no or negligible cross-reactivity between the antibodies for an analyte and the other analytes in the panel when MXMSM matrix is added to the standard curve. When cell culture media with 10% FBS is added to the standard curve the assay may have  $\leq 5\%$  cross-reactivity between the TNF $\beta$  standard and the IL-17E, IL-2, IL-27, IL13 and IL-31 assays.

## Assay Sensitivities (minimum detectable concentrations, pg/mL)

Minimum Detectable Concentration (MinDC) is calculated using MILLIPLEX® Analyst 5.1. It measures the true limits of detection for an assay by mathematically determining what the empirical MinDC would be if an infinite number of standard concentrations were run for the assay under the same conditions.

Analyte	Overnight Protocol (n=8)		2 Hour Protocol (n=3)	
	Min DC (pg/mL)	Min DC + 2 SD (pg/mL)	Min DC (pg/mL)	Min DC + 2 SD (pg/mL)
IL-17E/IL-25	357.3	475.3	425.7	530.6
GM-CSF	19.8	25.6	30.7	33.7
IFN $\gamma$	1.5	2.2	2.4	3.3
MIP-3 $\alpha$ /CCL20	9.4	11.8	12.7	13.8
IL-1 $\beta$	3.3	4.2	4.7	7.0
IL-2	1.5	2.2	1.0	1.3
IL-4	0.4	0.5	0.4	0.5
IL-5	5.1	8.3	4.6	5.1
IL-6	3.3	4.7	4.8	5.4
IL-21	35.8	19.8	35.8	19.8
IL-22	0.5	0.6	0.5	0.6
IL-28B	29.3	35.9	27.3	34.9
IL-10	5.6	7.5	5.7	6.8
IL-23	70.4	103.4	80.7	103.7
IL-12p70	12.9	17.3	13.7	14.8
IL-27	498.6	755.1	740.0	1071.6
IL-13	32.1	40.2	39.0	61.3
IL-15	8.6	11.0	12.0	17.3
IL-17A	18.6	28.9	33.7	38.3
IL-17F	5.2	6.4	9.6	10.9
IL-33	14.9	19.5	33.7	42.9
IL-31	25.8	33.9	51.7	63.4
TNF $\beta$	137.1	195.4	169.7	224.6
TNF $\alpha$	1.3	1.8	1.6	2.1
CD40L	14.6	19.4	45.0	62.1

## Precision

Intra-assay precision is generated from the mean of the %CV's from 8 reportable results across two different concentrations of analytes in a single assay. Inter-assay precision is generated from the mean of the %CV's across two different concentrations of analytes across 8 different assays.

### Overnight Protocol

<b>Analyte</b>	<b>Intra-assay %CV</b>	<b>Inter-assay %CV</b>
IL-17E/IL-25	3	8
GM-CSF	2	6
IFN $\gamma$	2	6
MIP-3 $\alpha$ /CCL20	3	7
IL-1 $\beta$	2	4
IL-2	3	3
IL-4	3	5
IL-5	2	6
IL-6	3	6
IL-21	3	7
IL-22	3	5
IL-28B	3	9
IL-10	3	5
IL-23	2	10
IL-12p70	2	3
IL-27	4	9
IL-13	4	7
IL-15	4	5
IL-17A	3	6
IL-17F	3	5
IL-33	4	5
IL-31	4	5
TNF $\beta$	4	12
TNF $\alpha$	3	5
CD40L	4	6

## Accuracy

Spike Recovery: The data represent mean percent recovery of spiked standards ranging from low, medium, and high concentration in serum matrices (n=6).

### Overnight Protocol

Analyte	% Recovery in Serum Matrix
IL-17E/IL-25	110
GM-CSF	105
IFN $\gamma$	97
MIP-3 $\alpha$ /CCL20	108
IL-1 $\beta$	103
IL-2	87
IL-4	96
IL-5	85
IL-6	100
IL-21	104
IL-22	101
IL-28B	106
IL-10	98
IL-23	101
IL-12p70	91
IL-27	101
IL-13	105
IL-15	102
IL-17A	92
IL-17F	102
IL-33	104
IL-31	99
TNF $\beta$	101
TNF $\alpha$	102
CD40L	103



## Troubleshooting

<b>Problem</b>	<b>Probable Cause</b>	<b>Solution</b>
Insufficient bead count	Plate washer aspirate height set too low	Adjust aspiration height according to manufacturers' instructions.
	Bead mix prepared inappropriately	Sonicate bead vials and vortex just prior to adding to bead mix bottle according to protocol. Agitate bead mix intermittently in reservoir while pipetting this into the plate.
	Samples cause interference due to particulate matter or viscosity	See above. Also sample probe may need to be cleaned with alcohol flushes, back flushes and washes; or if needed probe should be removed and sonicated.
	Probe height not adjusted correctly	When reading the assay on the Luminex® 200™ instrument, adjust probe height to the kit solid plate or to the recommended filter plates using 3 alignment discs. When reading the assay on the MAGPIX® instrument, adjust probe height to the kit solid plate or to the recommended filter plates using 2 alignment discs. When reading the assay on the FLEXMAP 3D® instrument, adjust probe height to the kit solid plate using 1 alignment disc. For the FLEXMAP 3D® instrument, when using the solid plate in the kit, the final resuspension should be with 150 µL Sheath Fluid PLUS in each well and 75 µL should be aspirated. When reading the assay on the xMAP® INTELLIFLEX instrument, adjust probe height based on the type of plate you are using, place an alignment disk or an alignment sphere in the well according to the protocol recommended by Luminex®.
Background is too high	Background wells were contaminated	Avoid cross-well contamination by using sealer appropriately and pipetting with multichannel pipettes without touching reagent in plate.
	Matrix used has endogenous analyte or interference	Check matrix ingredients for cross reacting components (for example, interleukin modified tissue culture medium).
	Insufficient washes	Increase number of washes.

<b>Problem</b>	<b>Probable Cause</b>	<b>Solution</b>
Beads not in region or gate	Luminex® instrument not calibrated correctly or recently	Calibrate Luminex® instrument based on manufacturer's instructions, at least once a week or if temperature has changed by > 3 °C.
	Gate settings not adjusted correctly	Some Luminex® instruments (for example, Bio-Plex®) require different gate settings than those described in the kit protocol. Use instrument default settings.
	Wrong bead regions in protocol template	Check kit protocol for correct bead regions or analyte selection.
	Incorrect sample type used	Samples containing organic solvents or if highly viscous should be diluted or dialyzed as required.
	Instrument not washed or primed	Prime the Luminex® 4 times to rid of air bubbles, wash 4 times with sheath fluid or water if there is any remnant alcohol or sanitizing liquid.
Signal for whole plate is same as background	Beads were exposed to light	Keep plate and bead mix covered with dark lid or aluminum foil during all incubation steps.
	Incorrect or no Detection Antibody was added	Add appropriate Detection Antibody and continue.
Low signal for standard curve	Streptavidin-Phycoerythrin was not added	Add Streptavidin-Phycoerythrin according to protocol. If Detection Antibody has already been removed, sensitivity may be low.
	Detection Antibody may have been removed prior to adding Streptavidin Phycoerythrin	May need to repeat assay if desired sensitivity not achieved.
Signals too high, standard curves are saturated	Incubations done at inappropriate temperatures, timings or agitation	Assay conditions need to be checked.
	Calibration target value set too high	With some Luminex® instruments (for example, Bio-Plex®) default target setting for RP1 calibrator is set at high PMT. Use low target value for calibration and reanalyze plate.
	Plate incubation was too long with standard curve and samples	Use shorter incubation time.

<b>Problem</b>	<b>Probable Cause</b>	<b>Solution</b>
Sample readings are out of range	Samples contain no or below detectable levels of analyte	If below detectable levels, it may be possible to use higher sample volume. Check with technical support for appropriate protocol modifications.
	Samples contain analyte concentrations higher than highest standard point	Samples may require dilution and reanalysis for just that particular analyte.
	Standard curve was saturated at higher end of curve	See above.
High variation in samples and/or standards	Multichannel pipette may not be calibrated	Calibrate pipettes.
	Plate washing was not uniform	Confirm all reagents are removed completely in all wash steps.
	Samples may have high particulate matter or other interfering substances	See above.
	Plate agitation was insufficient	Plate should be agitated during all incubation steps using a vertical plate shaker at a speed where beads are in constant motion without causing splashing.
	Cross-well contamination	Check when reusing plate sealer that no reagent has touched sealer. Care should be taken when using same pipette tips that are used for reagent additions and that pipette tip does not touch reagent in plate.

## FOR FILTER PLATES ONLY

<b>Problem</b>	<b>Probable Cause</b>	<b>Solution</b>
Filter plate will not vacuum	Vacuum pressure is insufficient	Increase vacuum pressure such that 0.2 mL buffer can be suctioned in 3-5 seconds.
	High lipid concentration	After centrifugation, remove lipid layer and use supernatant.
Plate leaked	Vacuum pressure too high	Adjust vacuum pressure such that 0.2 mL buffer can be suctioned in 3-5 seconds. May need to transfer contents to a new (blocked) plate and continue.
	Plate set directly on table or absorbent towels during incubations or reagent additions	Set plate on plate holder or raised edge so bottom of filter is not touching any surface.
	Insufficient blotting of filter plate bottom causing wicking	Blot the bottom of the filter plate well with absorbent towels after each wash step.
	Pipette touching plate filter during additions	Pipette to the side of plate.
	Probe height not adjusted correctly	Adjust probe to 3 alignment discs in well H6.
	Sample too viscous	May need to dilute sample.

## Product Ordering

<b>Replacement Reagents</b>	<b>Cat. No.</b>
Mouse Th17 Standard	MTH17-8047
Mouse Th17 Quality Controls 1 & 2	MTH17-6047
Serum Matrix	MXMSM
Mouse Th17 Detection Antibodies	MTH17-1047
Streptavidin-Phycoerythrin	L-SAPE3
Assay Buffer	L-AB
Set of two 96-Well plates with sealers	MAG-PLATE
10X Wash Buffer	L-WB
Mouse Th17 25 Plex Premixed Magnetic Bead Panel – BULK PACKAGED	MT17MAG47PMX25BK

### **Mouse Th17 Antibody-Immobilized Premixed Magnetic Beads**

Premixed 25-plex Beads	MTH17PMX25-MAG
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## Antibody-Immobilized Magnetic Beads

Analyte	Bead No.	Cat. No.
IL-17E/IL-25	14	MIL17E-MAG
GM-CSF	15	MGMCSF-MAG
IFN $\gamma$	19	MIFNG-MAG
MIP-3 $\alpha$ /CCL20	20	MTHMIP3A-MAG
IL-1 $\beta$	25	MIL1B-MAG
IL-2	26	MIL2-MAG
IL-4	28	MIL4-MAG
IL-5	30	MIL5-MAG
IL-6	34	MCYIL6-MAG
IL-21	35	MIL21-MAG
IL-22	37	MIL22-MAG
IL-28B	42	MIL28B-MAG
IL-10	43	MIL10-MAG
IL-23	46	MIL23-MAG
IL-12p70	47	MIL12P70-MAG
IL-27	48	MIL27-MAG
IL-13	52	MIL13-MAG
IL-15	54	MIL15-MAG
IL-17A	56	MIL17A-MAG
IL-17F	61	MIL17F-MAG
IL-33	67	MIL33-MAG
IL-31	72	MIL31-MAG
TNF $\beta$	73	MTNFB-MAG
TNF $\alpha$	77	MCYTNFA-MAG
CD40L	78	MCD40L-MAG

## Well Map

	1	2	3	4	5	6	7	8	9	10	11	12
A	Standard 0 (Background)	Standard 4	QC-2 Control									
B	Standard 0 (Background)	Standard 4	QC-2 Control									
C	Standard 1	Standard 5	Sample 1									
D	Standard 1	Standard 5	Sample 1									
E	Standard 2	Standard 6	Sample 2									
F	Standard 2	Standard 6	Sample 2									
G	Standard 3	QC-1 Control	Etc.									
H	Standard 3	QC-1 Control										

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