

Product Information

ADME/Tox Cell Lines

Caco-2 Engineered Control Cells 24 Well Assay Ready Plate

Catalog Number **MTOX1000CC24**
Store at Room Temperature

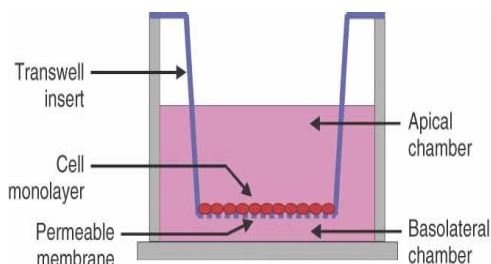
TECHNICAL BULLETIN

Product Description

ATP-binding cassette (ABC) transporters are a family of transmembrane proteins that utilize ATP hydrolysis for translocation of substrates across membranes. ABC transporters are known to play a critical role in the development of multidrug resistance. Evaluation of membrane transporter pharmacology in drug disposition and drug-drug interactions (DDI) is critical to the pharmaceutical safety evaluations of new drug entities. Selection of the targeted gene(s) was based on the considerable body of evidence supporting its crucial role in the development of multidrug resistance.¹

This product contains engineered Caco-2 cells (C2BBE1 sub-clone) that have been differentiated for 14 days in a Multiwell Insert – 24 Well Assay Ready Plate (see Figure 1). At day 14 the plate is prepared in an exclusive and proprietary shipping medium that is stable at room temperature and allows for up to 4 days of shipping. The plate is shipped overnight for next day delivery.

Figure 1.
Transwell of Multiwell Insert – 24 Well Assay Ready Plate



Components

This kit contains one Multiwell Insert – 24 Well Assay Ready Plate:

Engineered Caco-2 control cells (C2BBE1 sub-clone)
(Cat. No. MTOX1000CC24)

Note: Receiver tray not included and must be purchased separately for each Assay Ready Plate. [Falcon® 24-well plate: 351181 and receiver: 353047]

Cell Line Description

Parental Cell Line: ATCC® Cat. No. CRL-2102™

Note: Please see CRL-2102 product datasheet from ATCC for additional information about the origin of these cell lines. Cytogenetic information is based on initial seed stock at Sigma-Aldrich Life Science. Cytogenetic instability has been reported in the literature for some cell lines.

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Preparation Instructions

Caco-2 Medium: Fetal bovine serum, Catalog No. F4135, at a final concentration of 10% (v/v) in DMEM, Catalog No. D5671, supplemented with L-glutamine, Catalog No. G7513, to a final concentration of 2 mM and penicillin-streptomycin, Catalog No. P4333, at a final concentration of 1% (v/v). This medium is formulated for use with a 5% CO₂ in air atmosphere.

Procedures

Notes: Receiver tray not included and must be purchased separately for each Assay Ready Plate.

The shipping medium must be changed on the day the plate is received. The unpacking instructions should be followed for each plate that is received.

Unpacking:

Upon receipt, open the box and remove the plastic Ziploc® bag containing the Multiwell Insert – 24 well Assay Ready Plate.

Changing the Shipping Medium

1. On day of receipt, prepare everything needed to replace the shipping medium with fresh Caco-2 cell culture medium:
 - cell culture biosafety cabinet
 - standard Caco-2 medium, pre-warmed to 37 °C aspiration system
 - 24 well receiver tray (one for each plate received)
 - sterile containers for culture medium
2. Remove the Multiwell Insert – 24 well Assay Ready Plate (still at room temperature) from the Ziploc bag.
3. In a biosafety cabinet, unwrap the Multiwell Insert – 24 well Assay Ready Plate and carefully pull off the Parafilm®.
4. Open the Assay Ready Plate, placing the lid next to the plate, facing upwards. Carefully remove the ThermalSeal® film.
5. Add 400 µl of culture medium in each apical well.
6. Place the lid back on the 24 well Assay Ready Plate and place in the cell culture incubator for a minimum of 4 hours to allow the transport medium to liquefy.
7. Prepare the receiver tray for changing the medium:
 - a. Ensure the Caco-2 medium has been warmed to 37 °C.
 - b. In biosafety cabinet, unwrap one 24 well receiver tray (must be ordered separately) for each Assay Ready Plate. Place lid next to the plate, facing upwards.
 - c. Add 800 µl of warm Caco-2 medium in each basal well.
 - d. Place the lid on the plate and place the plate in the incubator.

Notes: Once the shipping medium has liquefied, proceed to step 8. The following must be performed using sterile technique in the biosafety cabinet.

Never handle more than one plate at a time while changing the shipping medium.

8. Take one Assay Ready Plate and one receiver tray out of the incubator and place both in biosafety cabinet.
9. Open the Assay Ready Plate, placing the lid next to the plate, facing upwards.
10. Gently lift the apical (upper) section of the Assay Ready Plate and place it on the new receiver tray.
11. Cover the receiver tray with its lid, then put it back into the cell culture incubator.
12. Repeat steps 1–11 for additional plates.

Notes: After the shipping medium has been changed to fresh Caco-2 medium, the plate should be kept in the incubator until day 21. Transwell assays can be performed on days 21–25.

Culture medium should be replaced every 48–72 hours.

TEER Measurement:

Read instructions for proper use of the TEER instrument in addition to these instructions.

1. Sterilize the electrodes (probe): submerge both electrodes in 70% ethanol for 30 minutes.
2. Equilibrate the electrodes (probe) for 30 minutes in Caco-2 Medium.
3. Insert the probe in the Transwell system so the shorter electrode is slightly submerged inside the culture medium of the apical well and the longer arm is placed through the lateral hole of the Transwell, so it is submerged in the medium of the basal well.
4. A TEER value of ≥ 240 ohms \cdot cm² is acceptable.
Note: It may be necessary to adjust X,Y coordinates on the TEER instrument for specific tissue culture plates.

Figure 2.**Representative TEER data**

Day 14	Ohms (Raw Data)					
	1215	1203	1198	1167	1052	1135
	1258	1256	1192	1163	1121	1153
	1252	1193	1211	1221	1130	1228
	1418	1228	1227	1164	1172	1285
Day 14	Ohms \cdot cm ² (Calculated Data)					
	851	842	839	817	736	794
	881	879	834	814	785	807
	876	835	848	855	791	859
	993	859	859	815	820	900
Day 21	Ohms (Raw Data)					
	918	943	922	915	746	724
	872	882	807	771	705	711
	867	827	754	557	802	829
	763	678	611	730	620	958
Day 21	Ohms \cdot cm ² (Calculated Data)					
	643	660	646	641	522	507
	610	618	565	539	494	497
	607	579	528	390	561	580
	534	475	427	511	434	671

Transwell Assay:

This protocol is designed to assess drug transporter functionality in Caco-2 cells. Transport is measured in both directions (apical-to-basal and basal-to-apical) across the cell monolayer, enabling an efflux ratio to be determined. In this study, buffer is taken from the receiver compartment after a designated time point. Compound concentrations in the receiver samples are quantified by LC-MS/MS, and the apparent permeability coefficient (P_{app}) and efflux ratio of the compound across the monolayer are calculated.

1. Materials

- Assay Ready Plates: Caco-2 engineered control cell type plates
- Caco-2 Medium
- Buffer B (see Reagent Preparation)
- Test compound working solution (see Reagent Preparation)
- Sample analysis equipment (fluorimeter, HPLC-UV/MS, liquid scintillation counter, etc)

2. Reagent Preparation

Use ultrapure water or equivalent to prepare reagents and in protocol steps.

- Buffer B - 500 ml of HBSS containing:
 - 12.5 ml of 1 M D-glucose
 - 10 ml of 1 M HEPES buffer
 - 1 ml of 625 mM CaCl₂
 - 1 ml of 250 mM MgCl₂
 - Adjust to pH 7.4
 - Store up to 4 weeks at 2–8 °C
- Test Compound Stock Solution: Dissolve compound at 200 \times concentration in DMSO and vortex to mix. If necessary, warm or sonicate to dissolve completely. Store up to 6 months at 2–8 °C.
- Test Compound Working Solution: Dilute Test Compound Stock Solution 200-fold with HBSS to make a working solution with a final DMSO concentration of 0.5% (v/v). Prepare fresh just before use.

3. Perform Transwell Assay
 - a. Aspirate medium from the apical and basal wells and replace with Buffer B (500 μ l in the apical wells and 1,000 μ l in the basal wells). Incubate at 37 °C for 15 minutes.
 - b. Aspirate all of Buffer B. Depending on the study design, add Test Compound Working Solution to the apical (500 μ l) or basal (1,000 μ l) wells, and add Buffer B to the other (basal or apical) wells. Incubate at 37 °C for 2 hours.
 - c. Take 250 μ l samples from the appropriate wells, depending on the direction of transport (i.e., from the basal well for A-to-B transport or the apical well for B-to-A transport).
 - d. Analyze samples.
 - e. Following quantitation of test compound, proceed to determination of (P_{app}) value and efflux ratio.

4. Determine P_{app} value and efflux ratio
 - a. Calculate the permeability coefficient as follows:

$$P_{app} = \frac{1}{A \times C_o} \times \frac{dM_r}{dt}$$

A = area (cm²)

C_o = mass of compound initially in the donor compartment

dM_r/dt = the rate of drug permeation across the cells

- b. Calculate the efflux ratio (ER) as the ratio of P_{app} determined in the A-to-B direction to P_{app} determined in the B-to-A direction:

$$ER = P_{app, B-to-A} / P_{app, A-to-B}$$

Measurement of Cell Monolayer Integrity using Lucifer Yellow

Evaluation of permeability characteristics of Caco-2 cells can be performed by measuring passive passage of different molecules across the monolayer. Small hydrophilic compounds cross the monolayer mainly via the paracellular space, such as through the tight junctions, and can be considered markers of passage by this route. Lucifer Yellow is one such marker that is easily detectable. It is used to check the barrier integrity and to determine whether the working concentration of a test compound disturbs the integrity of the monolayer. In this protocol, the Lucifer Yellow assay is performed after the Transwell assay.

1. Materials
 - Transwell assay plates
 - Buffer B
 - 0.1 mg/ml Lucifer Yellow Solution - (Lucifer Yellow CH dipotassium salt, Cat. No. L0144) in Buffer B
 - 96 well plate
 - Fluorescence multiwell plate reader
2. Perform Lucifer Yellow Assay
 - a. After removing samples for sample analysis, aspirate the remaining liquid from the apical and basal wells.
 - b. Add 500 μ l of 0.1 mg/ml Lucifer Yellow Solution to the apical wells and 1,000 μ l of Buffer B to the basal wells.
 - c. Incubate at 37 °C for 60 minutes.
 - d. Transfer 150 μ l from the basal wells to a 96 well plate and read in a spectrofluorometer with excitation at 485 nm and emission at 535 nm. Also measure fluorescence for Buffer B (blank) and 0.1 mg/ml Lucifer Yellow Solution.
3. Calculate the percent permeability from the fluorescence values as follows:

$$\% \text{ permeability} = \frac{\text{sample} - \text{blank}}{\text{Lucifer Yellow} - \text{blank}} \times 100$$

A permeability of <3% is acceptable.

Figure 3.
Representative Lucifer Yellow Data

0.037	0.017	0.013	0.009	0.014	0.056	Day 21
0.017	0.009	0.012	0.024	0.011	4.204	
0.009	0.016	0.008	0.009	0.018	0.012	
0.201	0.154	0.158	1.020	1.092	0.996	
0.021	0.023	0.029	0.027	0.029	0.027	Day 23
0.039	0.021	0.018	0.021	0.030	0.037	
0.016	0.038	0.046	0.029	0.016	0.036	
0.130	0.139	0.110	1.345	1.377	1.394	
0.238	0.094	0.074	0.094	0.106	0.183	Day 25
0.056	0.055	0.043	0.050	0.066	0.110	
0.250	0.117	0.096	0.058	0.044	0.066	
0.099	0.189	0.089	0.961	0.847	0.789	

References

1. The International Transporter Consortium (2010 White Paper), Membrane transporters in drug development. Nature Reviews Drug Discovery, **9**, 215-236 (2010).

Additional product and technical information can be obtained by searching for the catalog number at the following web page (www.sigmaaldrich.com).

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